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(71) Applicant: **DOW AGROSCIENCES LLC [US/US]; 9330 Zionsville Road, Indianapolis, IN 46268 (US).**

(72) Inventors: **PETELL, James, K.; 16825 Meyer Lane, Grass Valley, CA 95949 (US). MERLO, Donald, J.; 11845 Durbin Drive, Carmel, IN 46032 (US). HERMAN, Rod, A.; 11153 West 500 South, New Ross, IN 47968 (US). ROBERTS, Jean, L.; 26035 State Road 19, Arcadia, IN 46030 (US). GUO, Lining; 3212 Summit Ridge Loop, Morrisville, NC 27560 (US). SCHAFER, Barry, W.; 1429 Lighthouse Point, Cicero, IN 46034 (US). SUKHAPINDA, Kitisri; 4748 Ashwood Court, Zionsville, IN 46077 (US). MERLO, Ann, Owens; 11845 Durbin Drive, Carmel, IN 46032 (US).**

(74) Agent: **STUART, Donald, R.; Dow AgroSciences LLC, 9330 Zionsville Road, Indianapolis, IN 46268 (US).**

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(54) Title: **TRANSGENIC PLANTS EXPRESSING PHOTORHABDUS TOXIN**

(57) Abstract: Novel polynucleotide sequences that encode insect toxins TcdA and TcbA have base compositions that differ substantially from the native genes, making them more similar to plant genes. The new sequences are suitable for use for high expression in both monocots and dicots. Transgenic plants with a genome comprising a nucleic acid of SEQ ID NO: 3 or SEQ ID NO:4 are insect resistant.

TRANSGENIC PLANTS EXPRESSING PHOTORHABDUS TOXIN

## BACKGROUND OF THE INVENTION

As reported in WO98/08932, protein toxins from the genus *Photorhabdus* have been shown to have oral toxicity

5 against insects. The toxin complex produced by *Photorhabdus luminescens* (W-14), for example, has been shown to contain ten to fourteen proteins, and it is known that these are produced by expression of genes from four distinct genomic regions: *tca*, *tcb*, *tcc*, and *tcd*.

10 WO98/08932 discloses nucleotide sequences for the native toxin genes.

Of the separate toxins isolated from *Photorhabdus luminescens* (W-14), those designated Toxin A and Toxin B are especially potent against target insect species of

15 interest, for example corn rootworm. Toxin A is comprised of two different subunits. The native gene *tcdA* (SEQ ID NO:1) encodes protoxin *TcdA* (see SEQ ID NO:1). As determined by mass spectrometry, *TcdA* is processed by one or more proteases to provide Toxin A.

20 More specifically, *TcdA* is an approximately 282.9 kDa protein (2516 aa) that is processed to provide *TcdAii*, an approximately 208.2 kDa (1849 aa) protein encoded by nucleotides 265-5811 of SEQ ID NO:1, and *TcdAiii*, an approximately 63.5 kDa (579 aa) protein encoded by

25 nucleotides 5812-7551 of SEQ ID NO:1.

Toxin B is similarly comprised of two different subunits. The native gene *tcbA* (SEQ ID NO:2) encodes protoxin *TcbA* (see SEQ ID NO:2). As determined by mass spectrometry, *TcbA* is processed by one or more proteases

30 to provide Toxin B. More specifically, *TcbA* is an approximately 280.6 kDa (2504 aa) protein that is processed to provide *TcbAii*, an approximately 207.7 kDa (1844 aa) protein encoded by nucleotides 262-5793 of SEQ ID NO:2 and *TcbAiii*, an approximately 62.9 kDa (573 aa) protein encoded by nucleotides 5794-7512 of SEQ ID NO:2.

The native *tcdA* and *tcbA* genes are not well suited for high level expression in plants. They encode multiple destabilization sequences, mRNA splice sites, polyA addition sites and other possibly detrimental sequence motifs. In addition, the codon compositions are not like those of plant genes. WO98/08932 gives general guidance on how the toxin genes could be reengineered to more efficiently expressed in the cytoplasm of plants, and describes how plants can be transformed to incorporate the *Photorhabdus* toxin genes into their genomes.

#### SUMMARY OF THE INVENTION

In a preferred embodiment, the invention provides novel polynucleotide sequences that encode *TcdA* and *TcbA*. The novel sequences have base compositions that differ substantially from the native genes, making them more similar to plant genes. The new sequences are suitable for use for high expression in both monocots and dicots, and this feature is designated by referring to the sequences as the "hemicot" criteria, which is set forth in detail hereinafter. Other important features of the sequences are that potentially deleterious sequences have been eliminated, and unique restriction sites have been built in to enable adding or changing expression elements, organellar targeting signals, engineered protease sites and the like, if desired.

In a particularly preferred embodiment, the invention provides polynucleotide sequences that satisfy hemicot criteria and that comprise a sequence encoding an endoplasmic reticulum signal or similar targeting sequence for a cellular organelle in combination with a sequence encoding *TcdA* or *TdbA*.

More broadly, the invention provides engineered nucleic acids encoding functional *Photorhabdus* toxins wherein the sequences satisfy hemicot criteria.

The invention also provides transgenic plants with genomes comprising a novel sequence of the invention that imparts functional activity against insects.

5 BRIEF DESCRIPTION OF SEQUENCES

SEQ ID NO:1 is the native *tcdA* DNA sequence together with the corresponding encoded amino acid sequence for *TcdA*.

10 SEQ ID NO:2 is the native *tcbA* DNA sequence together with the corresponding encoded amino acid sequence for *TcbA*.

SEQ ID NO:3 is an artificial sequence encoding *TcdA* that is suitable for expression in monocot and dicot plants.

15 SEQ ID NO:4 is an artificial sequence encoding *TdbA* that is suitable for expression in monocot and dicot plants.

SEQ ID NO:5 is an artificial hemicot sequence that encodes the 21 amino acid ER signal peptide of 15 kDa 20 zein from Black Mexican Sweet maize.

SEQ ID NO:6 is an artificial hemicot sequence that encodes for the full-length native *TcdA* protein (amino acids 22-2537) fused to the modified 15 kDa zein endoplasmic reticulum signal peptide (amino acids 1-21).

25 DETAILED DESCRIPTION

The native *Photorhabdus* toxins are protein complexes that are produced and secreted by growing bacteria cells of the genus *Photorhabdus*. Of particular interest are the proteins produced by the species *Photorhabdus luminescens*. The protein complexes have a molecular size of approximately 1,000 kDa and can be separated by SDS-PAGE gel analysis into numerous component proteins. The toxins contain no hemolysin, lipase, type C phospholipase, or nuclease activities. The toxins 30 exhibit significant toxicity upon ingestion by a number of insects.

A unique feature of *Photorhabdus* is its bioluminescence. *Photorhabdus* may be isolated from a variety of sources. One such source is nematodes, more particularly nematodes of the genus *Heterorhabditis*.

5 Another such source is from human clinical samples from wounds, see Farmer et al. 1989 *J. Clin. Microbiol.* 27 pp. 1594-1600. These saprophytic strains are deposited in the American Type Culture Collection (Rockville, MD) ATCC #s 43948, 43949, 43950, 43951, and 43952, and are  
10 incorporated herein by reference. It is possible that other sources could harbor *Photorhabdus* bacteria that produce insecticidal toxins. Such sources in the environment could be either terrestrial or aquatic based.

The genus *Photorhabdus* is taxonomically defined as a  
15 member of the Family *Enterobacteriaceae*, although it has certain traits atypical of this family. For example, strains of this genus are nitrate reduction negative, yellow and red pigment producing and bioluminescent.

This latter trait is otherwise unknown within the  
20 *Enterobacteriaceae*. *Photorhabdus* has only recently been described as a genus separate from the *Xenorhabdus* (Boemare et al., 1993 *Int. J. Syst. Bacteriol.* 43, 249-255). This differentiation is based on DNA-DNA hybridization studies, phenotypic differences (e.g.,  
25 presence (*Photorhabdus*) or absence (*Xenorhabdus*) of catalase and bioluminescence) and the Family of the nematode host (*Xenorhabdus*; *Steinernematidae*, *Photorhabdus*; *Heterorhabditidae*). Comparative, cellular fatty-acid analyses (Janse et al. 1990, *Lett. Appl. Microbiol.* 10, 131-135; Suzuki et al. 1990, *J. Gen. Appl. Microbiol.*, 36, 393-401) support the separation of *Photorhabdus* from *Xenorhabdus*.

Currently, the bacterial genus *Photorhabdus* is comprised of a single defined species, *Photorhabdus*  
35 *luminescens* (ATCC Type strain #29999, Poinar et al., 1977, *Nematologica* 23, 97-102). A variety of related

strains have been described in the literature (e.g.,  
Akhurst et al. 1988 J. Gen. Microbiol., 134, 1835-1845;  
Boemare et al. 1993 Int. J. Syst. Bacteriol. 43 pp. 249-  
255; Putz et al. 1990, Appl. Environ. Microbiol., 56,  
5 181-186).

The following toxin producing *Photorhabdus* strains  
have been deposited:

strain	accession number	date of deposit
W-14	ATCC 55397	March 5, 1993
WX1	NRRL B-21710	April 29, 1997
WX2	NRRL B-21711	April 29, 1997
WX3	NRRL B-21712	April 29, 1997
WX4	NRRL B-21713	April 29, 1997
WX5	NRRL B-21714	April 29, 1997
WX6	NRRL B-21715	April 29, 1997
WX7	NRRL B-21716	April 29, 1997
WX8	NRRL B-21717	April 29, 1997
WX9	NRRL B-21718	April 29, 1997
WX10	NRRL B-21719	April 29, 1997
WX11	NRRL B-21720	April 29, 1997
WX12	NRRL B-21721	April 29, 1997
WX14	NRRL B-21722	April 29, 1997
WX15	NRRL B-21723	April 29, 1997
H9	NRRL B-21727	April 29, 1997
Hb	NRRL B-21726	April 29, 1997
Hm	NRRL B-21725	April 29, 1997
HP88	NRRL B-21724	April 29, 1997
NC-1	NRRL B-21728	April 29, 1997
W30	NRRL B-21729	April 29, 1997
WIR	NRRL B-21730	April 29, 1997
B2	NRRL B-21731	April 29, 1997
ATCC 43948	ATCC 55878	November 5, 1996
ATCC 43949	ATCC 55879	November 5, 1996
ATCC 43950	ATCC 55880	November 5, 1996
ATCC 53951	ATCC 55881	November 5, 1996
ATCC 43952	ATCC 55882	November 5, 1996
DEPI	NRRL B-21707	April 29, 1997
DEP2	NRRL B-21708	April 29, 1997
DEP3	NRRL B-21709	April 29, 1997
P. zealandica	NRRL B-21683	April 29, 1997
P. hepialus	NRRL B-21684	April 29, 1997
HB-Arg	NRRL B-21685	April 29, 1997
HB Oswego	NRRL B-21686	April 29, 1997
Hb Lewiston	NRRL B-21687	April 29, 1997
K-122	NRRL B-21688	April 29, 1997
HMGD	NRRL B-21689	April 29, 1997
Indicus	NRRL B-21690	April 29, 1997
GD	NRRL B-21691	April 29, 1997
PWH-5	NRRL B-21692	April 29, 1997
Megidis	NRRL B-21693	April 29, 1997
HF-85	NRRL B-21694	April 29, 1997
A. Cows	NRRL B-21695	April 29, 1997
MP1	NRRL B-21696	April 29, 1997
MP2	NRRL B-21697	April 29, 1997
MP3	NRRL B-21698	April 29, 1997
MP4	NRRL B-21699	April 29, 1997
MP5	NRRL B-21700	April 29, 1997
GL98	NRRL B-21701	April 29, 1997
GL101	NRRL B-21702	April 29, 1997
GL138	NRRL B-21703	April 29, 1997
GL155	NRRL B-21704	April 29, 1997
GL217	NRRL B-21705	April 29, 1997
GL257	NRRL B-21706	April 29, 1997

All strains were deposited in accordance with the terms of the Budapest Treaty. Strains having

accession numbers prefaced by "ATTC" were deposited on the indicated date in the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852 USA. Strains prefaced by "NRRL" were 5 deposited on the indicated date in the Agricultural Research Service Patent Culture Collection (NRRL), National Center for Agricultural Utilization Research, ARS-USDA, 1815 North University St., Peoria IL 61604 USA.

10 The present invention provides hemicot nucleic acid sequences encoding toxins from any *Photorhabdus* species or strain that produces a toxin having functional activity. Hemicot nucleic acid sequences encoding proteins homologous to such toxins are also encompassed 15 by the invention.

Several terms that are used herein have a particular meaning and are defined as follows:

By "functional activity" it is meant herein that the protein toxins) function as insect control agents in that 20 the proteins are orally active, or have a toxic effect, or are able to disrupt or deter feeding, which may or may not cause death of the insect. When an insect comes into contact with an effective amount of toxin delivered via transgenic plant expression, formulated protein 25 compositions), sprayable protein compositions), a bait matrix or other delivery system, the results are typically death of the insect, or the insects do not feed upon the source which makes the toxins available to the insects.

30 By "homolog" it is meant an amino acid sequence that is identified as possessing homology to a reference *Photorhabdus* toxin polypeptide amino acid sequence.

By "homology" it is meant an amino acid sequence that has a similarity index of at least 33% and/or an 35 identity index of at least 26% to a reference *Photorhabdus* toxin polypeptide amino acid sequence, as

scored by the GAP algorithm using the B10sum 62 protein scoring matrix Wisconsin Package Version 9.0, Genetics Computer Group GCG), Madison, WI).

By "identity" is meant an amino acid sequence that 5 contains an identical residue at a given position, following alignment with a reference *Photorhabdus* toxin polypeptide amino acid sequence by the GAP algorithm.

By the use of the term "Photorhabdus toxin" it is meant any protein produced by a *Photorhabdus* 10 microorganism strain which has functional activity against insects, where the *Photorhabdus* toxin could be formulated as a sprayable composition, expressed by a transgenic plant, formulated as a bait matrix, delivered via baculovirus, or delivered by any other applicable 15 host or delivery system.

By the use of the term "toxic" or "toxicity" as used herein it is meant that the toxins produced by *Photorhabdus* have "functional activity" as defined herein.

20 By "substantial sequence homology" is meant either: a DNA fragment having a nucleotide sequence sufficiently similar to another DNA fragment to produce a protein having similar biochemical properties; or a polypeptide having an amino acid sequence sufficiently similar to 25 another polypeptide to exhibit similar biochemical properties.

As with other bacterial toxins, the rate of mutation of the bacteria in a population causes many related toxins slightly different in sequence to exist. Toxins 30 of interest here are those which produce protein complexes toxic to a variety of insects upon exposure, as described herein. Preferably, the toxins are active against *Lepidoptera*, *Coleoptera*, *Homoptera*, *Diptera*, *Hymenoptera*, *Dictyoptera* and *Acarina*. The inventions 35 herein are intended to capture the protein toxins homologous to protein toxins produced by the strains

herein and any derivative strains thereof, as well as any protein toxins produced by *Photorhabdus*. These homologous proteins may differ in sequence, but do not differ in function from those toxins described herein. 5 Homologous toxins are meant to include protein complexes of between 300 kDa to 2,000 kDa and are comprised herein, at least two 2) subunits, where a subunit is a peptide which may or may not be the same as the other subunit. Various protein subunits have been identified and are taught in 10 the Examples herein. Typically, the protein subunits are between about 18 kDa to about 230 kDa; between about 160 kDa to about 230 kDa; and about 50 kDa to about 80 kDa. 15 As discussed above, some *Photorhabdus* strains can be isolated from nematodes. Some nematodes, elongated cylindrical parasitic worms of the phylum Nematoda, have evolved an ability to exploit insect larvae as a favored growth environment. The insect larvae provide a source of food for growing nematodes. One dramatic effect that follows invasion 20 to reproduce. Death results from the presence of, in certain nematodes, larval growth and inhibits feeding activity. 25 Interestingly, it appears that each genus of insect parasitic nematode hosts a particular species of bacterium, uniquely adapted for symbiotic growth with that nematode. In the interim since this research was initiated, the name of the bacterial genus *Photorhabdus* was reclassified into the genus *Xenorhabdus* and the 30 *Photorhabdus*. Bacteria of the genus *Photorhabdus* are characterized as being symbionts of *Heterorhabditus* nematodes while *Xenorhabdus* species are symbionts of the *Steinerinema* species. This change in nomenclature is reflected in this specification, but in no way should a 35

change in nomenclature alter the scope of the inventions described herein.

The peptides and genes that are disclosed herein are named according to the guidelines recently published in 5 the Journal of Bacteriology "Instructions to Authors" p. i-xii Jan. 1996), which is incorporated herein by reference.

Transformation methods useful in carrying out the invention are well known, and are described, for example, 10 in WO98/08932.

Hemicot tcdA and tcbA

SEQ ID NO: 3 is the nucleotide sequence for an engineered tcdA gene in accordance with the invention.

SEQ ID NO: 4 is the nucleotide sequence for an engineered 15 tcbA gene in accordance with the invention.

The following Tables 1 and 2 identify significant features of the engineered tcdA and tcbA genes.

Table 1  
tcdA

Feature	nucleotides of SEQ ID NO:3
<i>NcoI</i>	1-6
<i>HindIII</i>	48-53
<i>KpnI</i>	246-254
sequence encoding	267-5798
<i>TcbAii</i>	
<i>NheI</i>	333-338
<i>BglII</i>	1215-1220
<i>Clal</i>	2604-2609
<i>PstI</i>	4015-4020
<i>AgeI</i>	5088-5093
<i>MunI</i>	5598-5603
<i>XbaI</i>	5778-5783
sequence encoding	5799-7517
<i>TcbAiii</i>	
<i>AflIII</i>	5853-5858
<i>SphI</i>	6439-6444
<i>SfuI</i>	7392-7397
<i>SacI</i>	7519-7524
<i>XhoI</i>	7522-7527
<i>StuI</i>	7528-7533
<i>NotI</i>	7533-7538

20

Table 2  
tcbA

Feature	nucleotides of SEQ ID NO:5
<i>NcoI</i>	1-6
<i>HindIII</i>	48-53

<i>KpnI</i>	246-251
sequence encoding <i>TcbAii</i>	267-5798
<i>NheI</i>	333-338
<i>BglIII</i>	1215-1220
<i>ClaI</i>	2604-2609
<i>PstI</i>	4015-4020
<i>AgeI</i>	5088-5093
<i>MunI</i>	5598-5603
<i>XbaI</i>	5778-5783
sequence encoding <i>TcbAiii</i>	5799-7517
<i>AflIII</i>	5853-5858
<i>SphI</i>	6439-6444
<i>SfuI</i>	7392-7397
<i>SacI</i>	7519-7524
<i>SfuI</i>	7392-7397
<i>SacI</i>	7519-7524
<i>XbaI</i>	7522-7527
<i>StuI</i>	7528-7533
<i>NotI</i>	7535-7540

It should be noted that the proteins encoded by the plant-optimized *tcdA* (SEQ ID NO:3) and *tcbA* (SEQ ID NO:5) differ from the native proteins by the addition of 5 an Ala residue at position #2. This modification was made to accommodate the *NcoI* site which spans the ATG start codon.

10 The following Table 3 compares the codon composition of the engineered *tcdA* gene of SEQ ID NO:3 and engineered *tcbA* gene of SEQ ID NO:5 with the codon compositions of the native genes, the typical dicot genes, and maize genes.

Table 3

amino acid	codon	% in SEQ ID NO:3	% in <i>tcdA</i>	% in SEQ ID NO:5	% in <i>tcbA</i>	% in dicot	% in maize
Ala	GCT	62	21	69	41	42	24
	GCC	26	32	27	17	27	34
	GCA	11	25	4	22	25	18
	GCG	0	21	0	21	6	24
Arg	AGG	48	0	60	2	25	26
	CGC	22	36	18	16	11	24
	AGA	20	11	15	6	30	15
	CGT	11	39	7	57	21	11
	CGG	0	7	0	13	4	15
	CGA	0	8	0	6	8	9
Asn	AAC	100	32	100	33	55	68
	AAT	0	68	0	67	45	32
Asp	GAC	67	22	70	25	42	63

amino acid	codon	% in SEQ ID NO:3	% in tcdA	% in SEQ ID NO:5	% in tcbA	% in dicot	% in maize
	GAT	33	78	30	75	58	37
Cys	TGC	100	30	100	19	56	68
	TGT	0	70	0	81	44	32
End	TGA	100	0	100	0	33	59
	TAG	0	0	0	0	19	21
	TAA	0	100	0	100	48	20
Gln	CAA	65	61	74	53	59	38
	CAG	35	39	26	47	41	62
Glu	GAG	100	24	98	36	51	71
	GAA	0	76	2	64	49	29
Gly	GGT	67	37	64	44	33	20
	GGC	32	36	36	22	16	42
	GGA	1	20	0	19	38	19
	GGG	0	8	0	16	12	20
His	CAC	62	40	72	31	46	62
	CAT	38	60	28	69	54	38
Ile	ATC	73	34	65	24	37	58
	ATT	27	51	35	59	45	28
	ATA	0	15	0	17	18	14
Leu	CTC	54	11	59	7	28	26
	TTG	29	17	25	32	26	15
	CTT	16	9	15	7	19	17
	TTA	0	18	0	19	10	5
	CTG	0	32	0	29	9	29
	CTA	0	13	0	7	8	8
Lys	AAG	99	79	99	75	61	78
	AAA	1	21	1	25	39	22
Met	ATG	100	100	100	100	100	100
Phe	TTC	100	42	100	41	55	71
	TTT	0	58	0	59	45	29
Pro	CCA	74	30	91	26	42	26
	CCT	22	28	7	20	32	22
	CCC	4	14	3	7	17	24
	CCG	0	27	0	47	9	28
Ser	TCC	47	19	55	11	18	23
	TCT	35	15	30	15	25	15
	AGC	18	22	15	18	18	23
	AGT	0	20	0	31	14	9
	TCG	0	7	0	8	6	14
	TCA	0	17	0	17	19	16
Thr	ACC	60	41	64	31	30	37
	ACT	28	25	32	34	35	20
	ACA	12	21	4	18	27	21
	ACG	0	13	0	18	8	22
Trp	TGG	100	100	100	100	100	100
Tyr	TAC	100	24	100	19	57	73
	TAT	0	76	0	81	43	27
Val	GTC	69	27	73	11	20	31
	GTG	21	17	22	27	29	39
	GTT	10	34	3	48	39	21
	GTA	0	22	2	14	12	8

## EXAMPLE 1

Design Of Plant Codon-Biased Genes Encoding W-14 Peptides  
TcbA and TcdA

The coding strands of the native DNA sequences of the *Photorhabdus* W-14 genes encoding peptides TcbA and TcdA were scanned for the presence of deleterious sequences such as the Shaw/Kamen RNA destabilizing motif ATTTA, 5 intron splice recognition sites, and poly A addition motifs. This was done using the MacVector Sequence Analysis Software (Oxford Molecular Biology Group, Symantec Corp.), using a custom Nucleic Acid Subsequence File. The native sequence was also searched for runs of 10 4 or more of the same base.

Motif searching of the native W-14 *tcbA* and *tcdA* genes revealed the presence of many potentially deleterious sequences in the protein coding strands, as summarized in Table 4. Not shown, but also present, were 15 many runs of four or more single residues (e.g. the native *tcbA* gene has 81 runs of four A's).

Table 4

Native Gene	ATTTA	5' Splice	3' Splice	Poly A Addition*	RNAP II term.
<i>tcbA</i>	18	7	17	46	0
<i>tcdA</i>	18	7	13	77	1

\* Totals of 16 different motifs.

Analyses of eukaryotic genes and plant genes in 20 particular have shown that CG & TA doublets are underrepresented, while the genes are enriched in CT & TG doublets. The sequences of the hemicot biased genes have accordingly been adjusted to encompass these base compositions and to have G+C compositions of about 53%, 25 similar to many plant genes. When compared to the native W-14 *tcbA* and *tcdA* genes, the plant-biased genes have a much more uniform G+C distribution.

Nucleotide changes to remove potentially deleterious sequences were chosen to simultaneously adjust the codon 30 composition of the coding region to more closely reflect that of plant genes. A framework for these changes was provided by the codon bias tables prepared for maize and dicot genes shown in Table 3.

Comparison of codon compositions of the native W-14 genes to maize and dicot genes revealed that the W-14 genes contain a very different preference set of the degenerate codons for the 18 amino acids for which there 5 is a choice (Table 3). For each of 8 amino acids (Phe, Tyr, Cys, Arg, Asn, Lys, Glu, and Gly) in both W-14 genes, the most abundant codon is different from the preferred codons found in either maize or dicot genes. One might expect that translational difficulties would be 10 encountered in efforts to produce in plants proteins (such as TcbA and TcdA) having high relative amounts of these amino acids from mRNAs having large numbers of nonpreferred codons. There is a marked difference in distribution of the codon compositions specifying the 15 other 10 amino acids. For His, Gln, Ile, Val, and Asp, the dicot-preferred codons are found as the most abundant ones in both W-14 genes. For Leu, Thr, Ser, and Ala, the maize preferred codons are the most abundant codon choices found in the *tcdA* gene. In contrast, the *tcbA* 20 gene contains only the CCG (Pro) maize-preferred codon as the highest abundance choice.

In making the codon choices, doublet contents were considered, so that adjacent codons preferably did not form CG or TA doublets (which are underrepresented in 25 eukaryotic genes; 1, 4), while CT or TG doublets (which are enriched in eukaryotic genes *ibid.*) were created when possible.

Choices were also made to utilize a diversity of codons for Met, Trp, Asn, Asp, Cys, Glu, His, Ile, Lys, 30 Phe, Thr, and Tyr.

The sequences were also designed to encode unique 6-bp recognition sites for restriction enzymes, spaced about every 1200 bp. Finally, an additional codon (GCT; Ala) was inserted at the second position to encode an Nco 35 I recognition site encompassing the ATG (Met) start codon. Additional recognition sites were included after

the stop codon to facilitate subsequent cloning steps into expression vectors. These features are set forth above in Tables 1 and 2.

The new *tcdA* and *tcbA* genes of SEQ ID NO:3 and SEQ 5 ID NO:4 share 73.5%, and 72.6% identity, respectively, to their native W-14 counterparts (Wisconsin Genetics Computer Group, GAP algorithm).

#### B. Gene Synthesis

The complete synthesis of the plant codon-biased 10 *tcbA* and *tcdA* genes was performed under contract by Operon Technologies, Inc. (OPTI, Alameda, CA). Basically, chemically synthesized oligonucleotides of appropriate sequence were assembled into DNA pieces about 500 bases long. These were joined together end-to-end 15 (presumably by means of appropriately placed restriction enzyme sites) into four larger pieces of roughly 2 kilobase pairs (kbp) each; therefore each comprised about 1/4 of the entire coding region of the particular gene. DNA sequence of the pieces was confirmed at this step. 20 If mistakes in sequence were present, the appropriate oligonucleotides were re-synthesized, and the assembly process was repeated. Once gene fractional parts were sequence verified, they were assembled in pairs to make the gene halves, and again sequence verified. Finally, 25 the two halves were joined, and the sequences of the junctions between the halves was verified. Therefore, each part of the new gene was sequence verified at least twice.

It should be noted that attempts to express the 30 native *tcbA* or *tcdA* genes in standard *Escherichia coli* cloning strains suggests that production of these proteins is lethal. Lethality problems may be encountered if standard cloning vectors having leaky expression from inherent *lacZ* promoters are used to 35 assemble these genes.

C. Addition Of Endoplasmic Reticulum Targeting Peptide To  
Tcda Coding Region

It is known to those in the field of plant gene expression that proteins are specifically directed into 5 the endoplasmic reticulum (ER) by means of a short signal peptide which is removed during or after the transport process through the ER membrane. The mature (processed) protein is incorporated into the ER endomembrane or is released into the ER lumen where the transported protein 10 may be uniquely folded (aided by chaperonins), modified by glycosylation, accumulated in the vacuole, or additionally translocated (by secretion). These processes are reviewed by Gomord and Faye [V. Gomord and L. Faye, (1996) *Signals and mechanisms involved in 15 intracellular transport of secreted proteins in plants*. Plant Physiol. Biochem. 34:165-181] and by Bar-Peled et al. [M. Bar-Peled, D. C. Bassham, and N. V. Raikhel, (1996) *Transport of proteins in eukaryotic cells: more 20 questions ahead*. Plant Molec. Biology 32:223-249]. It is also known that the subcellular recognition mechanisms for an ER signal peptide are evolutionarily somewhat 25 conserved, since the ER signal for a protein normally produced in monocot (maize) cells is recognized and processed normally by dicot (tobacco) cells. This is exemplified by the maize 15 kDa zein ER signal peptide [L. M. Hoffman, D. D. Donaldson, R. Bookland, K. Rashka, and E. M. Herman, (1987) *Synthesis and protein body 30 deposition of maize 15-kd zein in transgenic tobacco seeds*. EMBO J. 6:3213-3221, and U.S. Patent 5589616]. Further, it is known that the ER signal peptide derived 35 from one protein can direct the translocation of a different protein if it is appropriately attached to the second protein by genetic engineering methods [D. C. Hunt and M. J. Chrispeels, (1991) *The signal peptide of a vacuolar protein is necessary and sufficient for the efficient secretion of a cytosolic protein*. Plant

Physiol. 96:18-25, and Denecke, J., J. Botterman, and R. Deblaere (1990) *Protein secretion in plants can occur via a default pathway*. Plant Cell 2:51-59]. Therefore, one may expose a protein *in vivo* to different biochemical environments by directing its accumulation in the cytosol (by not providing a signal peptide sequence), or in the ER/vacuole (by provision of an appropriate signal peptide.)

The ER signal peptide of maize 15 kDa zein proteins is known to comprise the first 20 amino acids encoded by the zein coding region. Two examples of such signal peptides the ER signal peptide of 15 kDa zein from A5707 maize, NCBI Accession # M72708, and the ER signal peptide of 15 kDa zein from Black Mexican Sweet maize, NCBI Accession # M13507. There is only a single amino acid difference (Ser vs Cys at residue 17) between these signal peptides.

SEQ ID NO:5 is a modified sequence coding the ER signal peptide of 15 kDa zein from Black Mexican Sweet maize. The modifications embodied in this sequence were made to accommodate the different monocot/dicot codon usages and other sequence motif considerations discussed above in the design of the plant-optimized *tcdA* coding region. The sequence includes an additional Ala residue at position #2 to accommodate the *NcoI* site which spans the ATG start codon.

SEQ ID NO:6 gives a sequence coding for the full-length native *TcdA* protein (amino acids 22-2537) fused to the modified 15 kDa zein endoplasmic reticulum signal peptide (amino acids 1-21).

#### Example 2

##### Transformation Of Tobacco With *Agrobacterium* Carrying Plasmid pDAB2041 Encoding *Photobacterium* Toxins

###### A. Plasmid pDAB2041

Preparation of tobacco transformation vectors was accomplished in three steps. First, a modified plant-optimized *tcdA* coding region was ligated into a tobacco

plant expression cassette plasmid. In this step, the coding region was placed under the transcriptional control of a promoter functional in tobacco plant cells. RNA transcription termination and polyadenylation were 5 mediated by a downstream copy of the terminator region from the *Agrobacterium* nopaline synthase gene. Two plasmids designed to function in this role are pDAB1507 and pDAB2006. In the second step, the complete gene comprised of the promoter, coding region, and terminator 10 region was ligated between the T-DNA borders of an *Agrobacterium* binary vector, pDAB1542. Also positioned between the T-DNA borders was a plant selectable marker gene to allow selection of transformed tobacco plant cells. In the third step, the engineered binary vector 15 plasmid was conjugated from its *E. coli* host strain into a disabled *Agrobacterium tumefaciens* strain capable of transforming tobacco plant cells that regenerate into fertile transgenic plants.

It is a feature of plasmid pDAB1507 that any coding 20 region having an *Nco*I site at its 5' end and a *Sac*I site 3' to the coding region, when cloned into the unique *Nco*I and *Sac*I sites of pDAB1507, is placed under the transcriptional control of an enhanced version of the CaMV 35S promoter. It is also a feature of pDAB1507 that 25 the 5' untranslated leader (UTR) sequence preceding the *Nco*I site comprises a modified version of the 5' UTR of the MSV coat protein gene, into which has been cloned an internally deleted version of the maize *Adh1S* intron 1. Additionally it is a feature of pDAB1507 that 30 transcription termination and polyadenylation of the mRNA containing the introduced coding region are mediated by termination/Poly A addition sequences derived from the nopaline synthase (Nos) gene. Finally, it is a feature 35 of pDAB1507 that the entire assembly of promoter/coding region/3'UTR can be obtained as a single DNA fragment by cleavage at the flanking *Not*I sites.

It is a feature of plasmid pDAB2006 that any coding region having an *Nco*I site at its 5' end and a *Sac*I site 3' to the coding region, when cloned into the unique *Nco*I and *Sac*I sites of pDAB2006, is placed under the transcriptional control of the CaMV 35S promoter. It is also a feature of pDAB2006 that the 5' untranslated leader (UTR) sequence preceding the *Nco*I site comprises a polylinker. Additionally it is a feature of pDAB2006 that transcription termination and polyadenylation of the mRNA containing the introduced coding region are mediated by termination/Poly A addition sequences derived from the nopaline synthase (Nos) gene. Finally, it is a feature of pDAB2006 that the entire assembly of promoter/coding region/3'UTR can be obtained as a single DNA fragment by cleavage at the flanking *Not*I sites.

It is a feature of pDAB1542 that any DNA fragment flanked by *Not*I sites can be cloned into the unique *Not*I site of pDAB1542, thus placing the introduced fragment between the T-DNA borders, and adjacent to the neomycin phosphotransferase II (kanamycin resistance) gene.

To prepare a plant-expressible gene to produce the non-targeted *TcdA* protein in tobacco plant cells, DNA of a plasmid (pAOH\_4-OPTI) containing the plant-optimized *tcdA* coding region, (SEQ ID No:3) was cleaved with restriction enzymes *Nco*I and *Sac*I, and the large 7550 bp fragment was ligated to similarly-cut DNA of plasmid pDAB1507 to produce plasmid pDAB2040. DNA of pDAB2040 was then digested with *Not*I, and the 8884 bp fragment was ligated to *Not*I digested DNA of pDAB1542 to produce plasmid pDAB2041. This plasmid was then conjugated by triparental mating [Firoozabady, E., D. L. DeBoer, D. J. Merlo, E. L. Halk, L. N. Amerson, K. E. Rashka, and E. E. Murray (1987) *Transformation of cotton (Gossypium hirsutum L.) by Agrobacterium tumefaciens and regeneration of transgenic plants.* Plant Molec. Biol.

10:105-116] from the host *Escherichia coli* strain (XL1-Blue, Stratagene, La Jolla, CA), into the nontumorigenic *Agrobacterium tumefaciens* strain EHA101S, which is a spontaneous streptomycin-resistant mutant of strain  
5 EHA101 (Hood, E. E., G. L. Helmer, R. T. Fraley, and M.-D. Chilton (1986) *The hypervirulence of Agrobacterium tumefaciens A281 is encoded in a region of pTiBo542 outside of T-DNA.* J. Bacteriol. 168:1291-1301). Strain EHA101S(pDAB2041) was then used to produce transgenic  
10 tobacco plants that expressed the TcdA protein.

B. Plasmid pRK2013

To prepare a plant-expressible gene to produce the endoplasmic reticulum-targeted TcdA protein in tobacco plant cells, DNA of a plasmid (pAOH\_4-ER) containing the  
15 plant-optimized, ER-targeted *tcdA* coding region, (SEQ ID No:6) was cleaved with restriction enzymes *Nco*I and *Sac*I, and the large 7610 bp fragment was ligated to similarly-cut DNA of plasmid pDAB2006 to produce plasmid pDAB1833. DNA of pDAB1833 was then digested with *Not*I, and the 8822  
20 bp fragment was ligated to *Not*I digested DNA of pDAB1542 to produce plasmid pDAB2052. This plasmid was then conjugated by triparental mating from the host *Escherichia coli* strain (XL1-Blue), into the nontumorigenic *Agrobacterium tumefaciens* strain EHA101S.  
25 Strain EHA101S(pDAB2052) was then used to produce transgenic tobacco plants that expressed the TcdA protein containing an amino terminus endoplasmic reticulum targeting peptide.

30 C. Transfer of Plasmid pDAB2041 Into *Agrobacterium* Strain EHA101S

Cultures of *E. coli* carrying the engineered Ti plasmid pDAB2041 (plasmid containing the rebuilt Toxin A gene, *tcdA*), *E. coli* carrying the plasmid pRK2013, and  
35 *Agrobacterium* strain EHA101S were grown overnight, then mixed 1:1:1 on plain LB medium solidified with agar and

cultured in the dark at 28°C. Two days later, the lawn of bacteria was scraped up with a loop, suspended in plain LB medium, vortexed, and then diluted 1:10<sup>4</sup>, 1:10<sup>5</sup>, and 1:10<sup>6</sup> fold in plain LB liquid medium. Aliquots of these 5 dilutions were spread on selective plates containing medium YEP plus erythromycin (100 mg/L) and streptomycin (250 mg/L) and grown at 28°C. Two days later, single colonies were picked and streaked onto the same medium, then spread to give single colonies. Single colonies were 10 picked again and streaked, then spread for single colonies. Single colonies were picked a third time, grown as streaks, then subjected to a quality analysis involving growth on lactose medium and chromogenic assay with Benedict's reagent. Of ten strains developed in this 15 way, the fastest coloring colony was chosen for further work.

D. Transformation Of Tobacco With *Agrobacterium* Carrying Plasmid pDAB2041

20 Tobacco transformation with *Agrobacterium tumefaciens* was carried out by a method similar, but not identical, to published methods (R Horsch et al, 1988. Plant Molecular Biology Manual, S. Gelvin et al, eds., Kluwer Academic Publishers, Boston). To provide source 25 tissue for the transformation, tobacco seed (*Nicotiana tabacum* cv. Kentucky 160) were surface sterilized and planted on the surface of TOB-, which is a hormone-free Murashige and Skoog medium (T. Murashige and F. Skoog, 1962). A revised medium for rapid growth and bioassays 30 with tobacco tissue culture. Plant Physiol. 75: 473-497) solidified with agar. Plants were grown for 6-8 weeks in a lighted incubator room at 28-30°C and leaves were collected steriley for use in the transformation protocol. Approximately one cm<sup>2</sup> pieces were steriley cut 35 from these leaves, excluding the midrib. Cultures of the

*Agrobacterium* strains (EHA101S containing pDAB2041), which had been grown overnight on a rotor at 28°C, were pelleted in a centrifuge and resuspended in sterile Murashige & Skoog salts, adjusted to a final optical 5 density of 0.7 at 600 nm. Leaf pieces were dipped in this bacterial suspension for approximately 30 seconds, then blotted dry on sterile paper towels and placed right side up on medium TOB+ (Murashige and Skoog medium containing 1 mg/L indole acetic acid and 2.5 mg/L 10 benzyladenine) and incubated in the dark at 28°C. Two days later the leaf pieces were moved to medium TOB+ containing 250 mg/L cefotaxime (Agri-Bio, North Miami, Florida) and 100 mg/L kanamycin sulfate (AgriBio) and incubated at 28-30°C in the light. Leaf pieces were moved 15 to fresh TOB+ with cefotaxime and kanamycin twice per week for the first two weeks and once per week thereafter. Leaf pieces which showed regrowth of the *Agrobacterium* strain were moved to medium TOB+ with cefotaxime and kanamycin, plus 100 mg/l carbenicillin 20 (Sigma). Four to six weeks after the leaf pieces were treated with the bacteria, small plants arising from transformed foci were removed from this tissue preparation and planted into medium TOB- containing 250 mg/L cefotaxime and 100 mg/L kanamycin in Magenta GA7 25 boxes (Magenta Corp., Chicago). These plantlets were grown in a lighted incubator room. After 3-4 weeks the primary transgenic plants had rooted and grown to a size sufficient that leaf samples could be analyzed for expression of protein from the transgene. Twenty-five 30 independent transgenic events were recovered as single plants from the pDAB2041 transformation.

Eight independent lines expressing various levels of transgenic protein from the T-DNA of pDAB2041 were propagated *in vitro* from leaf pieces as follows. Twelve 35 to sixteen approximately one cm<sup>2</sup> pieces were steriley cut from leaves of each primary transgenic plant, excluding

the midrib and all naturally occurring edges. These leaf pieces were placed on medium TOB+ containing 250 mg/L cefotaxime and 100 mg/L kanamycin, and cultured in the lighted incubator at 28-30°C for 3-4 weeks, at which time 5 small plants could be cut from the proliferating tissue mass. Several small plantlets from each transgenic line were moved into Magenta boxes containing medium TOB- plus cefotaxime and kanamycin and allowed to root and grow. The proliferating tissue mass was further cultured on 10 medium TOB+ with cefotaxime and kanamycin, and additional plants could be cut out and grown up as needed.

Plants were moved into the greenhouse by washing the agar from the roots, transplanting into soil in 5 1/2" square pots, placing the pot into a Ziploc bag 15 (DowBrands), placing plain water into the bottom of the bag, and placing in indirect light in a 30°C greenhouse for one week. After one week the bag could be opened; the plants were fertilized and allowed to grow further, until the plants were acclimated and the bag was removed. 20 Plants were grown under ordinary warm greenhouse conditions (30°C, 16 H light). Plants were suitable for sampling four weeks post transplant.

Example 3

25 Chacterization Of Transgenic Tobacco Plants Expressing  
Photorhabdus Toxin That Confer Insect Control.

A. Polyclonal Antibody Production

The *E. coli* produced recombinant TcdA protein was 30 purified by a series of column purification. The protein was sent to Berkley Antibody Company (Richmond, CA) for the production of antiserum in a rabbit. Inoculations with the antigen were initiated with 0.5 mg of protein followed by four boosting injections of 0.25 mg each at 35 about three week intervals. The rabbit serum was tested by the standard Western analysis using the recombinant TcdA protein as the antigen and enhanced chemi-

luminescens, ECL method (Amersham, Arlington Heights, IL) . The antibodies (PAb-EA<sub>0</sub>) were purified using a PURE I antibody purification kit (Sigma, St. Luis, MO). PAb-EA<sub>0</sub> antibodies recognize the full-length TcdA and its  
5 processed components.

B. Expression Of TcdA Protein In Tobacco

Protein was extracted from the leaf tissue of transformed and non-transformed tobacco plants following the procedure described immediately below.

10 Two leaf disks of 1.4 cm in diameter were harvested from the middle portion of a fully expanded leaf. The disks were placed on a 1.6 x 4 cm piece of 3M Whatman paper. The paper was folded lengthwise and inserted in a flexible straw. Four hundred micro liters of the  
15 extraction buffer (9.5 ml of 0.2 M NaH<sub>2</sub>PO<sub>4</sub>, 15.5 ml of 0.2 M Na<sub>2</sub>HPO<sub>4</sub>, 2 ml of 0.5 M Na<sub>2</sub>EDTA, 100 ml of Triton X100, 1 ml of 10% Sarkosyl, 78 ml of beta-mercaptoethanol, H<sub>2</sub>O to bring total volume to 100 ml) was pipetted on to the paper. The straw containing the sample was then passed  
20 through a rolling device used for squeezing out the extract 1.5 mL micro centrifuge tube was placed at the other end of the straw to collect the extract. The extract was centrifuged for 10 minutes at 14,000 rpm in an Eppendorf regrigerated microcentrifuge. The  
25 supernatant was transferred into a new tube. Protein quantitation analysis was performed using the standard Bio-Rad Protein Analysis protocol (Bio-Rad Laboratories, Hercules, CA). The extract was diluted to 2 mg/ml of total protein using the extraction buffer.  
30 For the detection of transgenic protein, Western blot analysis was performed. Following a standard procedure for protein separation (Laemmli, 1970), 40 µg of protein was loaded in each well of 4-20% gradient polyacrylamide gel (Owl Scientific Co., MA) for  
35 electrophoresis. Subsequently, the protein was

transferred onto a nitrocellulose membrane using a semi-dry electroblotter (Pharmacia LKB Biotechnology, Piscataway, NJ). The membrane was incubated for one hour in Blotto (5% milk in TBST solution; 25 mM Tris HCL pH 5.4, 136 mM NaCl, 2.7 mM KCl, 0.1% Tween 20). Thereafter, Blotto was replaced by the primary antibody solution (in Blotto). After one hour in the primary antibody, the membrane was washed with TBST for five minutes three times. Then the secondary antibody in Blotto (1:2000 dilution of goat anti-rabbit IgG conjugated to horseradish peroxidase; Bio-Rad Laboratories) was added to the membrane. After one hour of incubation, the membrane was washed with an excess amount of TBST for 10 minutes four times. The protein was visualized by using the enhanced chemi-luminescens, ECL method (Amersham, Arlington Heights, IL). The differential intensity of the protein bands were measured using densitometer (Molecular Dynamics Inc., Sunnyvale, CA).

To determine the expression of TcdA protein in tobacco transformed with pDAB2041, PAb-EA<sub>0</sub> antibodies were used as the primary antibodies. The expression levels of TcdA protein varied among independent transformation events. The primary plant generated from the event #2041-13 showed the highest level of pre-pro TcdA expression of extractable protein. When the leaf pieces from this plant (#2041-13) were used in *in vitro* propagation, several plants were obtained. Seven of these plants were analyzed for the expression of the TcdA protein. All but one plant produced the full-length TcdA protein as well as some processed peptide components. Using the antibodies specific to Neomycin phosphotransferase, NPT (5 prime-3 prime, Boulder, Co), the expression the selectable marker gene (*npt II*) was detected. Similar results were obtained for #2041-29.

## Western analysis of plants derived from event #2041-13.

Plant #	TcdA	NPT (selectable marker)
2041-13A	+	not done
2041-13B	+	not done
2041-13-1	-	+
2041-13-2	+	+
2041-13-3	+	+
2041-13-4	+	+
2041-13-5	+	+

## C. Nucleic Acid Analysis of Transgenic Tobacco Lines

Genomic DNA was prepared from a group of 2041

5 transgenic events. The lines included Magenta box stage 2041-13, and greenhouse stage plants 2041-13-1, 2041-13-2, 2041-13-5, 2041-9, 2041-20A and 2041-20B. A transgenic GUS line (2023) was included as a negative control. Southern analysis of these lines was performed.

10 The genomic tobacco DNA was restricted with the enzyme SstI which should result in a 8.9 kb hybridization product when hybridized to a *tcdA* gene specific probe. The 8.9 kb hybridization product should consist of the 35T promoter and the *tcdA* coding region. All 2041 plants

15 contained a band of the expected size. Events 2041-9 and -20 appear to be the same line with 5 identical hybridizing bands. Event 2041-13 produced 6 hybridization fragments with the *tcdA* coding region probe. Magenta box and various greenhouse plants of

20 2041-13 all produced the same hybridization profile. This hybridization pattern was different from that of events 2041-9 and -20.

RNA analysis, using the *tcdA* coding region probe, was performed on the same group of greenhouse 2041

25 plants. Immunoblot analysis had revealed that plants 2041-9, 2041-20A, 2041-20B, and 2041-13-1 produced no detectable TcdA protein; while 2041-13-2 and 2041-13-5 produced substantial amounts of full-length TcdA. Northern analysis was in agreement with the immunoblot

result. A faint RNA signal was detected for plants 2041-9, 2041-20A, 2041-20B, and 2041-13-1. Only faintly visible was a band corresponding to full-length *tcdA* transcript in plant 2041-13-1. In contrast, for plants 5 2041-13-2 and 2041-13-5 a strong RNA signal was detected, with a substantial amount of full-length size (~8.0 kb) *tcdA* transcript. These data support the observed bioassay activity for this group of plants.

Genomic DNA was prepared from a second functionally 10 active 2041 transgenic event, 2041-29. Southern analysis of this line was performed. A transgenic GUS line (2023) was included as a negative control, DNA of line 2041-9 was included as a positive control.

The genomic tobacco DNAs were restricted with the 15 enzyme *Sst*I which should result in a 8.9 kb hybridization product when hybridized to a *tcdA* gene specific probe. The 8.9 kb hybridization product should consist of the 35T promoter and the *tcdA* coding region. For plant 2041-29-5, three hybridization products larger than 8.9 kb the 20 were detected with the *tcdA* gene specific probe. Immunoblot analysis has demonstrated pre-pro TcdA protein is made by this plant, it is therefore likely that a 25 restriction site was lost during transformation or regeneration, or the 2041-29 genomic DNA was not thoroughly digested.

#### D. Tobacco Leaf-Disk Tests With Tobacco Hornworm Exhibiting Insect Control

Leaves were sampled from tobacco plants, *Nicotiana* 30 *tabaco*, previously transplanted into the greenhouse. A single leaf was sampled from each plant on each test date. Leaves were selected from the zone where younger elongate leaves transition into older ovate leaves. Excised leaves were placed into 12 oz. cups with the 35 petiole submerged in water to maintain turgor, and transported to the laboratory.

Eight, 1.4 cm disks were cut from the center portion of one side of each leaf (right adaxial side up, with distal portion facing away from the observer). Each disk was placed individually into a well of a C-D

5 International 128 well tray (Pitman, NJ.) into which 0.5 ml of a 1.6% aqueous agar solution had been previously pipetted. The solidified agar prevented the leaf disks from drying out. The adaxial surface of the disk was always oriented up.

10 A single neonate tobacco hornworm, *Manduca sexta*, was placed on each disk and the wells were sealed with vented plastic lids. The assay was held at 27°C and 40% RH. Larval mortality and live-weight data were collected after 3 days. Data were subjected to analysis of  
 15 variance and Duncan's multiple range test ( $\alpha = 0.05$ ) (Proc GLM, SAS Institute Inc., Cary, NC.). Data were transformed using a logarithmic function to correct a correlation between the magnitude of the mean and variance.

20 Table 6  
 Results of leaf-disk assays from greenhouse grown tobacco plants with event 2041-13.

TRT	Plant	Plant Age	Weight of Surviving Larvae (mg) & Duncan's Group <sup>1</sup>				
			Pretest	Test 1	Test 2	Test 3	3 Test Sum.
13	non-transformed - 2	young	---	---	---	18.8 a*	---
14	non-transformed - 3	young	---	---	---	17.0 ab	---
16	non-transformed - 5	young	---	---	---	16.4 ab	---
3	2041-13-1 (western -)	young	---	17.6 a	18.2 a	16.1 ab	17.3 a
9	Gus Control	old	19.3 a	14.6 a	16.3 a	14.5 ab	15.1 a
10	non-transformed - 1	young	---	8.3 b	16.8 a	13.9 b	13.0 b
11	2041-20B (western -)	old	---	10.0 b*	13.7 ab	14.6 ab	12.9 b
15	non-transformed - 4	young	---	---	---	13.0 bc	---
8	2041-20A (western -)	old	15.7 a	8.3 b	11.3 bc	9.2 cd	9.6 c
12	2041-9 (western -)	old	19.5 a	---	---	7.9 d	---
7	2041-13-5 (western +)	young	---	6.3 bc	9.6 cd	7.2 de	7.7 d
5	2041-13-3 (western +)	young	---	6.4	6.2 e	6.8 de**	6.4 de
				bc****			
1	2041-13A (western +)	old	7.2 b	6.8 bc*	7.0 de*	5.4 e	6.4 de
6	2041-13-4 (western +)	young	---	4.9 c****	5.8 e	7.6 d	6.4 de
4	2041-13-2 (western +)	young	---	5.7 bc	5.7 e**	7.5 d	6.3 de
2	2041-13B (western +)	old	---	4.7 c**	5.6 e	7.2 de	5.9 e

\* Number of stars corresponds to the number of dead larvae per 8 tested.

1. Data transformed (logarithm) for analysis.  
 Means followed by the same letter are not significantly different (alpha = 0.05).

5

TABLE 7  
 Results Of Leaf-Disk Assays From Greenhouse Grown Tobacco Plants  
 With Event 2041-29.

Plant	MEAN WGT (MG) / Duncan's Group				
	Test 1	Test 2	Test 3	Test 4	Four Test Summary
2014-6 GUS 1	15.8 a	16.6a	**5.5bc	*12.9ab	13.2 a
2014-6 GUS 2	14.4 a	*6.6 bc	*13.4a	15.2a	12.6 a
KY-160 NTC	13.4 a	6.7 bc	7.9b	8.5bc	9.1 b
2041-29 4P	*4.9 b	*7.3b	****6.9b	*****	6.3 c
2041-29 7	*5.9 b	5.1bc	****6.7b	***7.2c	6.1 c
2041-29 3P	*5.6 b	**7.9b	****6.5b	***3.6d	5.9 c
2041-29 2P	6.3 b	****4.7c	*****4.1c	*****4.6d	5.4 c

\* Number of stars corresponds to the number of dead larvae per 8 tested.

10 1. Data transformed (logarithm) for analysis.

Means followed by the same letter are not significantly different (alpha = 0.05).

15 All event 2041-29 plants significantly depressed THW larval weight gain compared to control plants. Average weight depression was 49%. Statistically significant mortality occurred in THW larvae exposed to foliage from 2041-29 plants. Mortality averaged 37.5% compared to 5.2% in controls.

20

E. Isolation and Characterization of Functional Photorhabdus Toxin Protein From Transgenic Plants

25 Seven grams of transgenic tobacco plants (2041-13) expressing TcdA (Toxin A) gene were homogenized with 10 ml 50 mM Potassium Phosphate buffer, pH 7.0 using a bead beater (Biospec Products, Bartlesville, OK) according to manufacturer's instructions. The homogenate was filtered through four layers of cheese cloth and then centrifuged at 35,000 g for 15 min. The supernant was collected and 30 filtered through 0.22  $\mu$ m Millipore Express<sup>TM</sup> membrane. It was then applied to a Superdex 200 column (2.6 x 40 cm)

which had been equilibrated with 20 mM Tris buffer, pH 8.0 (Buffer A). The protein was eluted in Buffer A at a flow rate of 3 ml/min. Fractions with 3 ml each were collected and subjected to southern corn rootworm (SCR) 5 bioassay. It was found that fractions corresponding to a native molecular weight around 860 kDa had the highest insecticidal activity. Western analysis of the active fraction using a polyclonal antibody specific to Toxin A indicated the presence of full-length TcdA peptide. The 10 active fractions were further combined and applied to a Mono Q 10/10 column which had been equilibrated with Buffer A. Proteins bound to the column were then eluted by a linear gradient of 0 to 1 M NaCl in Buffer A. Fractions with 2 ml each were collected and analyzed by 15 both SCR bioassay and Western using antibody specific to Toxin A. The results again demonstrated the correlation between insecticidal activity and presence of full-length TcdA peptide.

20 F. Characterization of Progeny Transgenic Plants

The inheritability of the genetically engineering plants containing the *Photorhabdus* toxin gene was evaluated by generating F1 progeny. Progeny was generated from 2041-13 event by selfing expression 25 positive plants. The 2041-13 plants in the greenhouse were allowed to self-pollinate. Seed capsules were collected when mature and were allowed to dry and after-ripen on the laboratory bench for two weeks. Seed from plant designated 2041-13A was surface-sterilized and 30 distributed on the surface of medium TOB- without selection, to allow recovery of nonexpressing or nontransgenic progeny as well as expressing and segregating transgenic siblings. Seed was germinated in a C lighted incubator room (16 H light, 28 C). After 1 35 month, fifty-one seedlings, designated 2041-13A-S1 through S51, were distributed into Magenta boxes

containing medium TOB- to grow further. Three weeks later, leaf samples from these Magenta-box grown seedlings were submitted for evaluation of the level of expression of TcdA toxin.

5 Leaf samples were tested for kanamycin response by placing sterile leaf segments on medium TOB+ containing 100 mg/L kanamycin in the light and scoring for tissue growth and color after two weeks. All leaf pieces showed some positive response, indicating complex segregation.

10 10 This group of in vitro grown event 2041-13 progeny seedlings were all transplanted into the greenhouse approximately two months after seeding onto medium, using the following method. After washing the agar from the roots, plants were transplanted into 5 ½ inch square pots  
15 in a soil mix containing 75% MetroMix and 25% mineral soil. They were enclosed in a zip-lock bag and plain water added to leave 1-2 inches of water in the bottom of the bag after soil absorption. These bags were closed and placed under a cart in the greenhouse to protect them  
20 from direct sunlight. The bags were opened after 5-6 days, and removed after 7 days, when the plants were adapted to soil and were moved to the top of the cart for normal greenhouse culture. Plants were ready to test in insect bioassays at four weeks post transplant.

25 F1 progeny were evaluated for expression of protein toxin by immunological screen and for biological activity by plant bioassays, as described previously, using tobacco hornworm. There existed a positive correlation between levels of expression protein toxin and degree of  
30 growth inhibition and at higher expression levels mortality was observed. The biological activity was observed to be statistical significance with high confidence levels between populations of non-transformed and transformed expressing protein toxin.

35 The following table summarizes the results of insect (tobacco hornworm) bioassays conducted with F1 progeny of

self-fertilized 2041-13 plants genetically engineered to produce the "204" A toxin. The tests included 6 non-expressing progeny (protein-negative controls), 45 toxin A expressors, and 4 non-transformed controls (KY-160).

5 Results are from three leaf-disk assays (method previously outlined) where eight disks were used per test. The data were analyzed using analysis of variance and were blocked by test.

The treatment effect for each of these analyses 10 indicated the  $Pr > F$  was less than 0.0001. The Toxin A expressors produced significant control of tobacco hornworm compared to each of the control groups based on each of the three measures of efficacy. The two control groups behaved similarly. Statistical analysis using 15 ANOVA and an LSD test with alpha equal to 0.01 (or 1%) showed differences between the 3 groups. The LSD test indicated that the non-expressors and the non-transformed plants were similar in larvae weights but the expressors gave weights significantly lower than either of the other 20 two groups of plants. These data demonstrated that the genetic basis for insect control was inheritable and corresponded to the presence of expressed toxin gene.

25 Table 8  
Tobacco hornworm results from F1 progeny of self-fertilized  
2041-13 tobacco plants.

Treatment Group	Mean Value and Duncan's Grouping <sup>d</sup>		
	Total Weight (mg) <sup>a</sup>	Survivor Weight (mg) <sup>b</sup>	Leaf Area (cm <sup>2</sup> ) <sup>c</sup>
Non-transformed Control	15.8 a	15.8 a	1.2 a
Protein-negative Control	16.4 a	16.5 a	1.2 a
Toxin A Expressor	8.1 b	9.2 b	4.9 b

<sup>a</sup> Average insect weight with dead insects considered to weigh nothing.

30 <sup>b</sup> Average insect weight with dead insects excluded from analysis.

<sup>c</sup> Total leaf area remaining per eight leaf disks. Initial area was approximately 12 cm<sup>2</sup>.

<sup>d</sup> Means followed by the same letter are not significantly different (alpha = 0.05).

## Example 4

5 Transformation Of Maize With a Vector Carrying Plasmid  
pDAB1834 Encoding *Photorhabdus* ToxinsA. Preparation Of Maize Transformation Vectors  
Containing Modified Plant-Optimized *Tcda* Coding Regions:  
Plasmid Pdab1834

10 Preparation of maize transformation vectors was  
accomplished in two steps. First, a modified plant-  
optimized *tcdA* coding region was ligated into a plant  
expression cassette plasmid. In this step, the coding  
15 region was placed under the transcriptional control of a  
promoter functional in maize plant cells. RNA  
transcription termination and polyadenylation were  
mediated by a downstream copy of the terminator region  
from the *Agrobacterium* nopaline synthase gene. One  
20 plasmid designed to function in this role is pDAB1538. In  
the second step, the complete gene comprised of the  
promoter, coding region, and 3' UTR terminator region was  
ligated to a plant transformation vector that contained a  
plant expressible selectable marker gene which allowed  
25 the selection of transformed maize plant cells amongst a  
background of nontransformed cells. An example of such a  
vector is pDAB367.

It is a feature of plasmid pDAB1538 that any coding  
region having an *Nco*I site at its 5' end and a *Sac*I site  
30 3' to the coding region, when cloned into the unique *Nco*I  
and *Sac*I sites of pDAB1538, is placed under the  
transcriptional control of the maize ubiquitin1 (ubil)  
promoter. It is also a feature of pDAB1538 that the 5'  
untranslated leader (UTR) sequence preceding the *Nco*I  
35 site comprises a polylinker. Additionally it is a  
feature of pDAB1538 that transcription termination and  
polyadenylation of the mRNA containing the introduced  
coding region are mediated by termination/Poly A addition

sequences derived from the nopaline synthase (Nos) gene. Finally, it is a feature of pDAB1538 that the entire assembly of promoter/coding region/3' UTR can be obtained as a single DNA fragment by cleavage at the flanking *NotI* sites.

It is a feature of pDAB367 that the phosphinothricin acetyl transferase protein, which has as its substrate phosphinothricin and related compounds, is produced in plant cells through transcription of its coding region mediated by the Cauliflower Mosaic Virus 35S promoter and that termination of transcription plus polyadenylation are mediated by the nopaline synthase terminator region. It is further a feature of pDAB367 that any DNA fragment containing flanking *NotI* sites can be cloned into the unique *NotI* site of pDAB367, thus physically linking the introduced DNA fragment to the aforementioned selectable marker gene.

To prepare a maize plant-expressible gene to produce the endoplasmic reticulum-targeted *TcdA* protein in plant cells, DNA of a plasmid (pAOH\_4-ER) containing the plant-optimized, ER-targeted *tcdA* coding region, (SEQ ID No:6) was cleaved with restriction enzymes *NcoI* and *SacI*, and the large 7610 bp fragment was ligated to similarly-cut DNA of plasmid pDAB1538 to produce plasmid pDAB1832. DNA of pDAB1832 was then digested with *NotI*, and the 9984 bp *NotI* fragment was ligated into the unique *NotI* site of pDAB367 to produce plasmid pDAB1834.

It is a feature of plasmids pDAB1834 that the ubil and 35S promoters are encoded on the same DNA strand.

30

#### B. Transformation and Regeneration of Transgenic Maize Isolates

Type II callus cultures were initiated from immature zygotic embryos of the genotype "Hi-II." (Armstrong et al, (1991) Maize Genet. Coop. Newslett., 65: 92-93). Embryos were isolated from greenhouse-grown ears from

crosses between Hi-II parent A and Hi-II parent B or  $F_2$  embryos derived from a self- or sib-pollination of a Hi-II plant. Immature embryos (1.5 to 3.5 mm) were cultured on initiation medium consisting of N6 salts and vitamins

5 (Chu et al, (1978) *The N6 medium and its application to anther culture of cereal crops.* Proc. Symp. Plant Tissue Culture, Peking Press, 43-56), 1.0 mg/L 2,4-D, 25mM L-proline, 100 mg/L casein hydrolysate, 10 mg/L  $AgNO_3$ , 2.5 g/L GELRITE (Schweizerhall, South Plainfield, NJ), and 20 10 g/L sucrose, with a pH of 5.8. After four to six weeks callus was subcultured onto maintenance medium (initiation medium in which  $AgNO_3$  was omitted and L-proline was reduced to 6 mM). Selection for Type II callus took place for ca. 12-16 weeks.

15 Plasmid pDAB1834 was transformed into embryogenic callus. For blasting, 140  $\mu$ g of plasmid DNA was precipitated onto 60 mg of alcohol-rinsed, spherical gold particles (1.5 - 3.0  $\mu$ m diameter, Aldrich Chemical Co., Inc., Milwaukee, WI) by adding 74  $\mu$ L of 2.5M  $CaCl_2$   $H_2O$  and 20 30  $\mu$ L of 0.1M spermidine (free base) to 300  $\mu$ L of plasmid DNA and  $H_2O$ . The solution was immediately vortexed and the DNA-coated gold particles were allowed to settle. The resulting clear supernatant was removed and the gold 25 particles were resuspended in 1 ml of absolute ethanol. This suspension was diluted with absolute ethanol to obtain 15 mg DNA-coated gold/mL.

30 Approximately 600 mg of embryogenic callus tissue was spread over the surface of Type II callus maintenance medium as described herein lacking casein hydrolysate and L-proline, but supplemented with 0.2 M sorbitol and 0.2 M mannitol as an osmoticum. Following a 4 h pre-treatment, tissue was transferred to culture dishes containing blasting medium (osmotic media solidified with 20 g/L TC agar (PhytoTechnology Laboratories, LLC, Shawnee Mission, 35 KS) instead of 7 g/L GELRITE. Helium blasting accelerated suspended DNA-coated gold particles towards

and into the prepared tissue targets. The device used was an earlier prototype of that described in US Patent 5,141,131 which is incorporated herein by reference. Tissues were covered with a stainless steel screen (104  $\mu\text{m}$  openings) and placed under a partial vacuum of 25 inches of Hg in the device chamber. The DNA-coated gold particles were further diluted 1:1 with absolute ethanol prior to blasting and were accelerated at the callus targets four times using a helium pressure of 1500 psi, with each blast delivering 20  $\mu\text{L}$  of the DNA/gold suspension. Immediately post-blasting, the tissue was transferred to osmotic media for a 16-24 h recovery period. Afterwards, the tissue was divided into small pieces and transferred to selection medium (maintenance medium lacking casein hydrolysate and L-proline but containing 30 mg/L BASTA® (AgrEvo, Berlin, Germany)). Every four weeks for 3 months, tissue pieces were non-selectively transferred to fresh selection medium. After 7 weeks and up to 22 weeks, callus sectors found proliferating against a background of growth-inhibited tissue were removed and isolated. The resulting BASTA®-resistant tissue was subcultured biweekly onto fresh selection medium. Following western analysis, positive transgenic lines were identified and transferred to regeneration media. Western-negative lines underwent subsequent RNA spot blot analysis to identify negative controls for regeneration.

Regeneration was initiated by transferring callus tissue to cytokinin-based induction medium, which 30 consisted of Murashige and Skoog salts, hereinafter MS salts, and vitamins (Murashige and Skoog, (1962) *Physiol. Plant.* 15: 473-497) 30 g/L sucrose, 100 mg/L myo-inositol, 30 g/L mannitol, 5 mg/L 6-benzylaminopurine, hereinafter BAP, 0.025 mg/L 2,4-D, 30 mg/L BASTA®, and 35 2.5 g/L GELRITE at pH 5.7. The cultures were placed in low light (125 ft-candles) for one week followed by one

week in high light (325 ft-candles). Following a two week induction period, tissue was non-selectively transferred to hormone-free regeneration medium, which was identical to the induction medium except that it 5 lacked 2,4-D and BAP, and was kept in high light. Small (1.5-3 cm) plantlets were removed and placed in 150x25 mm culture tubes containing SH medium (SH salts and vitamins (Schenk and Hildebrandt, (1972) Can. J. Bot. 50:199-204), 10 g/L sucrose, 100 mg/L myo-inositol, 5 mL/L FeEDTA, and 10 2.5 g/L GELRITE, pH 5.8). Plantlets were transferred to 12 cm pots containing approximately 0.25 kg of METRO-MIX 360 (The Scotts Co. Marysville, OH) in the greenhouse as soon as they exhibited growth and developed a sufficient root system. They were grown with a 16 h photoperiod 15 supplemented by a combination of high pressure sodium and metal halide lamps, and were watered as needed with a combination of three independent Peters Excel fertilizer formulations (Grace-Sierra Horticultural Products Company, Milpitas, CA). At the 6-8 leaf stage, plants 20 were transplanted to five gallon pots containing approximately 4 kg METRO-MIX 360, and grown to maturity.

#### EXAMPLE 5

##### Characterization Of Transgenic Maize Plants

###### 25 Expressing *Photorhabdus* Toxin That Confer Insect Control.

###### A. Insect Bioassays

A single leaf was sampled from each plant in each test. Eight, 1.4 cm disks were cut from the outer portion of each leaf (approximately 30cm long) avoiding the 30 center vein. Each disk was placed individually into a well of a C-D International 128 well tray (Pitman, NJ.) into which 0.5 ml of a 1.6% aqueous agar solution had been previously pipetted. The solidified agar prevented the leaf disks from drying out. The adaxial surface of 35 the disk was always oriented up.

Five neonate southern corn rootworms, *Diabrotica undecimpunctata howardi*, were placed on each disk and the wells were sealed with vented plastic lids. The assay was held at 27°C and 40% RH. Larval mortality and live-  
 5 weight data were collected after 3 days. Data were subjected to analysis of variance and Duncan's multiple range test ( $\alpha = 0.05$ ) (Proc GLM, SAS Institute Inc., Cary, NC.). Weight data were transformed using a logarithmic function to correct a correlation between the magnitude  
 10 of the mean and variance.

TABLE 9  
 Results of Maize Leaf-disk Test vs SCR

Treatment	Mean % Kill (Duncan's)	Mean Survival Weight (mg) (Duncan's)
1834 - 11	68 A	0.064 A
1834 - 17	44 B	0.098 B
1834 - 15	26 BC	0.127 C
HiII control	13 C	0.161 C

Note: Means followed by the same letter are not  
 15 significantly different based on Duncan's multiple range test ( $\alpha=0.05$ ). Insect groups weighing less than 0.1 mg were set to 0.03 mg instead of zero to conduct a more conservative analysis. Mortality ( $\text{arcsin}(\text{sqrt})$ ) and weight( $\log_{10}$ ) data were transformed for analyses.  
 20  
 The results shown in Table 9 demonstrated that two events expressing TcdA protein were statistically distinct from control lines bioassayed using SCR neonates by mortality and survival weight criteria. These results demonstrated that  
 25 southern corn rootworm were functionally effected by feeding on maize plants containing and expressing the *tcdA* gene. Those plants from 1834-11 were used to generate progeny for testing of inheritability of transgene.

B. PRODUCTION AND PROGENY TEST OF *tcdA* TRANSGENIC MAIZE

Origin and growth of progeny plants: Sibling plants 1834-11-07 and 1834-11-08, clonally derived by regeneration 5 from the callus of transgenic maize event 1834-11, were transplanted to the greenhouse and pollinated with inbred 0Q414. Seeds obtained from these crosses, comprising seed lots 1834-11-07A and 1834-11-08A, were planted in 10 Roottrainers (1  $\frac{1}{2}$  inch x 2 inch x 8 inch deep, product #647, C. Hummert Intl., Earth City, Mo.) filled with Metro-Mix 360 soilless mix (Scotts Terra-Lite, available from Hummert Intl.) and top irrigated with Hoagland's 15 nutrient solution. (Hoagland's solution contains 229 ppm nitrogen as nitrate, 24.6 ppm nitrogen as ammonium, 26 ppm P, 157 ppm K, 187 ppm Ca, 49 ppm Mg. and 30 ppm Na.)

Greenhouse conditions for this trial were: 16 hour days, daylight supplemented by metal halide lamps as needed to achieve a minimum of 600  $\text{Einstens/cm}^2$  PAR, and ambient temperature 30 C days, 22 C nights.

20 Leaves were sampled for protein determination approximately one week after planting. Leaf bioassays were conducted 2-3 weeks after planting; root bioassays were initiated approximately 3 weeks post planting.

25 Protein analysis of progeny plants: Protein was extracted from leaf and root samples harvested from transgenic plants, line 1834-11 progenies, and non-transformed plants. Each sample was placed on a 1.6 x 4 cm piece of 30 3M Whatman<sup>TM</sup> paper. The paper was folded lengthwise and inserted in a flexible straw. A volume of 350  $\mu\text{l}$  of an extraction buffer (9.5 ml of 0.2 M  $\text{Na}_2\text{PO}_4$ , 15.5 ml of 0.2 M  $\text{Na}_2\text{HPO}_4$ , 2 ml of 0.5 M  $\text{Na}_2\text{EDTA}$ , 100 ml of Triton X-100, 1 ml of 10% Sarkosyl, 78 ml of beta-mercaptoethanol,  $\text{H}_2\text{O}$  35 to bring total volume to 100 ml, 50  $\mu\text{g/ml}$  Antipain, 50  $\mu\text{g/ml}$  Leupeptin, 0.1 mM Chymostatin, 5  $\mu\text{g/ml}$  Pepstatin) was pipetted on to the paper. The straw containing the

sample was then passed through a rolling device used for squeezing the extract into a 1.5 ml microcentrifuge tube. The extract was centrifuged for 10 minutes at 14,000 rpm in an Eppendorf refrigerated micro-centrifuge. The 5 supernatant was transferred into a new tube. The amount of the total extractable protein was determined using a standard BioRad Protein Analysis protocol (BioRad Laboratories, Hercules, CA).

The presence of the TcdA protein was visualized by 10 Western blot analysis following a standard procedure for protein separation (Laemmli, 1970). A volume of twenty  $\mu$ l of extract was loaded in each well of 4-20% gradient polyacrylamide gel (Owl Scientific Co., MA) for electrophoresis. Subsequently, the protein was 15 transferred onto a nitrocellulose membrane using a semi-dry electroblotter (Pharmacia LKB Biotechnology, Piscataway, NJ). The membrane was incubated for one hour in TBST-M solution (10% milk in TBST solution; 25 mM Tris HCl pH 7.4, 136 mM NaCl, 2.7 mM KCl, 0.1% Tween 20). 20 Thereafter, the primary antibody (Anti-TcdA in TBST-M) was added. After one hour, the membrane was washed with TBST for five minutes, three times. Then the secondary antibody solution (goat anti-rabbit IgG conjugated to horseradish peroxidase; Bio-Rad Laboratories, in TBST-M) 25 was added to the membrane. After one hour of incubation, the membrane was washed with an excess amount of TBST for 10 minutes, four times. The protein was visualized using the Super Signal<sup>®</sup> West Pico chemiluminescence method (Pierce Chemical Co., Rockford, IL). The protein blot 30 was exposed on a Hyper-film (Amersham, Arlington Heights, IL) and was developed within 3 minutes. The intensity of the protein band was measured using a densitometer (Molecular Dynamics Inc., Sunnyvale, CA) and compared to standards.

35 Three of six plants from seed lot 1834-11-07A and three of six plants from seed lot 1834-11-08A produced

detectable levels of TcdA protein (Table 1). Approximately 3.8 to 13.3 ppm of TcdA were detected in the leaf blades and 4.1 to 8.4 ppm were detected in the leaf tips of the protein-positive plants. The amounts of 5 TcdA protein detected in the roots were slightly lower than those found in the leaves.

Insect bioassays with progeny plants: Plants were selected for bioassay based on results from Western blot 10 analysis. Twelve (12), 6.4 mm diameter leaf discs were cut from the youngest leaf of each 2 week old seedling. Each disc was placed in a well of a 128-well tray (CD International) containing approximately 0.5mL of a solidified 2% agar in water solution. Two neonate 15 southern corn rootworm, *Diabrotica undecimpunctata* *howardi* (Barber) (SCR), were placed in each well with a leaf disc. Trays were covered with perforated lids and maintained under a controlled environment for 3 days (28 C; 16 hours light:8 hours dark; approx. 60% relative 20 humidity). Living larvae from 4 leaf discs were pooled and weighed producing 3 weight determinations per plant. Average weights were calculated by dividing the pooled 25 weight by the number of survivors. Differences in average weights of SCR fed leaf discs from protein positive and protein negative plants were assessed using analysis of variance on the natural log-transformed average weights (Minitab, v. 12.2, Minitab Inc., State College, PA).

30 Root bioassays were initiated approximately 1 week after the initiation of the leaf disc bioassays. Approximately 24h prior to eclosion, SCR eggs were suspended in a 0.15% solution of agar in water to a concentration of 100 eggs/ml. Plants were inoculated 35 with SCR eggs by pipetting 2.0 ml of the egg suspension (ie., approximately 200 eggs) just below the soil surface at the base of each plant. Two weeks after inoculation, plants were removed from their Rootrainer pots, their

roots washed free of potting mix, and scored for rootworm damage based on a 1 (resistant) to 9 (susceptible) rating system (Welch, 1977). The results of the root ratings were examined using non-parametric tests to determine if 5 the distribution of root ratings from the protein positive plants was the same as the distribution of the ratings from the protein negative plants. Testing was done at the 5% significance level. (StatXact v.3, CYTEL Software Corporation, Cambridge MA)

10

Results from leaf and root bioassays of tcdA protein positive and protein negative progeny plants are summarized in Table 10. The average weights of SCR larvae fed leaf discs from protein positive plants were 15 significantly lower than those of larvae fed leaf discs from protein negative plants ( $F = 4.6$ ;  $d.f. = 1, 34$ ;  $P \leq 0.001$ ). The Kolmogorov-Smirnov 2 sample test ( $p=0.04$ ) and the Wald Wolfowitz runs test ( $p=0.001$ ) indicated that the protein positive and protein negative root rating 20 distributions were not similar. The Wilcoxon- Mann-Whitney test ( $p=0.0206$ ) and the Normal Scores test ( $p=0.206$ ) indicated that the average score for the protein positive plants was lower than the average root rating from the protein negative plants.

25

Table 10. Protein analysis and insect bioassay results with progeny of TcdA transgenic maize.

Plant Number	TcdA Protein	Leaf Disc Bioassay Avg. Wt. (mg)	Root Bioassay Root Rating (1-9)
1834-11-07A-30	PRO-	0.190	8
1834-11-08A-21	PRO-	0.196	9
1834-11-08A-16	PRO-	0.195	9
1834-11-08A-14	PRO-	0.137	9
1834-11-07A-22	PRO-	0.208	9
1834-11-07A-20	PRO-	0.175	9

1834-11-07A-26	PRO+	0.118	9
1834-11-08A-17	PRO+	0.132	8
1834-11-07A-14	PRO+	0.110	2
1834-11-07A-11	PRO+	0.106	4
1834-11-08A-28	PRO+	0.129	8
1834-11-08A-27	PRO+	0.108	4

DNA analysis of progeny plants: Leaf samples from 1834-11.7A and 1834-11.8A progeny plants were in conical 50 ml polypropylene tubes and dried in a Labconco Freeze Dry 5 Lyophilizer (Kansas City, MO) for 1-2 days. Lyophilized leaves were then ground in a Tecator Cyclotec 1093 Sample mill grinder (Hoganas, Sweden) and stored at -20C. Genomic DNA was extracted by the following procedure: (1) to a 25 ml Conical tube containing 300-500 mg of ground 10 tissue, 9 ml of CTAB (cetyl trimethylammonium bromide solution) was added, and incubated at 65°C for 1 hour; (2) 4.5 ml of chloroform: octanol (24:1) was added and mixed gently for 5 minutes; (3) samples were centrifuged at 2000 rpm and DNA was precipitated from the supernatant 15 with an equal volume of isopropanol; (4) DNA was collected on a glass hook, washed in ethanol, and dissolved in TE (10 mM Tris.HCl, 0.5 mM EDTA, pH8.0).

Genomic DNA was digested at 37 °C. for 2 hours in an 20 Eppendorf tube containing the following mixture: 8 µl of 800ug/ml DNA, 2 µl 1 mg/ml BSA (Bovine serum albumin), 2 µl 10x buffer, 1 µl SacI, 1 µl EcoRI, and 6 µl H2O. Digested DNA samples were electrophoresed overnight at 40 mA in a 0.85% SeaKem LE agarose gel (FMC, Rockland, 25 Maine). The gel was blotted onto Millipore Immobilon-Ny+ (Bedford, MA) membrane overnight in 20X SSC (NaCl 175.2 g/l, Na citrate 88 g/l). The probe DNA was cut with BamHI/SacI (NEB, Beverly, MA) from pDAB1551 plasmid, which released a 7356 bp fragment containing the open 30 reading frame of the rebuilt tcdA gene. This 7356 bp fragment was labeled with P32 using a Stratagene Prime-it

RmT dCTP-Labeling Reactions kit (La Jolla, CA) and used for Southern hybridization. Hybridization was conducted in hybridization buffer (10% polyethylene glycol, 7% SDS [Sodium dodecyl sulfate], 0.6X SSC, 10 mM NaPO<sub>4</sub>, 5 mM EDTA, 10 µg/ml denatured salmon sperm) at 60 °C overnight. After hybridization, the membrane was washed with 10X SSC plus 0.1% SDS at 60 °C for 30 min and exposed to X ray film (Hyperfilm® MP, Amersham Life Sciences, Piscataway, NJ) for 1-2 days.

10

Results summarized indicate that a pattern of 8 hybridizing bands (the size of the expected fragment and larger) cosegregated with protein expression in 50% of all progeny assayed. These results are characteristic of 15 a complex insertion at a single site. All seedlings containing the insert also expressed toxin protein.

20 Example 6  
Transformation Of Rice With a Vector Carrying Plasmid  
pDAB1553 Encoding *Photorhabdus* Toxins

A. Plasmid pDAB1553

25 Plasmid pDAB1553 containing *tcdA* driven by the maize ubiquitin1 promoter and *hpt* (hygromycin phosphotransferase providing resistance to the antibiotic hygromycin) under the control of 35T (a modified 35S promoter), was used for transformation.

30 Preparation of rice transformation vectors was accomplished in two steps. First, a modified plant-optimized *tcdA* coding region was ligated into a rice plant expression cassette plasmid. In this step, the coding region was placed under the transcriptional control of a promoter functional in plant cells. RNA 35 transcription termination and polyadenylation were mediated by a downstream copy of the terminator region from the *Agrobacterium* nopaline synthase gene. One

plasmid designed to function in this role is plasmid pDAB1538 (described in the section on maize transformation vectors). In the second step, the complete gene comprised of the promoter, coding region, 5 and terminator region was ligated to a rice plant transformation vector that contained a plant expressible selectable marker gene which allowed the selection of transformed rice plant cells amongst a background of nontransformed cells. An example of such a vector is 10 pDAB354-Not1.

It is a feature of pDAB354-Not1 that the hygromycin phosphotransferase protein, which has as its substrate hygromycin B and related compounds, is produced in plant cells through transcription of its coding region mediated 15 by the Cauliflower Mosaic Virus 35S promoter and that termination of transcription plus polyadenylation are mediated by the nopaline synthase terminator region. It is further a feature of pDAB354-Not1 that any DNA fragment containing flanking NotI sites can be cloned 20 into the unique NotI site of pDAB354-Not1, thus physically linking the introduced DNA fragment to the aforementioned selectable marker gene.

To prepare a plant-expressible gene to produce the non-targeted TcdA protein in rice plant cells, DNA of a 25 plasmid (pAOH\_4-OPTI) containing the plant-optimized *tcda* coding region, (SEQ ID No:3) was cleaved with restriction enzymes *Nco*I and *Sac*I, and the large 7550 bp fragment was ligated to similarly-cut DNA of plasmid pDAB1538 to produce plasmid pDAB1551. DNA of pDAB1551 was then 30 digested with *Not*I, and the large 9933 bp fragment was ligated to *Not*I digested DNA of pDAB354-Not1 to produce plasmid pDAB1553.

It is a feature of plasmid pDAB1553 that the *ubil* and 35S promoters are encoded on the same DNA strand.

35 B. Production of Rice transgenics

For initiation of embryogenic callus, mature seeds of a *Japonica* cultivar, Taipei 309 were dehusked and surface-sterilized in 70% ethanol for 2-5 min. followed by a 30-45 min soak in 50% commercial bleach (2.6% sodium 5 hypochlorite) with a few drops of 'Liquinox' soap. The seeds were then rinsed 3 times in sterile distilled water and placed on filter paper before transferring to 'callus induction' medium (i.e., NB). The NB medium consisted of N6 macro elements (Chu, 1978, The N6 medium and its 10 application to anther culture of cereal crops. Proc. Symp. Plant Tissue Culture, Peking Press, p43-56), B5 micro elements and vitamins (Gamborg et al., 1968, Nutrient requirements of suspension cultures of soybean root cells. *Exp. Cell Res.* 50: 151-158), 300 mg/L casein 15 hydrolysate, 500 mg/L L-proline, 500 mg/L L-glutamine, 30 g/L sucrose, 2 mg/L 2,4-dichloro-phenoxyacetic acid (2,4-D), and 2.5 g/L gelrite (Schweizerhall, NJ) with the pH adjusted to 5.8. The mature seed cultured on 'induction' media were incubated in the dark at 28°C. After 3 weeks 20 of culture, the emerging primary callus induced from the scutellar region of mature embryo was transferred to fresh NB medium for further maintenance.

About 140 µg of plasmid pDAB1553 DNA was precipitated onto 60 mg of 1.0 micron (Bio-Rad) gold 25 particles as described herein.

For helium blasting, actively growing embryogenic callus cultures, 2-4 mm in size, were subjected to a high osmoticum treatment. This treatment included placing of callus on NB medium with 0.2 M mannitol and 0.2 M 30 sorbitol (Vain et al., 1993, Osmoticum treatment enhances particle bombardment-mediated transient and stable transformation of maize. *Plant Cell Rep.* 12: 84-88) for 4 h before helium blasting. Following osmoticum 35 treatment, callus cultures were transferred to 'blasting' medium (NB+2% agar) and covered with a stainless steel screen (230 micron). The callus cultures were blasted at

2,000 psi helium pressures twice per target. After blasting, callus was transferred back to the media with high osmoticum overnight before placing on selection medium, which consisted NB medium with 30 mg/L hygromycin. After 2 weeks, the cultures were transferred to fresh selection medium with a higher concentration of selection agent, i.e., NB+50mg/L hygromycin (Li et al., 1993, An improved rice transformation system using the biolistic method. Plant Cell Rep. 12: 250-255).

10 Compact, white-yellow, embryogenic callus cultures, recovered on NB+50 mg/L hygromycin, were regenerated by transferring to 'pre-regeneration' (PR) medium + 50 mg/L hygromycin. The PR medium consisted of NB medium with 2 mg/L benzyl aminopurine (BAP), 1 mg/L naphthalene acetic acid (NAA), and 5 mg/L abscisic acid (ABA). After 2 weeks of culture in the dark, they were transferred to 'regeneration' (RN) medium. The composition of RN medium is NB medium with 3 mg/L BAP, and 0.5 mg/L NAA. The cultures on RN medium were incubated for 2 weeks at 15 28° C under high fluorescent light (325-ft-candles). The plantlets with 2 cm shoot were transferred to 1/2 MS medium (Murashige and Skoog, 1962, A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol. Plant.15:473-497) with 1/2 B5 vitamins, 10 g/L sucrose, 0.05 mg/L NAA, 50 mg/L hygromycin and 2.5 g/L gelrite adjusted to pH 5.8 in magenta boxes. When 20 plantlets were established with well-developed root systems, they were transferred to soil (1 metromix: 1 top soil) and raised in the greenhouse (29/24°C day/night cycle, 50-60% humidity, 12 h photoperiod) until maturity.

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## EXAMPLE 7

Chacterization Of Transgenic Rice Plants Expressing  
35 Photorhabdus Toxin That Confer Insect Control.

A. Insect bioassays

Insect bioassays were performed using leaf discs and shown to be highly effective in controlling Southern corn rootworm. *Diabrotica undecimpunctata howardi* eggs are obtained from French Ag Research and hatched in petri dishes held at 28.5°C and 40% RH. The aerial parts are sampled from the transgenic plants and placed, singly into inverted petri dishes (100x15mm) containing 15ml of 1.6% aqueous agar in the bottom to provide humidity and filter paper in the top to absorb condensation. These preparations are infested with five neonate larvae per dish and held at 28.5°C and 40% RH for 3 days. Mortality and larval weights are recorded. Weight data were transformed using a logarithmic function to correct a correlation between the magnitude of the mean and variance.

Table 11

Treatment	Average Survivor Weight in mg <sup>1</sup> (Duncan's Grouping)	Presence TcdA greenhouse-grown plants (number of +/number of plants tested)
GUS Control	0.390 A	-
1553-33	0.170 BCD	++
1553-44	0.167 BCD	+++
1553-62	0.125 CD	+++
1553-41	0.100 D	+++

Note: Means followed by the same letter are not significantly different based on Duncan's multiple range test (alpha=0.05).

Insect groups weighing less than 0.1 mg were set to 0.03 mg instead of zero to conduct a more conservative analysis. Weight data were transformed (Log10) for analyses. A single replicate was used on each of three test dates. Plants were sampled from magenta boxes. The results demonstrate that in leaf disc bioassays, several rice events derived by transformation with *tcdA* gene were demonstrated to statistically have a functional affect on corn rootworm neonate.

## Claims

1. An isolated nucleic acid of SEQ ID NO: 3 or SEQ ID NO: 4.
2. A transgenic monocot cell having a genome comprising SEQ ID NO:3 or SEQ ID NO:4.
3. A transgenic dicot cell having a genome comprising SEQ ID NO:3 or SEQ ID NO:4.
4. A transgenic plant with a genome comprising a nucleic acid of SEQ ID NO: 3 or SEQ ID NO:4 that imparts insect resistance.
5. A transgenic plant of claim 4 wherein the plant is rice.
6. A transgenic plant of claim 4 wherein the plant is maize.
- 15 7. A transgenic plant of claim 4 wherein the plant is tobacco.

## SEQUENCE LISTING

<110> Petell, Jim  
 Merlo, Donald  
 Herman, Rod  
 Roberts, Jean  
 Guo, Lining  
 Schafer, Barry  
 Sukhapinda, Kitisri  
 Owens Merlo, Ann

<120> Transgenic Plants Expressing *Photorhabdus* Toxin

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<140>

<141>

<150> US 60/148,356

<151> 1999-08-11

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<170> PatentIn Ver. 2.0

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 85 90 95

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cgt cct tcc ggc gca acg cct tat cat gat gct tat gaa aat gtg cgt Arg Pro Ser Gly Ala Thr Pro Tyr His Asp Ala Tyr Glu Asn Val Arg 195	200	205	624
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Pro	Gln	Val	Asn	Ile	Glu	Tyr	Ser	Ala	Asn	Ile	Thr	Leu	Asn	Thr	Ala	
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Gly	Ser	Trp	Ala	Tyr	Ala	Ala	Lys	Phe	Thr	Val	Glu	Glu	Tyr	Asn		
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Gln	Tyr	Ser	Phe	Leu	Leu	Lys	Leu	Asn	Lys	Ala	Ile	Arg	Leu	Ser	Arg	
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Phe	Ser	Thr	Gly	Asp	Glu	Glu	Ile	Asp	Leu	Asn	Ser	Gly	Ser	Thr	Gly	
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 1380 1385 1390

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 Val Thr Phe Thr Ala Phe Ala Glu Asp Gly Arg Lys Leu Gly Tyr Glu  
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agt ttc agt att cct gtt acc ctc aag gta agt acc gat aat gcc ctg 4752  
 Ser Phe Ser Ile Pro Val Thr Leu Lys Val Ser Thr Asp Asn Ala Leu  
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 Thr Leu His His Asn Glu Asn Gly Ala Gln Tyr Met Gln Trp Gln Ser  
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 Tyr Arg Thr Arg Leu Asn Thr Leu Phe Ala Arg Gln Leu Val Ala Arg  
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 Lys Ser Pro Ser Asp Gly Thr Trp Trp Gly Pro His Phe Val Arg Asp  
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 1730 1735 1740

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 Thr Ser Trp Asn Ser Asp Pro Leu Asp Ser Val Asp Pro Asp Ala Val  
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 Arg Asp Thr Leu Asn Glu Ala Lys Met Trp Tyr Met Gln Ala Leu His  
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 1985 1990 1995 2000

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2100	2105	2110	6336
caa gcc atg acg cta cga gcg tcc gcc ggg ctt acc acg gca gtt Gln Ala Met Thr Leu Arg Ala Ser Ala Ala Gly Leu Thr Thr Ala Val			
2115	2120	2125	6384
cag gca tcc cgt ctg gcc ggt gcg gct gat ctg gtg cct aac atc Gln Ala Ser Arg Leu Ala Gly Ala Ala Asp Leu Val Pro Asn Ile			
2130	2135	2140	6432
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gat aaa att agc caa tct gaa acc tac cgt cgc cgt cag gag tgg Asp Lys Ile Ser Gln Ser Glu Thr Tyr Arg Arg Arg Arg Gln Glu Trp			
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gag atc cag cgg aat aat gcc gaa gcg gaa ttg aag caa atc gat gct Glu Ile Gln Arg Asn Asn Ala Glu Ala Glu Leu Lys Gln Ile Asp Ala			
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tgc ctg atg gca gaa caa gct tac cgt tgg gaa ctc aat gat gac tct Cys Leu Met Ala Glu Gln Ala Tyr Arg Trp Glu Leu Asn Asp Asp Ser			
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gcc cgc ttc att aaa ccg ggc gcc tgg cag gga acc tat gcc ggt ctg Ala Arg Phe Ile Lys Pro Gly Ala Trp Gln Gly Thr Tyr Ala Gly Leu			
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His Leu Lys Arg Asp Lys Arg Ala Leu Glu Val Glu Arg Thr Val Ser	
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Leu Ala Glu Val Tyr Ala Gly Leu Pro Lys Asp Asn Gly Pro Phe Ser	
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Gly Ser Gly Asn Asn Asn Leu Ala Phe Gly Ala Gly Thr Asp Thr Lys	
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Thr Ser Leu Gln Ala Ser Val Ser Phe Ala Asp Leu Lys Ile Arg Glu	
2385 2390 2395 2400	
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Asp Tyr Pro Ala Ser Leu Gly Lys Ile Arg Arg Ile Lys Gln Ile Ser	
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Leu Ser Tyr Gly Asp Lys Ala Gly Leu Ala Asn Gly Cys Glu Ala Leu	
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Tyr Thr Ile Lys	
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 Gln Leu Thr Cys Pro Ala Glu Ile Ala Leu Tyr Pro Phe Asp Thr Phe  
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 Arg Glu Lys Thr Arg Gly Met Val Asn Trp Gly Glu Ala Lys Arg Ile  
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 Tyr Glu Ile Ala Gln Ala Glu Gln Asp Arg Asn Leu Leu His Glu Lys  
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 Arg Ile Phe Ala Tyr Ala Asn Pro Leu Leu Lys Asn Ala Val Arg Leu  
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 Gly Thr Arg Gln Met Leu Gly Phe Ile Gln Gly Tyr Ser Asp Leu Phe  
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 Gly Asn Arg Ala Asp Asn Tyr Ala Ala Pro Gly Ser Val Ala Ser Met  
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 Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu Tyr Arg Glu Ala Lys Asn  
 115 120 125  
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 Leu His Asp Ser Ser Ile Tyr Tyr Leu Asp Lys Arg Arg Pro Asp  
 130 135 140  
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 Leu Ala Ser Leu Met Leu Ser Gln Lys Asn Met Asp Glu Glu Ile Ser  
 145 150 155 160  
 acg ctg gct ctc tct aat gaa ttg tgc ctt gcc ggg atc gaa aca aaa 528  
 Thr Leu Ala Leu Ser Asn Glu Leu Cys Leu Ala Gly Ile Glu Thr Lys  
 165 170 175  
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 Thr Gly Lys Ser Gln Asp Glu Val Met Asp Met Leu Ser Thr Tyr Arg  
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 Leu Ser Gly Glu Thr Pro Tyr His His Ala Tyr Glu Thr Val Arg Glu  
 195 200 205



Arg Leu Leu Lys Ala Thr Gly Leu S r Phe Ala Thr Leu Glu Arg Ile			
450	455	460	
gtt gat agt gtt aat agc acc aaa tcc atc acg gtt gag gta tta aac			1440
Val Asp Ser Val Asn Ser Thr Lys Ser Ile Thr Val Glu Val Leu Asn			
465	470	475	480
aag gtt tat cgg gta aaa ttc tat att gat cgt tat ggc atc agt gaa			1488
Lys Val Tyr Arg Val Lys Phe Tyr Ile Asp Arg Tyr Gly Ile Ser Glu			
485	490	495	
gag aca gcc gct att ttg gct aat att aat atc tct cag caa gct gtt			1536
Glu Thr Ala Ala Ile Leu Ala Asn Ile Asn Ile Ser Gln Gln Ala Val			
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ggc aat cag ctt agc cag ttt gag caa cta ttt aat cac ccg ccg ctc			1584
Gly Asn Gln Leu Ser Gln Phe Glu Gln Leu Phe Asn His Pro Pro Leu			
515	520	525	
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Asn Gly Ile Arg Tyr Glu Ile Ser Glu Asp Asn Ser Lys His Leu Pro			
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aat cct gat ctg aac ctt aaa cca gac agt acc ggt gat gat caa cgc			1680
Asn Pro Asp Leu Asn Leu Lys Pro Asp Ser Thr Gly Asp Asp Gln Arg			
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Lys Ala Val Leu Lys Arg Ala Phe Gln Val Asn Ala Ser Glu Leu Tyr			
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Gln Met Leu Leu Ile Thr Asp Arg Lys Glu Asp Gly Val Ile Lys Asn			
580	585	590	
aac tta gag aat ttg tct gat ctg tat ttg gtt agt ttg ctg gcc cag			1824
Asn Leu Glu Asn Leu Ser Asp Leu Tyr Leu Val Ser Leu Leu Ala Gln			
595	600	605	
att cat aac ctg act att gct gaa ttg aac att ttg ttg gtg att tgt			1872
Ile His Asn Leu Thr Ile Ala Glu Leu Asn Ile Leu Leu Val Ile Cys			
610	615	620	
ggc tat ggc gac acc aac att tat cag att acc gac gat aat tta gcc			1920
Gly Tyr Gly Asp Thr Asn Ile Tyr Gln Ile Thr Asp Asp Asn Leu Ala			
625	630	635	640
aaa ata gtg gaa aca ttg ttg tgg atc act caa tgg ttg aag acc caa			1968
Lys Ile Val Glu Thr Leu Leu Trp Ile Thr Gln Trp Leu Lys Thr Gln			
645	650	655	
aaa tgg aca gtt acc gac ctg ttt ctg atg acc acg gcc act tac agc			2016
Lys Trp Thr Val Thr Asp Leu Phe Leu Met Thr Thr Ala Thr Tyr Ser			
660	665	670	
acc act tta acg cca gaa att agc aat ctg acg gct acg ttg tct tca			2064
Thr Thr Leu Thr Pro Glu Ile Ser Asn Leu Thr Ala Thr Leu Ser Ser			
675	680	685	
act ttg cat ggc aaa gag agt ctg att ggg gaa gat ctg aaa aga gca			2112
Thr Leu His Gly Lys Glu Ser L u Il Gly Glu Asp Leu Lys Arg Ala			

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705			710			715			720						
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Gln Val Ser Ala Asp Val Ile Thr Ser Arg Ile Ala Glu Ala Ile Ala	
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Gly Ile Gln Leu Tyr Val Asn Arg Ala Leu Asn Arg Asp Glu Gly Gln	
980 985 990	
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Leu Ala Ser Asp Val Ser Thr Arg Gln Phe Phe Thr Asp Trp Glu Arg	
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1075 1080 1085	
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Thr Tyr Tyr Trp Arg Ser Val Asp His Ser Lys Cys Glu Asn Gly Lys	
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ttt gcc gct aat gct tgg ggt gag tgg aat aaa att acc tgt gct gtc	3408
Phe Ala Ala Asn Ala Trp Gly Glu Trp Asn Lys Ile Thr Cys Ala Val	
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aat cct tgg aaa aat atc atc cgt ccg gtt gtt tat atg tcc cgc tta	3456
Asn Pro Trp Lys Asn Ile Ile Arg Pro Val Val Tyr Met Ser Arg Leu	
1140 1145 1150	
tat ctg cta tgg ctg gag cag caa tca aag aaa agt gat gat ggt aaa	3504
Tyr Leu Leu Trp Leu Glu Gln Gln Ser Lys Lys Ser Asp Asp Gly Lys	
1155 1160 1165	
acc acg att tat caa tat aac tta aaa ctg gct cat att cgt tac gac	3552
Thr Thr Ile Tyr Gln Tyr Asn Leu Lys Leu Ala His Il Arg Tyr Asp	
1170 1175 1180	

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 Gly Ser Trp Asn Thr Pro Phe Thr Phe Asp Val Thr Glu Lys Val Lys  
 1185 1190 1195 1200

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 Asn Tyr Thr Ser Ser Thr Asp Ala Ala Glu Ser Leu Gly Leu Tyr Cys  
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 Thr Gly Tyr Gln Gly Glu Asp Thr Leu Leu Val Met Phe Tyr Ser Met  
 1220 1225 1230

cag agt agt tat agc tcc tat acc gat aat aat gcg ccg gtc act ggg 3744  
 Gln Ser Ser Tyr Ser Ser Tyr Thr Asp Asn Asn Ala Pro Val Thr Gly  
 1235 1240 1245

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gca act aac tat tgg aat aac agt tat ccg caa ttt gat act gtg atg 3840  
 Ala Thr Asn Tyr Trp Asn Asn Ser Tyr Pro Gln Phe Asp Thr Val Met  
 1265 1270 1275 1280

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 Ala Asp Pro Asp Ser Asp Asn Lys Lys Val Ile Thr Arg Arg Val Asn  
 1285 1290 1295

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 Asn Arg Tyr Ala Glu Asp Tyr Glu Ile Pro Ser Ser Val Thr Ser Asn  
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agt aat tat tct tgg ggt gat cac agt tta acc atg ctt tat ggt ggt 3984  
 Ser Asn Tyr Ser Trp Gly Asp His Ser Leu Thr Met Leu Tyr Gly Gly  
 1315 1320 1325

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 Ser Val Pro Asn Ile Thr Phe Glu Ser Ala Ala Glu Asp Leu Arg Leu  
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tct acc aat atg gca ttg agt att att cat aat gga tat gcg gga acc 4080  
 Ser Thr Asn Met Ala Leu Ser Ile Ile His Asn Gly Tyr Ala Gly Thr  
 1345 1350 1355 1360

cgc cgt ata caa tgt aat ctt atg aaa caa tac gct tca tta ggt gat 4128  
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 Lys Phe Ile Ile Tyr Asp Ser Ser Phe Asp Asp Ala Asn Arg Phe Asn  
 1380 1385 1390

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 Leu Val Pro Leu Phe Lys Phe Gly Lys Asp Glu Asn Ser Asp Asp Ser  
 1395 1400 1405

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Phe Ser Ser Lys Asp Asp Asn Lys Thr Ala Asp Tyr Asn Gly Gly Thr			
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Gln Cys Ile Asp Ala Gly Thr Ser Asn Lys Asp Phe Tyr Tyr Asn Leu			
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Gln Glu Ile Glu Val Ile Ser Val Thr Gly Gly Tyr Trp Ser Ser Tyr			
1460	1465	1470	
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Lys Ile Ser Asn Pro Ile Asn Ile Asn Thr Gly Ile Asp Ser Ala Lys			
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Val Lys Val Thr Val Lys Ala Gly Gly Asp Asp Gln Ile Phe Thr Ala			
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Asp Asn Ser Thr Tyr Val Pro Gln Gln Pro Ala Pro Ser Phe Glu Glu			
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Met Ile Tyr Gln Phe Asn Asn Leu Thr Ile Asp Cys Lys Asn Leu Asn			
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Phe Ile Asp Asn Gln Ala His Ile Glu Ile Asp Phe Thr Ala Thr Ala			
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Gln Asp Gly Arg Phe Leu Gly Ala Glu Thr Phe Ile Ile Pro Val Thr			
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Lys Lys Val Leu Gly Thr Glu Asn Val Ile Ala Leu Tyr Ser Glu Asn			
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Asn Gly Val Gln Tyr Met Gln Ile Gly Ala Tyr Arg Thr Arg Leu Asn			
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Thr Leu Phe Ala Gln Gln Leu Val Ser Arg Ala Asn Arg Gly Ile Asp			
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His Gly Thr Asn Lys Ser Phe Ala Ile Glu Tyr Val Asp Ile Phe Lys			
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Glu Asn Asp Ser Ph Val Ile Tyr Gln Gly Glu Leu Ser Glu Thr Ser			

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gca tta aag aac gac agt gaa ccg atg gat ttc tct ggc gcc aat gct Ala Leu Lys Asn Asp Ser Glu Pro Met Asp Phe Ser Gly Ala Asn Ala 1745	1750	1755	1760	5280
ctc tat ttc tgg gaa ctg ttc tat tac acg ccg atg atg atg gct cat Leu Tyr Phe Trp Glu Leu Phe Tyr Tyr Pro Met Met Met Ala His 1765	1770	1775		5328
cgt ttg ttg cag gaa cag aat ttt gat gcg gcg aac cat tgg ttc cgt Arg Leu Leu Gln Glu Gln Asn Phe Asp Ala Ala Asn His Trp Phe Arg 1780	1785	1790		5376
tat gtc tgg agt cca tcc ggt tat atc gtt gat ggt aaa att gct atc Tyr Val Trp Ser Pro Ser Gly Tyr Ile Val Asp Gly Ile Ala Ile 1795	1800	1805		5424
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caa caa ctg gac tcc acc gat cca gat gct gta gcc caa gat gat ccg Gln Gln Leu Asp Ser Thr Asp Pro Asp Ala Val Ala Gln Asp Asp Pro 1825	1830	1835	1840	5520
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gcc cgt ggt gat gct gct tac cgc cag tta gag cgt gat acg ttg gct Ala Arg Gly Asp Ala Ala Tyr Arg Gln Leu Glu Arg Asp Thr Leu Ala 1860	1865	1870		5616
gaa gct aaa atg tgg tat aca cag gcg ctt aat ctg ttg ggt gat gag Glu Ala Lys Met Trp Tyr Thr Gln Ala Leu Asn Leu Leu Gly Asp Glu 1875	1880	1885		5664
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cgt cgt att aag caa atc agt gtt tcg cta cct gca ttg gtt ggg cct	7248

Arg Arg Ile Lys Gln Ile Ser Val Ser Leu Pro Ala Leu Val Gly Pro			
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Tyr Gln Asp Val Gln Ala Met Leu Ser Tyr Gly Gly Ser Thr Gln Leu			
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ccg aaa ggt tgt tca gcg ttg gct gtg tct cat ggt acc aat gat agt			7344
Pro Lys Gly Cys Ser Ala Leu Ala Val Ser His Gly Thr Asn Asp Ser			
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Gly Gln Phe Gln Leu Asp Phe Asn Asp Gly Lys Tyr Leu Pro Phe Glu			
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Gly Ile Ala Leu Asp Asp Gln Gly Thr Leu Asn Leu Gln Phe Pro Asn			
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gct acc gac aag cag aaa gca ata ttg caa act atg agc gat att att			7488
Ala Thr Asp Lys Gln Lys Ala Ile Leu Gln Thr Met Ser Asp Ile Ile			
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Gln Cys Gly Phe Asn Cys Leu Thr Asp Ile Ser His Ser Ser Phe Asn			
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gag ttc aga caa caa gtc tct gag cac ctc tcc tgg tcc gag acc cat			143
Glu Phe Arg Gln Gln Val Ser Glu His Leu Ser Trp Ser Glu Thr His			
35	40	45	
gac ctc tac cat gac gct cag caa gct cag aag gac aac agg ctc tac			191
Asp Leu Tyr His Asp Ala Gln Gln Ala Gln Lys Asp Asn Arg Leu Tyr			
50	55	60	
gag gct agg atc ctc aag agg gct aac cca caa ctc cag aac gct gtc			239
Glu Ala Arg Ile Leu Lys Arg Ala Asn Pro Gln Leu Gln Asn Ala Val			
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cac ctc gcc atc ttg gct cca aac gct gag ttg att ggt tac aac aac	287
His Leu Ala Ile Leu Ala Pro Asn Ala Glu Leu Ile Gly Tyr Asn Asn	
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Gln Phe Ser Gly Arg Ala Ser Gln Tyr Val Ala Pro Gly Thr Val Ser	
100 105 110	
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Ser Met Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu Tyr Arg Glu Ala	
115 120 125	
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Arg Asn Leu His Ala Ser Asp Ser Val Tyr Tyr Leu Asp Thr Arg Arg	
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Pro Asp Leu Lys Ser Met Ala Leu Ser Gln Gln Asn Met Asp Ile Glu	
145 150 155	
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Leu Ser Thr Leu Ser Leu Ser Asn Glu Leu Leu Leu Glu Ser Ile Lys	
160 165 170 175	
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Thr Glu Ser Lys Leu Glu Asn Tyr Thr Lys Val Met Glu Met Leu Ser	
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Thr Phe Arg Pro Ser Gly Ala Thr Pro Tyr His Asp Ala Tyr Glu Asn	
195 200 205	
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Val Arg Glu Val Ile Gln Leu Gln Asp Pro Gly Leu Glu Gln Leu Asn	
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Ala Ser Pro Ala Ile Ala Gly Leu Met His Gln Ala Ser Leu Leu Gly	
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Ile Asn Ala Ser Ile Ser Pro Glu Leu Phe Asn Ile Leu Thr Glu Glu	
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Ile Thr Glu Gly Asn Ala Glu Glu Leu Tyr Lys Lys Asn Phe Gly Asn	
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Ile Glu Pro Ala Ser Leu Ala Met Pro Glu Tyr Leu Lys Arg Tyr Tyr	
275 280 285	
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Asn Leu Ser Asp Glu Glu Leu Ser Gln Phe Ile Gly Lys Ala Ser Asn	
290 295 300	
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Phe Gly Gln Gln Glu Tyr S r Asn Asn Gln Leu Ile Thr Pro Val Val	
305 310 315	

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Asn Ser Ser Asp Gly Thr Val Lys Val Tyr Arg Ile Thr Arg Glu Tyr					
acc aca aac gcc tac caa atg gat gtt gag ttg ttc cca ttc ggt ggt	340	345	350		1055
Thr Thr Asn Ala Tyr Gln Met Asp Val Glu Leu Phe Pro Phe Gly Gly					
gag aac tac aga ctt gac tac aag ttc aag aac ttc tac aac gcc tcc	355	360	365		1103
Glu Asn Tyr Arg Leu Asp Tyr Lys Phe Lys Asn Phe Tyr Asn Ala Ser					
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Tyr Leu Ser Ile Lys Leu Asn Asp Lys Arg Glu Leu Val Arg Thr Glu					
ggt gct cct caa gtg aac att gag tac tct gcc aac atc acc ctc aac	385	390	395		1199
Gly Ala Pro Gln Val Asn Ile Glu Tyr Ser Ala Asn Ile Thr Leu Asn					
aca gct gac atc tct caa cca ttc gag att ggt ttg acc aga gtc ctt	400	405	410	415	1247
Thr Ala Asp Ile Ser Gln Pro Phe Glu Ile Gly Leu Thr Arg Val Leu					
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Pro Ser Gly Ser Trp Ala Tyr Ala Ala Lys Phe Thr Val Glu Glu					
tac aac cag tac tct ttc ctc ttg aag ctc aac aag gca att cgt ctc	435	440	445		1343
Tyr Asn Gln Tyr Ser Phe Leu Leu Lys Leu Asn Lys Ala Ile Arg Leu					
agc aga gcc act gag ttg tct ccc acc atc ttg gag ggc att gtg agg	450	455	460		1391
Ser Arg Ala Thr Glu Leu Ser Pro Thr Ile Leu Glu Gly Ile Val Arg					
tct gtc aac ctt caa ctt gac atc aac act gat gtg ctt ggc aag gtc	465	470	475		1439
Ser Val Asn Leu Gln Leu Asp Ile Asn Thr Asp Val Leu Gly Lys Val					
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Phe Leu Thr Lys Tyr Tyr Met Gln Arg Tyr Ala Ile His Ala Glu Thr					
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Ala Leu Ile Leu Cys Asn Ala Pro Ile Ser Gln Arg Ser Tyr Asp Asn					
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Gln Pro Ser Gln Phe Asp Arg Leu Phe Asn Thr Pro Leu Leu Asn Gly					
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Gln Tyr Phe Ser Thr Gly Asp Glu Glu Ile Asp Leu Asn Ser Gly Ser					
aca ggt gac tgg aga aag acc atc ttg aag agg gcc ttc aac att gat	545	550	555		1679
Thr Gly Asp Trp Arg Lys Thr Ile Leu Lys Arg Ala Phe Asn Ile Asp					
gat gtc tct ctc ttc cgt ctc ttg aag atc aca gat cac gac aac aag					1727

Asp Val Ser Leu Phe Arg Leu Leu Lys Ile Thr Asp His Asp Asn Lys			
560	565	570	575
gat ggc aag atc aag aac aac ttg aag aac ctt tcc aac ctc tac att			1775
Asp Gly Lys Ile Lys Asn Asn Leu Lys Asn Leu Ser Asn Leu Tyr Ile			
580	585	590	
ggc aag ttg ctt gca gac atc cac caa ctc acc att gat gag ttg gac			1823
Gly Lys Leu Leu Ala Asp Ile His Gln Leu Thr Ile Asp Glu Leu Asp			
595	600	605	
ctc ttg ctc att gca gtc ggt gag ggc aag acc aac ctc tct gca atc			1871
Leu Leu Leu Ile Ala Val Gly Glu Gly Lys Thr Asn Leu Ser Ala Ile			
610	615	620	
tct gac aag cag ttg gca acc ctc atc agg aag ttg aac acc atc acc			1919
Ser Asp Lys Gln Leu Ala Thr Leu Ile Arg Lys Leu Asn Thr Ile Thr			
625	630	635	
tcc tgg ctt cac acc cag aag tgg tct gtc ttc caa ctc ttc atc atg			1967
Ser Trp Leu His Thr Gln Lys Trp Ser Val Phe Gln Leu Phe Ile Met			
640	645	650	655
acc agc acc tcc tac aac aag acc ctc act cct gag atc aag aac ctc			2015
Thr Ser Thr Ser Tyr Asn Lys Thr Leu Thr Pro Glu Ile Lys Asn Leu			
660	665	670	
ttg gac aca gtc tac cac ggt ctc caa ggc ttc gac aag gac aag gct			2063
Leu Asp Thr Val Tyr His Gly Leu Gln Gly Phe Asp Lys Asp Lys Ala			
675	680	685	
gac ttg ctt cat gtc atg gct ccc tac att gca gcc acc ctc caa ctc			2111
Asp Leu Leu His Val Met Ala Pro Tyr Ile Ala Ala Thr Leu Gln Leu			
690	695	700	
tcc tct gag aac gtg gct cac tct gtc ttg ctc tgg gct gac aag ctc			2159
Ser Ser Glu Asn Val Ala His Ser Val Leu Leu Trp Ala Asp Lys Leu			
705	710	715	
caa cct ggt gat ggt gcc atg act gct gag aag ttc tgg gac tgg ctc			2207
Gln Pro Gly Asp Gly Ala Met Thr Ala Glu Lys Phe Trp Asp Trp Leu			
720	725	730	735
aac acc aag tac aca cca ggc tcc tct gag gct gtt gag act caa gag			2255
Asn Thr Lys Tyr Thr Pro Gly Ser Ser Glu Ala Val Glu Thr Gln Glu			
740	745	750	
cac att gtg caa tac tgc cag gct ctt gca cag ttg gag atg gtc tac			2303
His Ile Val Gln Tyr Cys Gln Ala Leu Ala Gln Leu Glu Met Val Tyr			
755	760	765	
cac tcc act ggc atc aac gag aac gct ttc aga ctc ttc gtc acc aag			2351
His Ser Thr Gly Ile Asn Glu Asn Ala Phe Arg Leu Phe Val Thr Lys			
770	775	780	
cct gag atg ttc ggt gct gcc aca ggt gct gca cct gct cat gat gct			2399
Pro Glu Met Phe Gly Ala Ala Thr Gly Ala Ala Pro Ala His Asp Ala			
785	790	795	
ctc tcc ctc atc atg ttg acc agg ttc gct gac tgg gtc aac gct ctt			2447
Leu Ser L u Ile Met Leu Thr Arg Phe Ala Asp Trp Val Asn Ala Leu			

800	805	810	815			
ggt gag aag gct tcc tct gtc ttg gct gcc ttc gag gcc aac tcc ctc				2495		
Gly	Glu	Lys	Ala	Ser Ser Val Leu Ala Ala Phe Glu Ala Asn Ser Leu		
			820	825	830	
act gct gag caa ctt gct gat gcc atg aac ctt gat gcc aac ctc ttg				2543		
Thr	Ala	Glu	Gln	Leu Ala Asp Ala Met Asn Leu Asp Ala Asn Leu Leu		
			835	840	845	
ctc caa gct tcc att caa gct cag aac cac caa cac ctc cca cct gtc				2591		
Leu	Gln	Ala	Ser Ile	Gln Ala Gln Asn His Gln His Leu Pro Pro Val		
			850	855	860	
act cca gag aac gct ttc tcc tgc tgg acc tcc atc aac acc atc ctc				2639		
Thr	Pro	Glu	Asn Ala	Phe Ser Cys Trp Thr Ser Ile Asn Thr Ile Leu		
			865	870	875	
caa tgg gtc aac gtg gct cag caa ctc aac gtg gct cca caa ggt gtc				2687		
Gln	Trp	Val	Asn Val	Ala Gln Gln Leu Asn Val Ala Pro Gln Gly Val		
			880	885	890	895
tct gct ttg gtc ggt ctt gac tac atc cag tcc atg aag gag aca cca				2735		
Ser	Ala	Leu	Val	Gly Leu Asp Tyr Ile Gln Ser Met Lys Glu Thr Pro		
			900	905	910	
acc tac gct caa tgg gag aac gca gct ggt gtc ttg act gct ggt ctc				2783		
Thr	Tyr	Ala	Gln	Trp Glu Asn Ala Ala Gly Val Leu Thr Ala Gly Leu		
			915	920	925	
aac tcc caa cag gcc aac acc ctc cat gct ttc ttg gat gag tct cgc				2831		
Asn	Ser	Gln	Gln	Ala Asn Thr Leu His Ala Phe Leu Asp Glu Ser Arg		
			930	935	940	
tct gct gcc ctc tcc acc tac tac atc agg caa gtc gcc aag gca gct				2879		
Ser	Ala	Ala	Leu	Ser Thr Tyr Tyr Ile Arg Gln Val Ala Lys Ala Ala		
			945	950	955	
gct gcc atc aag tct cgc gat gac ctc tac caa tac ctc ctc att gac				2927		
Ala	Ala	Ile	Lys	Ser Arg Asp Asp Leu Tyr Gln Tyr Leu Leu Ile Asp		
			960	965	970	975
aac cag gtc tct gct gcc atc aag acc acc agg atc gct gag gcc atc				2975		
Asn	Gln	Val	Ser	Ala Ala Ile Lys Thr Thr Arg Ile Ala Glu Ala Ile		
			980	985	990	
gct tcc atc caa ctc tac gtc aac cgc gct ctt gag aac gtt gag gag				3023		
Ala	Ser	Ile	Gln	Leu Tyr Val Asn Arg Ala Leu Glu Asn Val Glu Glu		
			995	1000	1005	
aac gcc aac tct ggt gtc atc tct cgc caa ttc ttc atc gac tgg gac				3071		
Asn	Ala	Asn	Ser	Gly Val Ile Ser Arg Gln Phe Phe Ile Asp Trp Asp		
			1010	1015	1020	
aag tac aac aag agg tac tcc acc tgg gct ggt gtc tct caa ctt gtc				3119		
Lys	Tyr	Asn	Lys	Arg Tyr Ser Thr Trp Ala Gly Val Ser Gln Leu Val		
			1025	1030	1035	
tac tac cca gag aac tac att gac cca acc atg agg att ggt cag acc				3167		
Tyr	Tyr	Pro	Glu	Asn Tyr Ile Asp Pro Thr Met Arg Ile Gly Gln Thr		
			1040	1045	1050	1055

aag atg atg gat gct ctc ttg caa tct gtc tcc caa agc caa ctc aac	3215
Lys Met Met Asp Ala Leu Leu Gln Ser Val Ser Gln Ser Gln Leu Asn	
1060 1065 1070	
gct gac act gtg gag gat gcc ttc atg agc tac ctc acc tcc ttc gag	3263
Ala Asp Thr Val Glu Asp Ala Phe Met Ser Tyr Leu Thr Ser Phe Glu	
1075 1080 1085	
caa gtt gcc aac ctc aag gtc atc tct gct tac cat gac aac atc aac	3311
Gln Val Ala Asn Leu Lys Val Ile Ser Ala Tyr His Asp Asn Ile Asn	
1090 1095 1100	
aac gac caa ggt ctc acc tac ttc att ggt ctc tct gag act gat gct	3359
Asn Asp Gln Gly Leu Thr Tyr Phe Ile Gly Leu Ser Glu Thr Asp Ala	
1105 1110 1115	
ggt gag tac tac tgg aga tcc gtg gac cac agc aag ttc aac gat ggc	3407
Gly Glu Tyr Tyr Trp Arg Ser Val Asp His Ser Lys Phe Asn Asp Gly	
1120 1125 1130 1135	
aag ttc gct gca aac gct tgg tct gag tgg cac aag att gac tgc cct	3455
Lys Phe Ala Ala Asn Ala Trp Ser Glu Trp His Lys Ile Asp Cys Pro	
1140 1145 1150	
atc aac cca tac aag tcc acc atc aga cct gtc atc tac aag agc cgc	3503
Ile Asn Pro Tyr Lys Ser Thr Ile Arg Pro Val Ile Tyr Lys Ser Arg	
1155 1160 1165	
ctc tac ttg ctc tgg ctt gag cag aag gag atc acc aag caa act ggc	3551
Leu Tyr Leu Leu Trp Leu Glu Gln Lys Glu Ile Thr Lys Gln Thr Gly	
1170 1175 1180	
aac tcc aag gat ggt tac caa act gag act gac tac cgc tac gag ttg	3599
Asn Ser Lys Asp Gly Tyr Gln Thr Glu Thr Asp Tyr Arg Tyr Glu Leu	
1185 1190 1195	
aag ttg gct cac atc cgc tac gat ggt acc tgg aac act cca atc acc	3647
Lys Leu Ala His Ile Arg Tyr Asp Gly Thr Trp Asn Thr Pro Ile Thr	
1200 1205 1210 1215	
. ttc gat gtc aac aag aag atc agc gag ttg aag ttg gag aag aac cgt	3695
Phe Asp Val Asn Lys Lys Ile Ser Glu Leu Lys Leu Glu Lys Asn Arg	
1220 1225 1230	
gct cct ggt ctc tac tgc gct ggt tac caa ggt gag gac acc ctc ttg	3743
Ala Pro Gly Leu Tyr Cys Ala Gly Tyr Gln Gly Glu Asp Thr Leu Leu	
1235 1240 1245	
gtc atg ttc tac aac cag caa gac acc ctt gac tcc tac aag aac gct	3791
Val Met Phe Tyr Asn Gln Gln Asp Thr Leu Asp Ser Tyr Lys Asn Ala	
1250 1255 1260	
tcc atg caa ggt ctc tac atc ttc gct gac atg gct tcc aag gac atg	3839
Ser Met Gln Gly Leu Tyr Ile Phe Ala Asp Met Ala Ser Lys Asp Met	
1265 1270 1275	
act cca gag caa agc aac gtc tac cgt gac aac tcc tac caa cag ttc	3887
Thr Pro Glu Gln Ser Asn Val Tyr Arg Asp Asn Ser Tyr Gln Gln Phe	
1280 1285 1290 1295	

gac acc aac aac gtc agg cgt gtc aac aac aga tac gct gag gac tac 3935  
 Asp Thr Asn Asn Val Arg Arg Val Asn Asn Arg Tyr Ala Glu Asp Tyr  
 1300 1305 1310  
  
 gag atc cca agc tct gtc agc tct cgc aag gac tac ggc tgg ggt gac 3983  
 Glu Ile Pro Ser Ser Val Ser Arg Lys Asp Tyr Gly Trp Gly Asp  
 1315 1320 1325  
  
 tac tac ctc agc atg gtg tac aac ggt gac atc cca acc atc aac tac 4031  
 Tyr Tyr Leu Ser Met Val Tyr Asn Gly Asp Ile Pro Thr Ile Asn Tyr  
 1330 1335 1340  
  
 aag gct gcc tct tcc gac ctc aaa atc tac atc agc cca aag ctc agg 4079  
 Lys Ala Ala Ser Ser Asp Leu Lys Ile Tyr Ile Ser Pro Lys Leu Arg  
 1345 1350 1355  
  
 atc atc cac aac ggc tac gag ggt cag aag agg aac cag tgc aac ttg 4127  
 Ile Ile His Asn Gly Tyr Glu Gly Gln Lys Arg Asn Gln Cys Asn Leu  
 1360 1365 1370 1375  
  
 atg aac aag tac ggc aag ttg ggt gac aag ttc att gtc tac acc tct 4175  
 Met Asn Lys Tyr Gly Lys Leu Gly Asp Lys Phe Ile Val Tyr Thr Ser  
 1380 1385 1390  
  
 ctt ggt gtc aac cca aac aac agc tcc aac aag ctc atg ttc tac cca 4223  
 Leu Gly Val Asn Pro Asn Asn Ser Ser Asn Lys Leu Met Phe Tyr Pro  
 1395 1400 1405  
  
 gtc tac caa tac tct ggc aac acc tct ggt ctc aac cag ggt aga ctc 4271  
 Val Tyr Gln Tyr Ser Gly Asn Thr Ser Gly Leu Asn Gln Gly Arg Leu  
 1410 1415 1420  
  
 ttg ttc cac agg gac acc acc tac cca agc aag gtg gag gct tgg att 4319  
 Leu Phe His Arg Asp Thr Thr Tyr Pro Ser Lys Val Glu Ala Trp Ile  
 1425 1430 1435  
  
 cct ggt gcc aag agg tcc ctc acc aac cag aac gct gcc att ggt gat 4367  
 Pro Gly Ala Lys Arg Ser Leu Thr Asn Gln Asn Ala Ala Ile Gly Asp  
 1440 1445 1450 1455  
  
 gac tac gcc aca gac tcc ctc aac aag cct gat gac ctc aag cag tac 4415  
 Asp Tyr Ala Thr Asp Ser Leu Asn Lys Pro Asp Asp Leu Lys Gln Tyr  
 1460 1465 1470  
  
 atc ttc atg act gac tcc aag ggc aca gcc act gat gtc tct ggt cca 4463  
 Ile Phe Met Thr Asp Ser Lys Gly Thr Ala Thr Asp Val Ser Gly Pro  
 1475 1480 1485  
  
 gtg gag atc aac act gca atc agc cca gcc aag gtc caa atc att gtc 4511  
 Val Glu Ile Asn Thr Ala Ile Ser Pro Ala Lys Val Gln Ile Ile Val  
 1490 1495 1500  
  
 aag gct ggt ggc aag gag caa acc ttc aca gct gac aag gat gtc tcc 4559  
 Lys Ala Gly Gly Lys Glu Gln Thr Phe Thr Ala Asp Lys Asp Val Ser  
 1505 1510 1515  
  
 atc cag cca agc cca tcc ttc gat gag atg aac tac caa ttc aac gct 4607  
 Ile Gln Pro Ser Pro Ser Phe Asp Glu Met Asn Tyr Gln Phe Asn Ala  
 1520 1525 1530 1535  
  
 ctt gag att gat ggt tct ggc ctc aac ttc atc aac aac tct gtc tcc 4655

Leu Glu Ile Asp Gly Ser Gly L u Asn Phe Ile Asn Asn Ser Ala Ser  
 1540 1545 1550  
 att gat gtc acc ttc act gcc ttc gct gag gat ggc cgc aag ttg ggt 4703  
 Ile Asp Val Thr Phe Thr Ala Phe Ala Glu Asp Gly Arg Lys Leu Gly  
 1555 1560 1565  
 tac gag agc ttc tcc atc cca gtc acc ctt aag gtt tcc act gac aac 4751  
 Tyr Glu Ser Phe Ser Ile Pro Val Thr Leu Lys Val Ser Thr Asp Asn  
 1570 1575 1580  
 gca ctc acc ctt cat cac aac gag aac ggt gct cag tac atg caa tgg 4799  
 Ala Leu Thr Leu His His Asn Glu Asn Gly Ala Gln Tyr Met Gln Trp  
 1585 1590 1595  
 caa agc tac cgc acc agg ttg aac acc ctc ttc gca agg caa ctt gtg 4847  
 Gln Ser Tyr Arg Thr Arg Leu Asn Thr Leu Phe Ala Arg Gln Leu Val  
 1600 1605 1610 1615  
 gcc cgt gcc acc aca ggc att gac acc atc ctc agc atg gag acc cag 4895  
 Ala Arg Ala Thr Thr Gly Ile Asp Thr Ile Leu Ser Met Glu Thr Gln  
 1620 1625 1630  
 aac atc caa gag cca cag ttg ggc aag ggt ttc tac gcc acc ttc gtc 4943  
 Asn Ile Gln Glu Pro Gln Leu Gly Lys Gly Phe Tyr Ala Thr Phe Val  
 1635 1640 1645  
 atc cca cct tac aac ctc agc act cat ggt gat gag agg tgg ttc aag 4991  
 Ile Pro Pro Tyr Asn Leu Ser Thr His Gly Asp Glu Arg Trp Phe Lys  
 1650 1655 1660  
 ctc tac atc aag cac gtg gtt gac aac aac tcc cac atc atc tac tct 5039  
 Leu Tyr Ile Lys His Val Val Asp Asn Asn Ser His Ile Ile Tyr Ser  
 1665 1670 1675  
 ggt caa ctc act gac acc aac atc aac atc acc ctc ttc atc cca ctt 5087  
 Gly Gln Leu Thr Asp Thr Asn Ile Asn Ile Thr Leu Phe Ile Pro Leu  
 1680 1685 1690 1695  
 gac gat gtc cca ctc aac cag gac tac cat gcc aag gtc tac atg acc 5135  
 Asp Asp Val Pro Leu Asn Gln Asp Tyr His Ala Lys Val Tyr Met Thr  
 1700 1705 1710  
 ttc aag aag tct cca tct gat ggc acc tgg tgg ggt cca cac ttc gtc 5183  
 Phe Lys Lys Ser Pro Ser Asp Gly Thr Trp Trp Gly Pro His Phe Val  
 1715 1720 1725  
 cgt gat gac aag ggc atc gtc acc atc aac cca aag tcc atc ctc acc 5231  
 Arg Asp Asp Lys Gly Ile Val Thr Ile Asn Pro Lys Ser Ile Leu Thr  
 1730 1735 1740  
 cac ttc gag tct gtc aac gtt ctc aac aac atc tcc tct gag cca atg 5279  
 His Phe Glu Ser Val Asn Val Leu Asn Asn Ile Ser Ser Glu Pro Met  
 1745 1750 1755  
 gac ttc tct ggt gcc aac tcc ctc tac ttc tgg gag ttg ttc tac tac 5327  
 Asp Phe Ser Gly Ala Asn Ser Leu Tyr Phe Trp Glu Leu Phe Tyr Tyr  
 1760 1765 1770 1775  
 aca cca atg ctt gtg gct caa agg ttg ctc cat gag cag aac ttc gat 5375  
 Thr Pro Met Leu Val Ala Gln Arg Leu Leu His Glu Gln Asn Phe Asp

1780	1785	1790	
gag gcc aac agg tgg ctc aag tac gtc tgg agc cca tct ggt tac att Glu Ala Asn Arg Trp Leu Lys Tyr Val Trp Ser Pro Ser Gly Tyr Ile			5423
1795	1800	1805	
gtg cat ggt caa atc cag aac tac caa tgg aac gtc agg cca ttg ctt Val His Gly Gln Ile Gln Asn Tyr Gln Trp Asn Val Arg Pro Leu Leu			5471
1810	1815	1820	
gag gac acc tcc tgg aac tct gac cca ctt gac tct gtg gac cct gat Glu Asp Thr Ser Trp Asn Ser Asp Pro Leu Asp Ser Val Asp Pro Asp			5519
1825	1830	1835	
gct gtg gct caa cat gac cca atg cac tac aag gtc tcc acc ttc atg Ala Val Ala Gln His Asp Pro Met His Tyr Lys Val Ser Thr Phe Met			5567
1840	1845	1850	1855
agg acc ttg gac ctc ttg att gcc aga ggt gac cat gct tac cgc caa Arg Thr Leu Asp Leu Leu Ile Ala Arg Gly Asp His Ala Tyr Arg Gln			5615
1860	1865	1870	
ttg gag agg gac acc ctc aac gag gca aag atg tgg tac atg caa gct Leu Glu Arg Asp Thr Leu Asn Glu Ala Lys Met Trp Tyr Met Gln Ala			5663
1875	1880	1885	
ctc cac ctc ttg ggt gac aag cca tac ctc cca ctc agc acc act tgg Leu His Leu Leu Gly Asp Lys Pro Tyr Leu Pro Leu Ser Thr Thr Trp			5711
1890	1895	1900	
tcc gac cca agg ttg gac cgt gct gac atc acc act cag aac gct Ser Asp Pro Arg Leu Asp Arg Ala Ala Asp Ile Thr Thr Gln Asn Ala			5759
1905	1910	1915	
cat gac tct gcc att gtt gct ctc agg cag aac atc cca act cct gct His Asp Ser Ala Ile Val Ala Leu Arg Gln Asn Ile Pro Thr Pro Ala			5807
1920	1925	1930	1935
cca ctc tcc ctc aga tct gct aac acc ctc act gac ttg ttc ctc cca Pro Leu Ser Leu Arg Ser Ala Asn Thr Leu Thr Asp Leu Phe Leu Pro			5855
1940	1945	1950	
cag atc aac gag gtc atg atg aac tac tgg caa acc ttg gct caa agg Gln Ile Asn Glu Val Met Met Asn Tyr Trp Gln Thr Leu Ala Gln Arg			5903
1955	1960	1965	
gtc tac aac ctc aga cac aac ctc tcc att gat ggt caa cca ctc tac Val Tyr Asn Leu Arg His Asn Leu Ser Ile Asp Gly Gln Pro Leu Tyr			5951
1970	1975	1980	
ctc cca atc tac gcc aca cca gct gac cca aag gct ctt ctc tct gct Leu Pro Ile Tyr Ala Thr Pro Ala Asp Pro Lys Ala Leu Leu Ser Ala			5999
1985	1990	1995	
gct gtg gct acc agc caa ggt ggt ggc aag ctc cca gag tcc ttc atg Ala Val Ala Thr Ser Gln Gly Gly Lys Leu Pro Glu Ser Phe Met			6047
2000	2005	2010	2015
tcc ctc tgg agg ttc cca cac atg ttg gag aac gcc cgt ggc atg gtc Ser Leu Trp Arg Phe Pro His Met Leu Glu Asn Ala Arg Gly Met Val			6095
2020	2025	2030	

tcc caa ctc acc cag ttc ggt tcc acc ctc cag aac atc att gag agg	6143
Ser Gln Leu Thr Gln Phe Gly S r Thr Leu Gln Asn Ile Ile Glu Arg	
2035 2040 2045	
caa gat gct gag gct ctc aac gct ttg ctc cag aac cag gca gct gag	6191
Gln Asp Ala Glu Ala Leu Asn Ala Leu Leu Gln Asn Gln Ala Ala Glu	
2050 2055 2060	
ttg atc ctc acc aac ttg tcc atc caa gac aag acc att gag gag ctt	6239
Leu Ile Leu Thr Asn Leu Ser Ile Gln Asp Lys Thr Ile Glu Glu Leu	
2065 2070 2075	
gat gct gag aag aca gtc ctt gag aag agc aag gct ggt gcc caa tct	6287
Asp Ala Glu Lys Thr Val Leu Glu Lys Ser Lys Ala Gly Ala Gln Ser	
2080 2085 2090 2095	
cgc ttc gac tcc tac ggc aag ctc tac gat gag aac atc aac gct ggt	6335
Arg Phe Asp Ser Tyr Gly Lys Leu Tyr Asp Glu Asn Ile Asn Ala Gly	
2100 2105 2110	
gag aac cag gcc atg acc ctc agg gct tcc gca gct ggt ctc acc act	6383
Glu Asn Gln Ala Met Thr Leu Arg Ala Ser Ala Ala Gly Leu Thr Thr	
2115 2120 2125	
gct gtc caa gcc tct cgc ttg gct gca gct gct gac ctc gtt cca	6431
Ala Val Gln Ala Ser Arg Leu Ala Gly Ala Ala Asp Leu Val Pro	
2130 2135 2140	
aac atc ttc ggt ttc gct ggt ggt ggc tcc aga tgg ggt gcc att gct	6479
Asn Ile Phe Gly Phe Ala Gly Gly Ser Arg Trp Gly Ala Ile Ala	
2145 2150 2155	
gag gct acc ggt tac gtc atg gag ttc tct gcc aac gtc atg aac act	6527
Glu Ala Thr Gly Tyr Val Met Glu Phe Ser Ala Asn Val Met Asn Thr	
2160 2165 2170 2175	
gag gct gac aag atc agc caa tct gag acc tac aga agg cgc cgt caa	6575
Glu Ala Asp Lys Ile Ser Gln Ser Glu Thr Tyr Arg Arg Arg Gln	
2180 2185 2190	
gag tgg gag atc caa agg aac aac gct gag gca gag ttg aag caa atc	6623
Glu Trp Glu Ile Gln Arg Asn Asn Ala Glu Ala Glu Leu Lys Gln Ile	
2195 2200 2205	
gat gct caa ctc aag tcc ttg gct gtc aga agg gag gct gct gtc ctc	6671
Asp Ala Gln Leu Lys Ser Leu Ala Val Arg Arg Glu Ala Ala Val Leu	
2210 2215 2220	
cag aag acc tcc ctc aag acc caa cag gag caa acc cag tcc cag ttg	6719
Gln Lys Thr Ser Leu Lys Thr Gln Gln Glu Gln Thr Gln Ser Gln Leu	
2225 2230 2235	
gct ttc ctc caa agg aag ttc tcc aac cag gct ctc tac aac tgg ctc	6767
Ala Phe Leu Gln Arg Lys Phe Ser Asn Gln Ala Leu Tyr Asn Trp Leu	
2240 2245 2250 2255	
aga ggc cgc ttg gct gcc atc tac ttc caa ttc tac gac ctt gct gtg	6815
Arg Gly Arg Leu Ala Ala Ile Tyr Phe Gln Phe Tyr Asp Leu Ala Val	
2260 2265 2270	

gcc agg tgc ctc atg gct gag caa gcc tac cgc tgg gag ttg aac gat Ala Arg Cys Leu Met Ala Glu Gln Ala Tyr Arg Trp Glu Leu Asn Asp 2275 2280 2285	6863
gac tcc gcc agg ttc atc aag cca ggt gct tgg caa ggc acc tac gct Asp Ser Ala Arg Phe Ile Lys Pro Gly Ala Trp Gln Gly Thr Tyr Ala 2290 2295 2300	6911
ggt ctc ctt gct ggt gag acc ctc atg ctc tcc ttg gct caa atg gag Gly Leu Leu Ala Gly Glu Thr Leu Met Leu Ser Leu Ala Gln Met Glu 2305 2310 2315	6959
gat gct cac ctc aag agg gac aag agg gct ttg gag gtg gag agg aca Asp Ala His Leu Lys Arg Asp Lys Arg Ala Leu Glu Val Glu Arg Thr 2320 2325 2330 2335	7007
gtc tcc ctt gct gag gtc tac gct ggt ctc cca aag gac aac ggt cca Val Ser Leu Ala Glu Val Tyr Ala Gly Leu Pro Lys Asp Asn Gly Pro 2340 2345 2350	7055
ttc tcc ctt gct caa gag att gac aag ttg gtc agc caa ggt tct ggt Phe Ser Leu Ala Gln Glu Ile Asp Lys Leu Val Ser Gln Gly Ser Gly 2355 2360 2365	7103
tct gct ggt tct ggt aac aac aac ttg gct ttc ggc gct ggt act gac Ser Ala Gly Ser Gly Asn Asn Asn Leu Ala Phe Gly Ala Gly Thr Asp 2370 2375 2380	7151
acc aag acc tcc ctc caa gcc tct gtc tcc ttc gct gac ctc aag atc Thr Lys Thr Ser Leu Gln Ala Ser Val Ser Phe Ala Asp Leu Lys Ile 2385 2390 2395	7199
agg gag gac tac cca gct tcc ctt ggc aag atc agg cgc atc aag caa Arg Glu Asp Tyr Pro Ala Ser Leu Gly Lys Ile Arg Arg Ile Lys Gln 2400 2405 2410 2415	7247
atc tct gtc acc ctc cca gct ctc ttg ggt cca tac caa gat gtc caa Ile Ser Val Thr Leu Pro Ala Leu Leu Gly Pro Tyr Gln Asp Val Gln 2420 2425 2430	7295
gca atc ctc tcc tac ggt gac aag gct ggt ttg gcg aac ggt tgc gag Ala Ile Leu Ser Tyr Gly Asp Lys Ala Gly Leu Ala Asn Gly Cys Glu 2435 2440 2445	7343
gct ctt gct gtc tct cat ggc atg aac gac tct ggt caa ttc caa ctt Ala Leu Ala Val Ser His Gly Met Asn Asp Ser Gly Gln Phe Gln Leu 2450 2455 2460	7391
gac ttc aac gat ggc aag ttc ctc cca ttc gag ggc att gcc att gac Asp Phe Asn Asp Gly Lys Phe Leu Pro Phe Glu Gly Ile Ala Ile Asp 2465 2470 2475	7439
caa ggc acc ctc acc ctc tcc cca aac gct tcc atg cca gag aag Gln Gly Thr Leu Thr Leu Ser Phe Pro Asn Ala Ser Met Pro Glu Lys 2480 2485 2490 2495	7487
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Lys Leu Gln Leu Thr Cys Pro Ala Glu Ile Ala Leu Tyr Pro Phe Asp	
20 25 30	

acc ttc cgt gag aag acc aga ggc atg gtc aac tgg ggt gag gcc aag	143
Thr Phe Arg Glu Lys Thr Arg Gly Met Val Asn Trp Gly Glu Ala Lys	
35 40 45	

agg atc tac gag att gct caa gct gag caa gac agg aac ctc ctt cat	191
Arg Ile Tyr Glu Ile Ala Gln Ala Glu Gln Asp Arg Asn Leu Leu His	
50 55 60	

gag aag agg atc ttc gcc tac gct aac cca ttg ctc aag aac gct gtc	239
Glu Lys Arg Ile Phe Ala Tyr Ala Asn Pro Leu Leu Lys Asn Ala Val	
65 70 75	

agg ctt ggt acc agg caa atg ttg ggt ttc atc caa ggt tac tct gac	287
Arg Leu Gly Thr Arg Gln Met Leu Gly Phe Ile Gln Gly Tyr Ser Asp	
80 85 90 95	

ttg ttc ggc aac agg gct gac aac tac gca gct cct ggt tct gtt gct	335
Leu Phe Gly Asn Arg Ala Asp Asn Tyr Ala Ala Pro Gly Ser Val Ala	
100 105 110	

agc atg ttc agc cca gct gcc tac ctc act gag ttg tac cgt gag gcc	383
Ser Met Phe Ser Pro Ala Ala Tyr Leu Thr Glu Leu Tyr Arg Glu Ala	
115 120 125	

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Lys Asn Leu His Asp Ser Ser Ser Ile Tyr Tyr Leu Asp Lys Arg Arg	
130 135 140	

cca gac ctt gct tcc ttg atg ctc tcc cag aag aac atg gat gag gag	479
Pro Asp Leu Ala Ser Leu Met Leu Ser Gln Lys Asn Met Asp Glu Glu	
145 150 155	

atc agc acc ttg gct ctc tcc aac gag ctt tgc ttg gct ggc att gag	527
Ile Ser Thr Leu Ala Leu Ser Asn Glu Leu Cys Leu Ala Gly Ile Glu	
160 165 170 175	

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Thr Lys Thr Gly Lys Ser Gln Asp Glu Val Met Asp M t Leu Ser Thr	
180 185 190	
tac cgc ctc tct ggt gag act cca tac cac cat gct tac gag act gtc	623
Tyr Arg Leu Ser Gly Glu Thr Pro Tyr His His Ala Tyr Glu Thr Val	
195 200 205	
agg gag att gtc cat gag agg gac cca ggt ttc cgc cac ctc tcc caa	671
Arg Glu Ile Val His Glu Arg Asp Pro Gly Phe Arg His Leu Ser Gln	
210 215 220	
gct ccc att gtg gct gcc aag ttg gac cca gtc acc ctc ttg ggc atc	719
Ala Pro Ile Val Ala Ala Lys Leu Asp Pro Val Thr Leu Leu Gly Ile	
225 230 235	
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Ser Ser His Ile Ser Pro Glu Leu Tyr Asn Leu Leu Ile Glu Glu Ile	
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Pro Glu Lys Asp Glu Ala Ala Leu Asp Thr Leu Tyr Lys Thr Asn Phe	
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Gly Asp Ile Thr Thr Ala Gln Leu Met Ser Pro Ser Tyr Leu Ala Arg	
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Tyr Tyr Gly Val Ser Pro Glu Asp Ile Ala Tyr Val Thr Thr Ser Leu	
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Ser His Val Gly Tyr Ser Ser Asp Ile Leu Val Ile Pro Leu Val Asp	
305 310 315	
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Gly Val Gly Lys Met Glu Val Val Arg Val Thr Arg Thr Pro Ser Asp	
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aac tac acc tcc cag acc aac tac att gag ttg tac cca caa ggt ggt	1055
Asn Tyr Thr Ser Gln Thr Asn Tyr Ile Glu Leu Tyr Pro Gln Gly Gly	
340 345 350	
gac aac tac ctc atc aag tac aac ctc tcc aac tct ttc ggt ttg gat	1103
Asp Asn Tyr Leu Ile Lys Tyr Asn Leu Ser Asn Ser Phe Gly Leu Asp	
355 360 365	
gac ttc tac ctc cag tac aag gat ggt tct gct gac tgg act gag att	1151
Asp Phe Tyr Leu Gln Tyr Lys Asp Gly Ser Ala Asp Trp Thr Glu Ile	
370 375 380	
gct cac aac cca tac cca gac atg gtc atc aac cag aag tac gag tcc	1199
Ala His Asn Pro Tyr Pro Asp Met Val Ile Asn Gln Lys Tyr Glu Ser	
385 390 395	
caa gcc acc atc aag aga tct gac tct gac aac atc ctc tcc att ggt	1247
Gln Ala Thr Ile Lys Arg Ser Asp Ser Asp Asn Ile L u Ser Ile Gly	
400 405 410 415	
ctc caa agg tgg cac tct ggt tcc tac aac ttc gct gct gcc aac ttc	1295

Leu Gln Arg Trp His Ser Gly Ser Tyr Asn Phe Ala Ala Ala Asn Phe  
 420 425 430  
 aag att gac caa tac tct cca aag gct ttc ctc ttg aag atg aac aag 1343  
 Lys Ile Asp Gln Tyr Ser Pro Lys Ala Phe Leu Leu Lys Met Asn Lys  
 435 440 445  
 gcc atc agg ctc ttg aag gcc act ggt ctc tcc ttc gcc acc ctt gag 1391  
 Ala Ile Arg Leu Leu Lys Ala Thr Gly Leu Ser Phe Ala Thr Leu Glu  
 450 455 460  
 agg att gtg gac tct gtc aac tcc acc aag tcc atc act gtg gag gtc 1439  
 Arg Ile Val Asp Ser Val Asn Ser Thr Lys Ser Ile Thr Val Glu Val  
 465 470 475  
 ctc aac aag gtc tac aga gtc aag ttc tac att gac cgc tac ggc atc 1487  
 Leu Asn Lys Val Tyr Arg Val Lys Phe Tyr Ile Asp Arg Tyr Gly Ile  
 480 485 490 495  
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 Ser Glu Glu Thr Ala Ala Ile Leu Ala Asn Ile Asn Ile Ser Gln Gln  
 500 505 510  
 gct gtc ggc aac cag ctc tcc caa ttc gag caa ctc ttc aac cac cct 1583  
 Ala Val Gly Asn Gln Leu Ser Gln Phe Glu Gln Leu Phe Asn His Pro  
 515 520 525  
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 Pro Leu Asn Gly Ile Arg Tyr Glu Ile Ser Glu Asp Asn Ser Lys His  
 530 535 540  
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 Leu Pro Asn Pro Asp Leu Asn Leu Lys Pro Asp Ser Thr Gly Asp Asp  
 545 550 555  
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 Gln Arg Lys Ala Val Leu Lys Arg Ala Phe Gln Val Asn Ala Ser Glu  
 560 565 570 575  
 ctt tac caa atg ctc ttg atc act gac agg aag gag gat ggt gtc atc 1775  
 Leu Tyr Gln Met Leu Leu Ile Thr Asp Arg Lys Glu Asp Gly Val Ile  
 580 585 590  
 aag aac aac ttg gag aac ctc tct gac ctc tac ctt gtc tcc ctc ttg 1823  
 Lys Asn Asn Leu Glu Asn Leu Ser Asp Leu Tyr Leu Val Ser Leu Leu  
 595 600 605  
 gcc caa atc cac aac ttg acc att gct gag ttg aac atc ctc ttg gtc 1871  
 Ala Gln Ile His Asn Leu Thr Ile Ala Glu Leu Asn Ile Leu Leu Val  
 610 615 620  
 atc tgc ggt tac ggt gac acc aac atc tac caa atc act gac gac aac 1919  
 Ile Cys Gly Tyr Gly Asp Thr Asn Ile Tyr Gln Ile Thr Asp Asp Asn  
 625 630 635  
 ctt gcc aag att gtg gag acc ctc ttg tgg atc acc caa tgg ctc aag 1967  
 Leu Ala Lys Ile Val Glu Thr Leu Leu Trp Ile Thr Gln Trp Leu Lys  
 640 645 650 655  
 acc cag aag tgg act gtc aca gac ctc ttc ctc atg acc act gcc acc 2015  
 Thr Gln Lys Trp Thr Val Thr Asp Leu Ph Leu Met Thr Thr Ala Thr

660	665	670	
tac tcc acc act ctc act cca gag att tcc aac ctc act gcc acc ctc			2063
Tyr Ser Thr Thr Leu Thr Pro Glu Ile Ser Asn Leu Thr Ala Thr Leu			
675	680	685	
agc tcc acc ctc cac ggc aag gag tcc ctc att ggt gag gac ctc aag			2111
Ser Ser Thr Leu His Gly Lys Glu Ser Leu Ile Gly Glu Asp Leu Lys			
690	695	700	
agg gca atg gct cca tgc ttc acc tct gct ctc cac ctc acc tcc caa			2159
Arg Ala Met Ala Pro Cys Phe Thr Ser Ala Leu His Leu Thr Ser Gln			
705	710	715	
gag gtg gct tac gac ctc ctt ctc tgg att gac caa atc caa cca gct			2207
Glu Val Ala Tyr Asp Leu Leu Leu Trp Ile Asp Gln Ile Gln Pro Ala			
720	725	730	735
caa atc act gtg gat ggt ttc tgg gag gag gtc caa acc act cca acc			2255
Gln Ile Thr Val Asp Gly Phe Trp Glu Glu Val Gln Thr Thr Pro Thr			
740	745	750	
tcc ctc aag gtc atc acc ttc gct caa gtc ttg gct caa ctc tcc ctc			2303
Ser Leu Lys Val Ile Thr Phe Ala Gln Val Leu Ala Gln Leu Ser Leu			
755	760	765	
atc tac aga agg att ggt ctc tct gag act gag ttg tcc ctc att gtc			2351
Ile Tyr Arg Arg Ile Gly Leu Ser Glu Thr Glu Leu Ser Leu Ile Val			
770	775	780	
acc caa tcc agc ctc ttg gtc gct ggc aag tcc atc ctt gat cat ggt			2399
Thr Gln Ser Ser Leu Leu Val Ala Gly Lys Ser Ile Leu Asp His Gly			
785	790	795	
ctc ttg acc ctc atg gct ctt gag ggt ttc cac acc tgg gtc aac ggt			2447
Leu Leu Thr Leu Met Ala Leu Glu Gly Phe His Thr Trp Val Asn Gly			
800	805	810	815
ttg ggt caa cat gct tcc ctc atc ttg gct gca ctc aag gat ggt gct			2495
Leu Gly Gln His Ala Ser Leu Ile Leu Ala Ala Leu Lys Asp Gly Ala			
820	825	830	
ctc acc gtc acc gat gtg gct caa gcc atg aac aag gag gag tcc ctc			2543
Leu Thr Val Thr Asp Val Ala Gln Ala Met Asn Lys Glu Ser Leu			
835	840	845	
ttg caa atg gct gcc aac cag gtg gag aag gac ctc acc aag ctc acc			2591
Leu Gln Met Ala Ala Asn Gln Val Glu Lys Asp Leu Thr Lys Leu Thr			
850	855	860	
tcc tgg acc caa atc gat gcc atc ctc caa tgg ctc caa atg tcc tct			2639
Ser Trp Thr Gln Ile Asp Ala Ile Leu Gln Trp Leu Gln Met Ser Ser			
865	870	875	
gct ctt gct gtc agc cca ttg gac ctt gct ggc atg atg gct ctc aag			2687
Ala Leu Ala Val Ser Pro Leu Asp Leu Ala Gly Met Met Ala Leu Lys			
880	885	890	895
tac ggc att gat cac aac tac gct gcc tgg caa gca gct gcc gct gcc			2735
Tyr Gly Ile Asp His Asn Tyr Ala Ala Trp Gln Ala Ala Ala Ala			
900	905	910	

ctc atg gct gac cat gcc aac cag gct cag aag ttg gat gag acc Leu Met Ala Asp His Ala Asn Gln Ala Gln Lys Lys Leu Asp Glu Thr 915 920 925	2783
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gct gcc ggt gtc agg gac agg aac ggt ctc tac acc tac ctc ttg att Ala Ala Gly Val Arg Asp Arg Asn Gly Leu Tyr Thr Tyr Leu Leu Ile 945 950 955	2879
gac aac cag gtc tct gct gat gtc atc acc tcc aga att gct gag gcc Asp Asn Gln Val Ser Ala Asp Val Ile Thr Ser Arg Ile Ala Glu Ala 960 965 970 975	2927
att gct ggc atc caa ctc tac gtc aac agg gct ctc aac agg gat gag Ile Ala Gly Ile Gln Leu Tyr Val Asn Arg Ala Leu Asn Arg Asp Glu 980 985 990	2975
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gag agg tac aac aag agg tac tcc acc tgg gct ggt gtc tct gag ttg Glu Arg Tyr Asn Lys Arg Tyr Ser Thr Trp Ala Gly Val Ser Glu Leu 1010 1015 1020	3071
gtc tac tac cca gag aac tac gtg gac cca acc caa agg att ggt cag Val Tyr Tyr Pro Glu Asn Tyr Val Asp Pro Thr Gln Arg Ile Gly Gln 1025 1030 1035	3119
acc aag atg atg gat gct ttg ctc caa tcc atc aac cag tcc caa ctc Thr Lys Met Met Asp Ala Leu Leu Gln Ser Ile Asn Gln Ser Gln Leu 1040 1045 1050 1055	3167
aac gct gac act gtg gag gat gct ttc aag acc tac ctc acc tcc ttc Asn Ala Asp Thr Val Glu Asp Ala Phe Lys Thr Tyr Leu Thr Ser Phe 1060 1065 1070	3215
gag caa gtg gcc aac ctc aag gtc atc tct gct tac cat gac aac gtc Glu Gln Val Ala Asn Leu Lys Val Ile Ser Ala Tyr His Asp Asn Val 1075 1080 1085	3263
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cct ggc acc tac tac tgg agg tct gtg gac cac tcc aag tgc gag aac Pro Gly Thr Tyr Tyr Trp Arg Ser Val Asp His Ser Lys Cys Glu Asn 1105 1110 1115	3359
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Arg Leu Tyr Leu L u Trp L u Glu Gln Gln Ser Lys Lys S r Asp Asp	
1155 1160 1165	
ggc aag aca act atc tac cag tac aac ctc aag ttg gct cac atc cgc	3551
Gly Lys Thr Thr Ile Tyr Gln Tyr Asn Leu Lys Leu Ala His Ile Arg	
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Tyr Asp Gly Ser Trp Asn Thr Pro Phe Thr Phe Asp Val Thr Glu Lys	
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Val Lys Asn Tyr Thr Ser Ser Thr Asp Ala Ala Glu Ser Leu Gly Leu	
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Tyr Cys Thr Gly Tyr Gln Gly Glu Asp Thr Leu Leu Val Met Phe Tyr	
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Ser Met Gln Ser Ser Tyr Ser Ser Tyr Thr Asp Asn Asn Ala Pro Val	
1235 1240 1245	
act ggt ctc tac atc ttc gct gac atg tcc tct gac aac atg acc aac	3791
Thr Gly Leu Tyr Ile Phe Ala Asp Met Ser Ser Asp Asn Met Thr Asn	
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gct caa gcc acc aac tac tgg aac aac tcc tac cca caa ttc gac act	3839
Ala Gln Ala Thr Asn Tyr Trp Asn Asn Ser Tyr Pro Gln Phe Asp Thr	
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gtc atg gct gac cca gac tct gac aac aag gtc atc acc agg cgt	3887
Val Met Ala Asp Pro Asp Ser Asp Asn Lys Lys Val Ile Thr Arg Arg	
1280 1285 1290 1295	
gtc aac aac cgc tac gct gag gac tac gag atc cca agc tct gtc acc	3935
Val Asn Asn Arg Tyr Ala Glu Asp Tyr Glu Ile Pro Ser Ser Val Thr	
1300 1305 1310	
tcc aac agc aac tac tcc tgg ggt gac cac tcc ctc acc atg ctc tac	3983
Ser Asn Ser Asn Tyr Ser Trp Gly Asp His Ser Leu Thr Met Leu Tyr	
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Gly Gly Ser Val Pro Asn Ile Thr Phe Glu Ser Ala Ala Glu Asp Leu	
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agg ctc tcc acc aac atg gct ctc tcc atc att cac aac ggt tac gct	4079
Arg Leu Ser Thr Asn Met Ala Leu Ser Ile Ile His Asn Gly Tyr Ala	
1345 1350 1355	
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Gly Thr Arg Arg Ile Gln Cys Asn Leu Met Lys Gln Tyr Ala Ser Leu	
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Gly Asp Lys Phe Ile Ile Tyr Asp Ser Ser Phe Asp Asp Ala Asn Arg	
1380 1385 1390	
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Phe Asn Leu Val Pro Leu Phe Lys Phe Gly Lys Asp Glu Asn Ser Asp			
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gac tcc atc tgc atc tac aac gag aac cca agc tct gag gac aag aag			4271
Asp Ser Ile Cys Ile Tyr Asn Glu Asn Pro S r Ser Glu Asp Lys Lys			
1410	1415	1420	
tgg tac ttc agc tcc aag gac gac aac aag act gct gac tac aac ggt			4319
Trp Tyr Phe Ser Ser Lys Asp Asp Asn Lys Thr Ala Asp Tyr Asn Gly			
1425	1430	1435	
ggc acc caa tgc att gat gct ggc acc tcc aac aag gac ttc tac tac			4367
Gly Thr Gln Cys Ile Asp Ala Gly Thr Ser Asn Lys Asp Phe Tyr Tyr			
1440	1445	1450	1455
aac ctc caa gag att gag gtc atc tct gtc act ggt ggc tac tgg tcc			4415
Asn Leu Gln Glu Ile Glu Val Ile Ser Val Thr Gly Gly Tyr Trp Ser			
1460	1465	1470	
agc tac aag atc agc aac ccc atc aac atc aac act ggc att gac tct			4463
Ser Tyr Lys Ile Ser Asn Pro Ile Asn Ile Asn Thr Gly Ile Asp Ser			
1475	1480	1485	
gcc aag gtc aag gtc act gtc aag gct ggt ggc gat gac caa atc ttc			4511
Ala Lys Val Lys Val Thr Val Lys Ala Gly Gly Asp Asp Gln Ile Phe			
1490	1495	1500	
act gct gac aac tcc acc tac gtc cca cag caa cct gct cca tcc ttc			4559
Thr Ala Asp Asn Ser Thr Tyr Val Pro Gln Gln Pro Ala Pro Ser Phe			
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Glu Glu Met Ile Tyr Gln Phe Asn Asn Leu Thr Ile Asp Cys Lys Asn			
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Leu Asn Phe Ile Asp Asn Gln Ala His Ile Glu Ile Asp Phe Thr Ala			
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Thr Ala Gln Asp Gly Arg Phe Leu Gly Ala Glu Thr Phe Ile Ile Pro			
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Val Thr Lys Lys Val Leu Gly Thr Glu Asn Val Ile Ala Leu Tyr Ser			
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1585	1590	1595	
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Ile Asp Ala Val Leu Ser Met Glu Thr Gln Asn Ile Gln Glu Pro Gln			
1620	1625	1630	
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Leu Gly Ala Gly Thr Tyr Val Gln Leu Val L u Asp Lys Tyr Asp Glu			

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ttc aag gag aac gac tcc ttc gtc atc tac caa ggt gag ttg tct gag Phe Lys Glu Asn Asp Ser Phe Val Ile Tyr Gln Gly Glu Leu Ser Glu 1665	1670	1675	5039
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acc act gac aag atc ctc ttc gac agg act gat gag aag gac cca cat Thr Thr Asp Lys Ile Leu Phe Asp Arg Thr Asp Glu Lys Asp Pro His 1715	1720	1725	5183
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gcc atc tac cac tgg aac gtc agg cca ttg gag gag gac acc tcc tgg Ala Ile Tyr His Trp Asn Val Arg Pro Leu Glu Glu Asp Thr Ser Trp 1810	1815	1820	5471
aac gct cag caa ctt gac tcc act gac cca gat gct gtg gct caa gat Asn Ala Gln Gln Leu Asp Ser Thr Asp Pro Asp Ala Val Ala Gln Asp 1825	1830	1835	5519
gac cca atg cac tac aag gtg gcc acc ttc atg gcc acc ttg gac ctt Asp Pro Met His Tyr Lys Val Ala Thr Phe Met Ala Thr Leu Asp Leu 1840	1845	1850	5567
ctc atg gcc aga ggt gat gct gtc tac cgc caa ttg gag agg gac acc Leu Met Ala Arg Gly Asp Ala Ala Tyr Arg Gln Leu Glu Arg Asp Thr 1860	1865	1870	5615
ttg gct gag gcc aag atg tgg tac acc caa gct ctc aac ttg ctg ggt Leu Ala Glu Ala Lys Met Trp Tyr Thr Gln Ala Leu Asn Leu Leu Gly 1875	1880	1885	5663

gat gag cca caa gtc atg ctc tcc aca acc tgg gcc aac cca acc ttg Asp Glu Pro Gln Val Met Leu Ser Thr Thr Trp Ala Asn Pro Thr Leu 1890 1895 1900	5711
ggc aac gct gcc tcc aag acc aca caa cag gtc agg caa cag gtc ctc Gly Asn Ala Ala Ser Lys Thr Thr Gln Gln Val Arg Gln Gln Val Leu 1905 1910 1915	5759
acc caa ctc agg ctc aac tct aga gtc aag act cca ctc ttg ggc act Thr Gln Leu Arg Leu Asn Ser Arg Val Lys Thr Pro Leu Leu Gly Thr 1920 1925 1930 1935	5807
gcc aac tcc ctc act gct ctc ttc cca caa gag aac tcc aaa ctt Ala Asn Ser Leu Thr Ala Leu Phe Leu Pro Gln Glu Asn Ser Lys Leu 1940 1945 1950	5855
aag ggt tac tgg agg acc ctt gct caa cgc atg ttc aac ctc agg cac Lys Gly Tyr Trp Arg Thr Leu Ala Gln Arg Met Phe Asn Leu Arg His 1955 1960 1965	5903
aac ctc tcc att gat ggt caa cca ctc tcc ttg cca ctc tac gct aag Asn Leu Ser Ile Asp Gly Gln Pro Leu Ser Leu Pro Leu Tyr Ala Lys 1970 1975 1980	5951
cca gct gac cca aag gct ctc ctt tcc gct gct gtc tcc gca tcc caa Pro Ala Asp Pro Lys Ala Leu Leu Ser Ala Ala Val Ser Ala Ser Gln 1985 1990 1995	5999
ggt ggt gct gac ctc cca aag gct cca ctc acc atc cac agg ttc cca Gly Gly Ala Asp Leu Pro Lys Ala Pro Leu Thr Ile His Arg Phe Pro 2000 2005 2010 2015	6047
caa atg ttg gag ggt gcc cgt ggt ctt gtc aac cag ctc atc caa ttc Gln Met Leu Glu Gly Ala Arg Gly Leu Val Asn Gln Leu Ile Gln Phe 2020 2025 2030	6095
ggt tcc tct ctc ctt ggt tac tct gag agg caa gat gct gag gcc atg Gly Ser Ser Leu Leu Gly Tyr Ser Glu Arg Gln Asp Ala Glu Ala Met 2035 2040 2045	6143
tcc caa ctc ttg caa acc cag gct tct gag ttg atc ctc acc tcc atc Ser Gln Leu Leu Gln Ala Ser Glu Leu Ile Leu Thr Ser Ile 2050 2055 2060	6191
agg atg caa gac aac cag ctt gct gag ttg gac tct gag aag act gct Arg Met Gln Asp Asn Gln Leu Ala Glu Leu Asp Ser Glu Lys Thr Ala 2065 2070 2075	6239
ctc caa gtc tcc ctt gct ggt gtc caa cag agg ttc gac agc tac tcc Leu Gln Val Ser Leu Ala Gly Val Gln Gln Arg Phe Asp Ser Tyr Ser 2080 2085 2090 2095	6287
caa ctc tac gag gag aac atc aac gct ggt gag caa agg gct ttg gct Gln Leu Tyr Glu Glu Asn Ile Asn Ala Gly Glu Gln Arg Ala Leu Ala 2100 2105 2110	6335
ctc agg tct gag tct gcc att gag tcc caa ggt gct caa atc tcc cgc Leu Arg Ser Glu Ser Ala Ile Glu Ser Gln Gly Ala Gln Ile Ser Arg 2115 2120 2125	6383

atg gct ggt gct ggc gtg gac atg gct cca aac atc ttc ggt ctt gct	6431
Met Ala Gly Ala Gly Val Asp Met Ala Pro Asn Ile Phe Gly Leu Ala	
2130 2135 2140	
gat ggt ggc atg cac tac ggt gcc att gct tac gcc att gct gat ggc	6479
Asp Gly Gly Met His Tyr Gly Ala Ile Ala Tyr Ala Ile Ala Asp Gly	
2145 2150 2155	
att gag ctt tct gct tct gcc aag atg gtt gat gct gag aag gtg gct	6527
Ile Glu Leu Ser Ala Ser Ala Lys Met Val Asp Ala Glu Lys Val Ala	
2160 2165 2170 2175	
caa tct gaa atc tac cgt cgc aga cgc caa gaa tgg aag atc caa agg	6575
Gln Ser Glu Ile Tyr Arg Arg Arg Gln Glu Trp Lys Ile Gln Arg	
2180 2185 2190	
gac aac gct caa gct gag atc aac cag ctc aac gct caa ctt gag tcc	6623
Asp Asn Ala Gln Ala Glu Ile Asn Gln Leu Asn Ala Gln Leu Glu Ser	
2195 2200 2205	
ctc agc atc agg cgt gag gct gct gag atg cag aag gag tac ctc aag	6671
Leu Ser Ile Arg Arg Glu Ala Ala Glu Met Gln Lys Glu Tyr Leu Lys	
2210 2215 2220	
acc caa cag gct caa gct cag gct caa ctc acc ttc ctc agg tcc aag	6719
Thr Gln Gln Ala Gln Ala Gln Leu Thr Phe Leu Arg Ser Lys	
2225 2230 2235	
ttc tcc aac cag gct ctc tac tcc tgg ctc aga ggc cgc ctc tct ggc	6767
Phe Ser Asn Gln Ala Leu Tyr Ser Trp Leu Arg Gly Arg Leu Ser Gly	
2240 2245 2250 2255	
atc tac ttc caa ttc tac gac ttg gct gtc tcc cgc tgc ctc atg gct	6815
Ile Tyr Phe Gln Phe Tyr Asp Leu Ala Val Ser Arg Cys Leu Met Ala	
2260 2265 2270	
gag caa tcc tac caa tgg gag gcc aac gac aac agc atc tcc ttc gtc	6863
Glu Gln Ser Tyr Gln Trp Glu Ala Asn Asp Asn Ser Ile Ser Phe Val	
2275 2280 2285	
aag cca ggt gct tgg caa ggc acc tac gct ggt ctc ctt tgc ggt gag	6911
Lys Pro Gly Ala Trp Gln Gly Thr Tyr Ala Gly Leu Leu Cys Gly Glu	
2290 2295 2300	
gct ctc atc cag aac ttg gct caa atg gag gag gct tac ctc aag tgg	6959
Ala Leu Ile Gln Asn Leu Ala Gln Met Glu Glu Ala Tyr Leu Lys Trp	
2305 2310 2315	
gag tcc aga gct ttg gag gta gag agg act gtc tcc ctt gct gta gtc	7007
Glu Ser Arg Ala Leu Glu Val Glu Arg Thr Val Ser Leu Ala Val Val	
2320 2325 2330 2335	
tac gac tcc ttg gag ggc aac gac agg ttc aac ctt gct gag caa atc	7055
Tyr Asp Ser Leu Glu Gly Asn Asp Arg Phe Asn Leu Ala Glu Gln Ile	
2340 2345 2350	
cca gct ctc ttg gac aag ggt gag ggc act gct ggc acc aag gag aac	7103
Pro Ala Leu Leu Asp Lys Gly Glu Gly Thr Ala Gly Thr Lys Glu Asn	
2355 2360 2365	
ggt ctc tcc ttg gcc aac gcc atc ctc tct gct tct gtc aag ctc tct	7151

Gly Leu Ser Leu Ala Asn Ala Ile Leu Ser Ala Ser Val Lys Leu Ser				
2370	2375	2380		
gac ctc aag ttg ggt act gac tac cca gac tcc att gtg ggt tcc aac				7199
Asp Leu Lys Leu Gly Thr Asp Tyr Pro Asp Ser Ile Val Gly Ser Asn				
2385	2390	2395		
aag gtc aga agg atc aag caa atc tct gtc tcc ctc cca gct ttg gtg				7247
Lys Val Arg Arg Ile Lys Gln Ile Ser Val Ser Leu Pro Ala Leu Val				
2400	2405	2410	2415	
ggt cca tac caa gat gtc caa gcc atg ctc tcc tac ggt ggc tcc acc				7295
Gly Pro Tyr Gln Asp Val Gln Ala Met Leu Ser Tyr Gly Gly Ser Thr				
2420	2425	2430		
caa ctc cca aag ggt tgc tct gct ttg gct gtc tcc cac ggc acc aac				7343
Gln Leu Pro Lys Gly Cys Ser Ala Leu Ala Val Ser His Gly Thr Asn				
2435	2440	2445		
gac tct ggt caa ttc caa ctt gac ttc aac gat ggc aag tac ctc cca				7391
Asp Ser Gly Gln Phe Leu Asp Phe Asn Asp Gly Lys Tyr Leu Pro				
2450	2455	2460		
ttc gaa ggc att gct ttg gat gac caa ggc acc ctc aac ctc caa ttc				7439
Phe Glu Gly Ile Ala Leu Asp Asp Gln Gly Thr Leu Asn Leu Gln Phe				
2465	2470	2475		
cca aac gcc act gac aag cag aag gcc atc ctc caa acc atg tct gac				7487
Pro Asn Ala Thr Asp Lys Gln Lys Ala Ile Leu Gln Thr Met Ser Asp				
2480	2485	2490	2495	
atc atc ctc cac atc agg tac acc atc agg tgagctcgag aggcctgcgg				7537
Ile Ile Leu His Ile Arg Tyr Thr Ile Arg				
2500	2505			
ccgc				7541
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encoding ER signal from 15 kDa zein from Black				
Mexican Sweet maize				
<220>				
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Met Ala Lys Met Val Ile Val Leu Val Val Cys Leu Ala Leu Ser Ala				
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gcc tgt gct tca gcc				63
Ala Cys Ala Ser Ala				
20				

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 reticulum signal peptide

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Met	Ala	Lys	Met	Val	Ile	Val	Leu	Val	Val	Cys	Leu	Ala	Leu	Ser		
1															15	

gct	gcc	tgt	gct	tca	gcc	atg	aac	gag	tcc	gtc	aag	gag	atc	cca	gac	96
Ala	Ala	Cys	Ala	Ser	Ala	Met	Asn	Glu	Ser	Val	Lys	Glu	Ile	Pro	Asp	
20															30	

gtc	ctc	aag	tcc	caa	tgc	ggt	ttc	aac	tgc	ctc	act	gac	atc	tcc	cac	144
Val	Leu	Lys	Ser	Gln	Cys	Gly	Phe	Asn	Cys	Leu	Thr	Asp	Ile	Ser	His	
35															45	

agc	tcc	ttc	aac	gag	ttc	aga	caa	caa	gtc	tct	gag	cac	ctc	tcc	tgg	192
Ser	Ser	Phe	Asn	Glu	Phe	Arg	Gln	Gln	Val	Ser	Glu	His	Leu	Ser	Trp	
50															60	

tcc	gag	acc	cat	gac	ctc	tac	cat	gac	gct	cag	caa	gct	cag	aag	gac	240
Ser	Glu	Thr	His	Asp	Leu	Tyr	His	Asp	Ala	Gln	Gln	Ala	Gln	Lys	Asp	
65															75	

aac	agg	ctc	tac	gag	gtc	agg	atc	ctc	aag	agg	gct	aac	cca	caa	ctc	288
Asn	Arg	Leu	Tyr	Glu	Ala	Arg	Ile	Leu	Lys	Arg	Ala	Asn	Pro	Gln	Leu	
80															95	

cag	aac	gct	gtc	cac	ctc	gcc	atc	ttg	gct	cca	aac	gct	gag	ttg	att	336
Gln	Asn	Ala	Val	His	Leu	Ala	Ile	Leu	Ala	Pro	Asn	Ala	Glu	Leu	Ile	
100															110	

ggt	tac	aac	aac	cag	ttc	tct	ggc	aga	gct	agc	cag	tac	gtg	gct	cct	384
Gly	Tyr	Asn	Asn	Gln	Phe	Ser	Gly	Arg	Ala	Ser	Gln	Tyr	Val	Ala	Pro	
115															125	

ggt	aca	gtc	tcc	tcc	atg	ttc	agc	cca	gcc	gct	tac	ctc	act	gag	ttg	432
Gly	Thr	Val	Ser	Ser	Met	Phe	Ser	Pro	Ala	Ala	Tyr	Leu	Thr	Glu	Leu	
130															140	

tac	cgc	gag	gct	agg	aac	ctt	cat	gct	tct	gac	tcc	gtc	tac	tac	ttg	480
Tyr	Arg	Glu	Ala	Arg	Asn	Leu	His	Ala	Ser	Asp	Ser	Val	Tyr	Tyr	Leu	
145															155	

gac	aca	cgc	aga	cca	gac	ctc	aag	agc	atg	gcc	ctc	agc	caa	cag	aac	528
Asp	Thr	Arg	Arg	Pro	Asp	Leu	Lys	Ser	Met	Ala	Leu	Ser	Gln	Gln	Asn	
160															175	

atg	gac	att	gag	ttg	tcc	acc	ctc	tcc	ttg	agc	aac	gag	ctt	ctc	ttg	576
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Met Asp Ile Glu Leu Ser Thr Leu Ser Leu Ser Asn Glu Leu Leu			
180	185	190	
gag tcc atc aag act gag agc aag ttg gag aac tac acc aag gtc atg			624
Glu Ser Ile Lys Thr Glu Ser Lys Leu Glu Asn Tyr Thr Lys Val Met			
195	200	205	
gag atg ctc tcc acc ttc aga cca agc ggt gca act cca tac cat gat			672
Glu Met Leu Ser Thr Phe Arg Pro Ser Gly Ala Thr Pro Tyr His Asp			
210	215	220	
gcc tac gag aac gtc agg gag gtc atc caa ctt caa gac cct ggt ctt			720
Ala Tyr Glu Asn Val Arg Glu Val Ile Gln Leu Gln Asp Pro Gly Leu			
225	230	235	
gag caa ctc aac gct tct cca gcc att gct ggt ttg atg cac cag gca			768
Glu Gln Leu Asn Ala Ser Pro Ala Ile Ala Gly Leu Met His Gln Ala			
240	245	250	255
tcc ttg ctc ggt atc aac gcc tcc atc tct cct gag ttg ttc aac atc			816
Ser Leu Leu Gly Ile Asn Ala Ser Ile Ser Pro Glu Leu Phe Asn Ile			
260	265	270	
ttg act gag gag atc act gag ggc aac gct gag gag ttg tac aag aag			864
Leu Thr Glu Glu Ile Thr Glu Gly Asn Ala Glu Glu Leu Tyr Lys Lys			
275	280	285	
aac ttc ggc aac att gag cca gcc tct ctt gca atg cct gag tac ctc			912
Asn Phe Gly Asn Ile Glu Pro Ala Ser Leu Ala Met Pro Glu Tyr Leu			
290	295	300	
aag agg tac tac aac ttg tct gat gag gag ctt tct caa ttc att ggc			960
Lys Arg Tyr Tyr Asn Leu Ser Asp Glu Glu Leu Ser Gln Phe Ile Gly			
305	310	315	
aag gct tcc aac ttc ggt caa cag gag tac agc aac aac cag ctc atc			1008
Lys Ala Ser Asn Phe Gly Gln Gln Glu Tyr Ser Asn Asn Gln Leu Ile			
320	325	330	335
act cca gtt gtg aac tcc tct gat ggc act gtg aag gtc tac cgc atc			1056
Thr Pro Val Val Asn Ser Ser Asp Gly Thr Val Lys Val Tyr Arg Ile			
340	345	350	
aca cgt gag tac acc aca aac gcc tac caa atg gat gtt gag ttg ttc			1104
Thr Arg Glu Tyr Thr Asn Ala Tyr Gln Met Asp Val Glu Leu Phe			
355	360	365	
cca ttc ggt ggt gag aac tac aga ctt gac tac aag ttc aag aac ttc			1152
Pro Phe Gly Gly Glu Asn Tyr Arg Leu Asp Tyr Lys Phe Lys Asn Phe			
370	375	380	
tac aac gcc tcc tac ctc tcc atc aag ttg aac gac aag agg gag ctt			1200
Tyr Asn Ala Ser Tyr Leu Ser Ile Lys Leu Asn Asp Lys Arg Glu Leu			
385	390	395	
gtc agg act gag ggt gct cct caa gtg aac att gag tac tct gcc aac			1248
Val Arg Thr Glu Gly Ala Pro Gln Val Asn Ile Glu Tyr Ser Ala Asn			
400	405	410	415
atc acc ctc aac aca gct gac atc tct caa cca ttc gag att ggt ttg			1296
Ile Thr Leu Asn Thr Ala Asp Ile Ser Gln Pro Phe Glu Ile Gly Leu			

420	425	430	
acc aga gtc ctt ccc tct ggc tcc tgg gcc tac gct gca gcc aag ttc			1344
Thr Arg Val Leu Pro Ser Gly Ser Trp Ala Tyr Ala Ala Lys Phe			
435	440	445	
act gtt gag gag tac aac cag tac tct ttc ctc ttg aag ctc aac aag			1392
Thr Val Glu Glu Tyr Asn Gln Tyr Ser Phe Leu Leu Lys Leu Asn Lys			
450	455	460	
gca att cgt ctc agc aga gcc act gag ttg tct ccc acc atc ttg gag			1440
Ala Ile Arg Leu Ser Arg Ala Thr Glu Leu Ser Pro Thr Ile Leu Glu			
465	470	475	
ggc att gtg agg tct gtc aac ctt caa ctt gac atc aac act gat gtg			1488
Gly Ile Val Arg Ser Val Asn Leu Gln Leu Asp Ile Asn Thr Asp Val			
480	485	490	495
ctt ggc aag gtc ttc ctc acc aag tac tac atg caa cgc tac gcc atc			1536
Leu Gly Lys Val Phe Leu Thr Lys Tyr Tyr Met Gln Arg Tyr Ala Ile			
500	505	510	
cat gct gag act gca ctc atc ctc tgc aac gca ccc atc tct caa cgc			1584
His Ala Glu Thr Ala Leu Ile Leu Cys Asn Ala Pro Ile Ser Gln Arg			
515	520	525	
tcc tac gac aac cag cct tcc cag ttc gac agg ctc ttc aac act cct			1632
Ser Tyr Asp Asn Gln Pro Ser Gln Phe Asp Arg Leu Phe Asn Thr Pro			
530	535	540	
ctc ttg aac ggc cag tac ttc tcc act ggt gat gag gag att gac ctc			1680
Leu Leu Asn Gly Gln Tyr Phe Ser Thr Gly Asp Glu Glu Ile Asp Leu			
545	550	555	
aac tct ggc tcc aca ggt gac tgg aga aag acc atc ttg aag agg gcc			1728
Asn Ser Gly Ser Thr Gly Asp Trp Arg Lys Thr Ile Leu Lys Arg Ala			
560	565	570	575
ttc aac att gat gtc tct ctc ttc cgt ctc ttg aag atc aca gat			1776
Phe Asn Ile Asp Asp Val Ser Leu Phe Arg Leu Leu Lys Ile Thr Asp			
580	585	590	
cac gac aac aag gat ggc aag atc aag aac aac ttg aag aac ctt tcc			1824
His Asp Asn Lys Asp Gly Lys Ile Lys Asn Asn Leu Lys Asn Leu Ser			
595	600	605	
aac ctc tac att ggc aag ttg ctt gca gac atc cac caa ctc acc att			1872
Asn Leu Tyr Ile Gly Lys Leu Leu Ala Asp Ile His Gln Leu Thr Ile			
610	615	620	
gat gag ttg gac ctc ttg ctc att gca gtc ggt gag ggc aag acc aac			1920
Asp Glu Leu Asp Leu Leu Ile Ala Val Gly Glu Gly Lys Thr Asn			
625	630	635	
ctc tct gca atc tct gac aag cag ttg gca acc ctc atc agg aag ttg			1968
Leu Ser Ala Ile Ser Asp Lys Gln Leu Ala Thr Leu Ile Arg Lys Leu			
640	645	650	655
aac acc atc acc tcc tgg ctt cac acc cag aag tgg tct gtc ttc caa			2016
Asn Thr Ile Thr Ser Trp Leu His Thr Gln Lys Trp Ser Val Phe Gln			
660	665	670	

ctc ttc atc atg acc agc acc tcc tac aac aag acc ctc act cct gag	2064
Leu Phe Ile Met Thr Ser Thr Ser Tyr Asn Lys Thr Leu Thr Pro Glu	
675	680
685	
atc aag aac ctc ttg gac aca gtc tac cac ggt ctc caa ggc ttc gac	2112
Ile Lys Asn Leu Leu Asp Thr Val Tyr His Gly Leu Gln Gly Phe Asp	
690	695
700	
aag gac aag gct gac ttg ctt cat gtc atg gct ccc tac att gca gcc	2160
Lys Asp Lys Ala Asp Leu Leu His Val Met Ala Pro Tyr Ile Ala Ala	
705	710
715	
acc ctc caa ctc tcc tct gag aac gtg gct cac tct gtc ttg ctc tgg	2208
Thr Leu Gln Leu Ser Ser Glu Asn Val Ala His Ser Val Leu Leu Trp	
720	725
730	735
gct gac aag ctc caa cct ggt gat ggt gcc atg act gct gag aag ttc	2256
Ala Asp Lys Leu Gln Pro Gly Asp Gly Ala Met Thr Ala Glu Lys Phe	
740	745
750	
tgg gac tgg ctc aac acc aag tac aca cca ggc tcc tct gag gct gtt	2304
Trp Asp Trp Leu Asn Thr Lys Tyr Thr Pro Gly Ser Ser Glu Ala Val	
755	760
765	
gag act caa gag cac att gtg caa tac tgc cag gct ctt gca cag ttg	2352
Glu Thr Gln Glu His Ile Val Gln Tyr Cys Gln Ala Leu Ala Gln Leu	
770	775
780	
gag atg gtc tac cac tcc act ggc atc aac gag aac gct ttc aga ctc	2400
Glu Met Val Tyr His Ser Thr Gly Ile Asn Glu Asn Ala Phe Arg Leu	
785	790
795	
ttc gtc acc aag cct gag atg ttc ggt gct gcc aca ggt gct gca cct	2448
Phe Val Thr Lys Pro Glu Met Phe Gly Ala Ala Thr Gly Ala Ala Pro	
800	805
810	815
gct cat gat gct ctc tcc ctc atc atg ttg acc agg ttc gct gac tgg	2496
Ala His Asp Ala Leu Ser Leu Ile Met Leu Thr Arg Phe Ala Asp Trp	
820	825
830	
gtc aac gct ctt ggt gag aag gct tcc tct gtc ttg gct gcc ttc gag	2544
Val Asn Ala Leu Gly Glu Lys Ala Ser Ser Val Leu Ala Ala Phe Glu	
835	840
845	
gcc aac tcc ctc act gct gag caa ctt gct gat gcc atg aac ctt gat	2592
Ala Asn Ser Leu Thr Ala Glu Gln Leu Ala Asp Ala Met Asn Leu Asp	
850	855
860	
gcc aac ctc ttg ctc caa gct tcc att caa gct cag aac cac caa cac	2640
Ala Asn Leu Leu Gln Ala Ser Ile Gln Ala Gln Asn His Gln His	
865	870
875	
ctc cca cct gtc act cca gag aac gct ttc tcc tgc tgg acc tcc atc	2688
Leu Pro Pro Val Thr Pro Glu Asn Ala Phe Ser Cys Trp Thr Ser Ile	
880	885
890	895
aac acc atc ctc caa tgg gtc aac gtg gct cag caa ctc aac gtg gct	2736
Asn Thr Ile Leu Gln Trp Val Asn Val Ala Gln Gln Leu Asn Val Ala	
900	905
910	

cca caa ggt gtc tct gct ttg gtc ggt ctt gac tac atc cag tcc atg Pro Gln Gly Val Ser Ala Leu Val Gly Leu Asp Tyr Ile Gln Ser Met 915 920 925	2784
aag gag aca cca acc tac gct caa tgg gag aac gca gct ggt gtc ttg Lys Glu Thr Pro Thr Tyr Ala Gln Trp Glu Asn Ala Ala Gly Val Leu 930 935 940	2832
act gct ggt ctc aac tcc caa cag gcc aac acc ctc cat gct ttc ttg Thr Ala Gly Leu Asn Ser Gln Gln Ala Asn Thr Leu His Ala Phe Leu 945 950 955	2880
gat gag tct cgc tct gct gcc ctc acc tac tac atc agg caa gtc Asp Glu Ser Arg Ser Ala Ala Leu Ser Thr Tyr Tyr Ile Arg Gln Val 960 965 970 975	2928
gcc aag gca gct gct gcc atc aag tct cgc gat gac ctc tac caa tac Ala Lys Ala Ala Ala Ile Lys Ser Arg Asp Asp Leu Tyr Gln Tyr 980 985 990	2976
ctc ctc att gac aac cag gtc tct gct gcc atc aag acc acc agg atc Leu Leu Ile Asp Asn Gln Val Ser Ala Ala Ile Lys Thr Thr Arg Ile 995 1000 1005	3024
gct gag gcc atc gct tcc atc caa ctc tac gtc aac cgc gct ctt gag Ala Glu Ala Ile Ala Ser Ile Gln Leu Tyr Val Asn Arg Ala Leu Glu 1010 1015 1020	3072
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tct caa ctt gtc tac tac cca gag aac tac att gac cca acc atg agg Ser Gln Leu Val Tyr Tyr Pro Glu Asn Tyr Ile Asp Pro Thr Met Arg 1060 1065 1070	3216
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agc caa ctc aac gct gac act gtg gag gat gcc ttc atg agc tac ctc Ser Gln Leu Asn Ala Asp Thr Val Glu Asp Ala Phe Met Ser Tyr Leu 1090 1095 1100	3312
acc tcc ttc gag caa gtt gcc aac ctc aag gtc atc tct gct tac cat Thr Ser Phe Glu Gln Val Ala Asn Leu Lys Val Ile Ser Ala Tyr His 1105 1110 1115	3360
gac aac atc aac aac gac caa ggt ctc acc tac ttc att ggt ctc tct Asp Asn Ile Asn Asn Asp Gln Gly Leu Thr Tyr Phe Ile Gly Leu Ser 1120 1125 1130 1135	3408
gag act gat gct ggt gag tac tac tgg aga tcc gtg gac cac agc aag Glu Thr Asp Ala Gly Glu Tyr Tyr Trp Arg Ser Val Asp His Ser Lys 1140 1145 1150	3456
ttc aac gat ggc aag ttc gct gca aac gct tgg tct gag tgg cac aag	3504

Phe Asn Asp Gly Lys Phe Ala Ala Asn Ala Trp Ser Glu Trp His Lys			
1155	1160	1165	
att gac tgc cct atc aac cca tac aag tcc acc atc aga cct gtc atc			3552
Ile Asp Cys Pro Ile Asn Pro Tyr Lys Ser Thr Ile Arg Pro Val Ile			
1170	1175	1180	
tac aag aac cgc ctc tac ttg ctc tgg ctt gag cag aag gag atc acc			3600
Tyr Lys Ser Arg Leu Tyr Leu Leu Trp Leu Glu Gln Lys Glu Ile Thr			
1185	1190	1195	
aag caa act ggc aac tcc aag gat ggt tac caa act gag act gac tac			3648
Lys Gln Thr Gly Asn Ser Lys Asp Gly Tyr Gln Thr Glu Thr Asp Tyr			
1200	1205	1210	1215
cgc tac gag ttg aag ttg gct cac atc cgc tac gat ggt acc tgg aac			3696
Arg Tyr Glu Leu Lys Leu Ala His Ile Arg Tyr Asp Gly Thr Trp Asn			
1220	1225	1230	
act cca atc acc ttc gat gtc aac aag atc agc gag ttg aag ttg			3744
Thr Pro Ile Thr Phe Asp Val Asn Lys Ile Ser Glu Leu Lys Leu			
1235	1240	1245	
gag aag aac cgt gct cct ggt ctc tac tgc gct ggt tac caa ggt gag			3792
Glu Lys Asn Arg Ala Pro Gly Leu Tyr Cys Ala Gly Tyr Gln Gly Glu			
1250	1255	1260	
gac acc ctc ttg gtc atg ttc tac aac cag caa gac acc ctt gac tcc			3840
Asp Thr Leu Leu Val Met Phe Tyr Asn Gln Gln Asp Thr Leu Asp Ser			
1265	1270	1275	
tac aag aac gct tcc atg caa ggt ctc tac atc ttc gct gac atg gct			3888
Tyr Lys Asn Ala Ser Met Gln Gly Leu Tyr Ile Phe Ala Asp Met Ala			
1280	1285	1290	1295
tcc aag gac atg act cca gag caa agc aac gtc tac cgt gac aac tcc			3936
Ser Lys Asp Met Thr Pro Glu Gln Ser Asn Val Tyr Arg Asp Asn Ser			
1300	1305	1310	
tac caa cag ttc gac acc aac gtc agg cgt gtc aac aac aga tac			3984
Tyr Gln Gln Phe Asp Thr Asn Asn Val Arg Arg Val Asn Asn Arg Tyr			
1315	1320	1325	
gct gag gac tac gag atc cca agc tct gtc agc tct cgc aag gac tac			4032
Ala Glu Asp Tyr Glu Ile Pro Ser Ser Val Ser Arg Lys Asp Tyr			
1330	1335	1340	
ggc tgg ggt gac tac tac ctc agc atg gtg tac aac ggt gac atc cca			4080
Gly Trp Gly Asp Tyr Tyr Leu Ser Met Val Tyr Asn Gly Asp Ile Pro			
1345	1350	1355	
acc atc aac tac aag gct gcc tct tcc gac ctc aaa atc tac atc agc			4128
Thr Ile Asn Tyr Lys Ala Ala Ser Ser Asp Leu Lys Ile Tyr Ile Ser			
1360	1365	1370	1375
cca aag ctc agg atc atc cac aac ggc tac gag ggt cag aag agg aac			4176
Pro Lys Leu Arg Ile Ile His Asn Gly Tyr Glu Gly Gln Lys Arg Asn			
1380	1385	1390	
cag tgc aac ttg atg aac aag tac ggc aag ttg ggt gac aag ttc att			4224
Gln Cys Asn Leu Met Asn Lys Tyr Gly Lys Leu Gly Asp Lys Phe Ile			

1395	1400	1405	
gtc tac acc tct ctt ggt gtc aac cca aac aac agc tcc aac aag ctc Val Tyr Thr Ser Leu Gly Val Asn Pro Asn Asn Ser Ser Asn Lys Leu 1410	1415	1420	4272
atg ttc tac cca gtc tac caa tac tct ggc aac acc tct ggt ctc aac Met Phe Tyr Pro Val Tyr Gln Tyr Ser Gly Asn Thr Ser Gly Leu Asn 1425	1430	1435	4320
cag ggt aga ctc ttg ttc cac agg gac acc acc tac cca agc aag gtg Gln Gly Arg Leu Leu Phe His Arg Asp Thr Thr Tyr Pro Ser Lys Val 1440	1445	1450	4368
gag gct tgg att cct ggt gcc aag agg tcc ctc acc aac cag aac gct Glu Ala Trp Ile Pro Gly Ala Lys Arg Ser Leu Thr Asn Gln Asn Ala 1460	1465	1470	4416
gcc att ggt gat gac tac gcc aca gac tcc ctc aac aag cct gat gac Ala Ile Gly Asp Asp Tyr Ala Thr Asp Ser Leu Asn Lys Pro Asp Asp 1475	1480	1485	4464
ctc aag cag tac atc ttc atg act gac tcc aag ggc aca gcc act gat Leu Lys Gln Tyr Ile Phe Met Thr Asp Ser Lys Gly Thr Ala Thr Asp 1490	1495	1500	4512
gtc tct ggt cca gtg gag atc aac act gca atc agc cca gcc aag gtc Val Ser Gly Pro Val Glu Ile Asn Thr Ala Ile Ser Pro Ala Lys Val 1505	1510	1515	4560
caa atc att gtc aag gct ggt ggc aag gag caa acc ttc aca gct gac Gln Ile Ile Val Lys Ala Gly Gly Lys Glu Gln Thr Phe Thr Ala Asp 1520	1525	1530	4608
aag gat gtc tcc atc cag cca agc cca tcc ttc gat gag atg aac tac Lys Asp Val Ser Ile Gln Pro Ser Pro Phe Asp Glu Met Asn Tyr 1540	1545	1550	4656
caa ttc aac gct ctt gag att gat ggt tct ggc ctc aac ttc atc aac Gln Phe Asn Ala Leu Glu Ile Asp Gly Ser Gly Leu Asn Phe Ile Asn 1555	1560	1565	4704
aac tct gct tcc att gat gtc acc ttc act gcc ttc gct gag gat ggc Asn Ser Ala Ser Ile Asp Val Thr Phe Thr Ala Phe Ala Glu Asp Gly 1570	1575	1580	4752
cgc aag ttg ggt tac gag agc ttc tcc atc cca gtc acc ctt aag gtt Arg Lys Leu Gly Tyr Glu Ser Phe Ser Ile Pro Val Thr Leu Lys Val 1585	1590	1595	4800
tcc act gac aac gca ctc acc ctt cat cac aac gag aac ggt gct cag Ser Thr Asp Asn Ala Leu Thr Leu His His Asn Glu Asn Gly Ala Gln 1600	1605	1610	4848
tac atg caa tgg caa agc tac cgc acc agg ttg aac acc ctc ttc gca Tyr Met Gln Trp Gln Ser Tyr Arg Thr Arg Leu Asn Thr Leu Phe Ala 1620	1625	1630	4896
agg caa ctt gtg gcc cgt gcc acc aca ggc att gac acc atc ctc agc Arg Gln Leu Val Ala Arg Ala Thr Thr Gly Ile Asp Thr Ile Leu Ser 1635	1640	1645	4944

atg gag acc cag aac atc caa gag cca cag ttg ggc aag ggt ttc tac	4992
Met Glu Thr Gln Asn Ile Gln Glu Pro Gln Leu Gly Lys Gly Phe Tyr	
1650 1655 1660	
gcc acc ttc gtc atc cca cct tac aac ctc agc act cat ggt gat gag	5040
Ala Thr Phe Val Ile Pro Pro Tyr Asn Leu Ser Thr His Gly Asp Glu	
1665 1670 1675	
agg tgg ttc aag ctc tac atc aag cac gtg gtt gac aac aac tcc cac	5088
Arg Trp Phe Lys Leu Tyr Ile Lys His Val Val Asp Asn Asn Ser His	
1680 1685 1690 1695	
atc atc tac tct ggt caa ctc act gac acc aac atc aac atc acc ctc	5136
Ile Ile Tyr Ser Gly Gln Leu Thr Asp Thr Asn Ile Asn Ile Thr Leu	
1700 1705 1710	
ttc atc cca ctt gac gat gtc cca ctc aac cag gac tac cat gcc aag	5184
Phe Ile Pro Leu Asp Asp Val Pro Leu Asn Gln Asp Tyr His Ala Lys	
1715 1720 1725	
gtc tac atg acc ttc aag aag tct cca tct gat ggc acc tgg tgg ggt	5232
Val Tyr Met Thr Phe Lys Lys Ser Pro Ser Asp Gly Thr Trp Trp Gly	
1730 1735 1740	
cca cac ttc gtc cgt gat gac aag ggc atc gtc acc atc aac cca aag	5280
Pro His Phe Val Arg Asp Asp Lys Gly Ile Val Thr Ile Asn Pro Lys	
1745 1750 1755	
tcc atc ctc acc cac ttc gag tct gtc aac gtt ctc aac aac atc tcc	5328
Ser Ile Leu Thr His Phe Glu Ser Val Asn Val Leu Asn Asn Ile Ser	
1760 1765 1770 1775	
tct gag cca atg gac ttc tct ggt gcc aac tcc ctc tac ttc tgg gag	5376
Ser Glu Pro Met Asp Phe Ser Gly Ala Asn Ser Leu Tyr Phe Trp Glu	
1780 1785 1790	
ttg ttc tac tac aca cca atg ctt gtg gct caa agg ttg ctc cat gag	5424
Leu Phe Tyr Tyr Pro Met Leu Val Ala Gln Arg Leu Leu His Glu	
1795 1800 1805	
cag aac ttc gat gag gcc aac agg tgg ctc aag tac gtc tgg agc cca	5472
Gln Asn Phe Asp Glu Ala Asn Arg Trp Leu Lys Tyr Val Trp Ser Pro	
1810 1815 1820	
tct ggt tac att gtg cat ggt caa atc cag aac tac caa tgg aac gtc	5520
Ser Gly Tyr Ile Val His Gly Gln Ile Gln Asn Tyr Gln Trp Asn Val	
1825 1830 1835	
agg cca ttg ctt gag gac acc tcc tgg aac tct gac cca ctt gac tct	5568
Arg Pro Leu Leu Glu Asp Thr Ser Trp Asn Ser Asp Pro Leu Asp Ser	
1840 1845 1850 1855	
gtg gac cct gat gct gtg gct caa cat gac cca atg cac tac aag gtc	5616
Val Asp Pro Asp Ala Val Ala Gln His Asp Pro Met His Tyr Lys Val	
1860 1865 1870	
tcc acc ttc atg agg acc ttg gac ctc ttg att gcc aga ggt gac cat	5664
Ser Thr Phe Met Arg Thr Leu Asp Leu Leu Ile Ala Arg Gly Asp His	
1875 1880 1885	

gct tac cgc caa ttg gag agg gac acc ctc aac gag gca aag atg tgg Ala Tyr Arg Gln Leu Glu Arg Asp Thr Leu Asn Glu Ala Lys Met Trp 1890 1895 1900	5712
tac atg caa gct ctc cac ctc ttg ggt gac aag cca tac ctc cca ctc Tyr Met Gln Ala Leu His Leu Leu Gly Asp Lys Pro Tyr Leu Pro Leu 1905 1910 1915	5760
agc acc act tgg tcc gac cca agg ttg gac cgt gct gct gac atc acc Ser Thr Thr Trp Ser Asp Pro Arg Leu Asp Arg Ala Ala Asp Ile Thr 1920 1925 1930 1935	5808
act cag aac gct cat gac tct gcc att gtt gct ctc agg cag aac atc Thr Gln Asn Ala His Asp Ser Ala Ile Val Ala Leu Arg Gln Asn Ile 1940 1945 1950	5856
cca act cct gct cca ctc tcc ctc aga tct gct aac acc ctc act gac Pro Thr Pro Ala Pro Leu Ser Leu Arg Ser Ala Asn Thr Leu Thr Asp 1955 1960 1965	5904
ttg ttc ctc cca cag atc aac gag gtc atg atg aac tac tgg caa acc Leu Phe Leu Pro Gln Ile Asn Glu Val Met Met Asn Tyr Trp Gln Thr 1970 1975 1980	5952
ttg gct caa agg gtc tac aac ctc aga cac aac ctc tcc att gat ggt Leu Ala Gln Arg Val Tyr Asn Leu Arg His Asn Leu Ser Ile Asp Gly 1985 1990 1995	6000
caa cca ctc tac ctc cca atc tac gcc aca cca gct gac cca aag gct Gln Pro Leu Tyr Leu Pro Ile Tyr Ala Thr Pro Ala Asp Pro Lys Ala 2000 2005 2010 2015	6048
ctt ctc tct gct gtc gct acc agc caa ggt ggt ggc aag ctc cca Leu Leu Ser Ala Ala Val Ala Thr Ser Gln Gly Gly Gly Lys Leu Pro 2020 2025 2030	6096
gag tcc ttc atg tcc ctc tgg agg ttc cca cac atg ttg gag aac gcc Glu Ser Phe Met Ser Leu Trp Arg Phe Pro His Met Leu Glu Asn Ala 2035 2040 2045	6144
cgt ggc atg gtc tcc caa ctc acc cag ttc ggt tcc acc ctc cag aac Arg Gly Met Val Ser Gln Leu Thr Gln Phe Gly Ser Thr Leu Gln Asn 2050 2055 2060	6192
atc att gag agg caa gat gct gag gct ctc aac gct ttg ctc cag aac Ile Ile Glu Arg Gln Asp Ala Glu Ala Leu Asn Ala Leu Leu Gln Asn 2065 2070 2075	6240
cag gca gct gag ttg atc ctc acc aac ttg tcc atc caa gac aag acc Gln Ala Ala Glu Leu Ile Leu Thr Asn Leu Ser Ile Gln Asp Lys Thr 2080 2085 2090 2095	6288
att gag gag ctt gat gct gag aag aca gtc ctt gag aag agc aag gct Ile Glu Glu Leu Asp Ala Glu Lys Thr Val Leu Glu Lys Ser Lys Ala 2100 2105 2110	6336
ggt gcc caa tct cgc ttc gac tcc tac ggc aag ctc tac gat gag aac Gly Ala Gln Ser Arg Phe Asp Ser Tyr Gly Lys Leu Tyr Asp Glu Asn 2115 2120 2125	6384
atc aac gct ggt gag aac cag gcc atg acc ctc agg gct tcc gca gct	6432

Ile Asn Ala Gly Glu Asn Gln Ala Met Thr Leu Arg Ala Ser Ala Ala			
2130	2135	2140	
ggt ctc acc act gct gtc caa gcc tct cgc ttg gct ggt gca gct gct			6480
Gly Leu Thr Thr Ala Val Gln Ala Ser Arg Leu Ala Gly Ala Ala Ala			
2145	2150	2155	
gac ctc gtt cca aac atc ttc ggt ttc gct ggt ggc tcc aga tgg			6528
Asp Leu Val Pro Asn Ile Phe Gly Phe Ala Gly Gly Ser Arg Trp			
2160	2165	2170	2175
ggt gcc att gct gag gct acc ggt tac gtc atg gag ttc tct gcc aac			6576
Gly Ala Ile Ala Glu Ala Thr Gly Tyr Val Met Glu Phe Ser Ala Asn			
2180	2185	2190	
gtc atg aac act gag gct gac aag atc agc caa tct gag acc tac aga			6624
Val Met Asn Thr Glu Ala Asp Lys Ile Ser Gln Ser Glu Thr Tyr Arg			
2195	2200	2205	
agg cgc cgt caa gag tgg gag atc caa agg aac aac gct gag gca gag			6672
Arg Arg Arg Gln Glu Trp Glu Ile Gln Arg Asn Asn Ala Glu Ala Glu			
2210	2215	2220	
ttg aag caa atc gat gct caa ctc aag tcc ttg gct gtc aga agg gag			6720
Leu Lys Gln Ile Asp Ala Gln Leu Lys Ser Leu Ala Val Arg Arg Glu			
2225	2230	2235	
gct gct gtc ctc cag aag acc tcc ctc aag acc caa cag gag caa acc			6768
Ala Ala Val Leu Gln Lys Thr Ser Leu Lys Thr Gln Gln Glu Gln Thr			
2240	2245	2250	2255
cag tcc cag ttg gct ttc ctc caa agg aag ttc tcc aac cag gct ctc			6816
Gln Ser Gln Leu Ala Phe Leu Gln Arg Lys Phe Ser Asn Gln Ala Leu			
2260	2265	2270	
tac aac tgg ctc aga ggc cgc ttg gct gcc atc tac ttc caa ttc tac			6864
Tyr Asn Trp Leu Arg Gly Arg Leu Ala Ala Ile Tyr Phe Gln Phe Tyr			
2275	2280	2285	
gac ctt gct gtg gcc agg tgc ctc atg gct gag caa gcc tac cgc tgg			6912
Asp Leu Ala Val Ala Arg Cys Leu Met Ala Glu Gln Ala Tyr Arg Trp			
2290	2295	2300	
gag ttg aac gat gac tcc gcc agg ttc atc aag cca ggt gct ttg caa			6960
Glu Leu Asn Asp Asp Ser Ala Arg Phe Ile Lys Pro Gly Ala Trp Gln			
2305	2310	2315	
ggc acc tac gct ggt ctc ctt gct ggt gag acc ctc atg ctc tcc ttg			7008
Gly Thr Tyr Ala Gly Leu Leu Ala Gly Glu Thr Leu Met Leu Ser Leu			
2320	2325	2330	2335
gct caa atg gag gat gct cac ctc aag agg gac aag agg gct ttg gag			7056
Ala Gln Met Glu Asp Ala His Leu Lys Arg Asp Lys Arg Ala Leu Glu			
2340	2345	2350	
gtg gag agg aca gtc tcc ctt gct gag gtc tac gct ggt ctc cca aag			7104
Val Glu Arg Thr Val Ser Leu Ala Glu Val Tyr Ala Gly Leu Pro Lys			
2355	2360	2365	
gac aac ggt cca ttc tcc ctt gct caa gag att gac aag ttg gtc agc			7152
Asp Asn Gly Pro Phe Ser Leu Ala Gln Glu Ile Asp Lys Leu Val Ser			

2370	2375	2380	
caa ggt tct ggt tct gct ggt tct ggt aac aac aac ttg gct ttc ggc Gln Gly Ser Gly Ser Ala Gly Ser Gly Asn Asn Asn Leu Ala Phe Gly 2385 2390 2395			7200
gct ggt act gac acc aag acc tcc ctc caa gcc tct gtc tcc ttc gct Ala Gly Thr Asp Thr Lys Ser Leu Gln Ala Ser Val Ser Phe Ala 2400 2405 2410 2415			7248
gac ctc aag atc agg gag gac tac cca gct tcc ctt ggc aag atc agg Asp Leu Lys Ile Arg Glu Asp Tyr Pro Ala Ser Leu Gly Lys Ile Arg 2420 2425 2430			7296
cgc atc aag caa atc tct gtc acc ctc cca gct ctc ttg ggt cca tac Arg Ile Lys Gln Ile Ser Val Thr Leu Pro Ala Leu Leu Gly Pro Tyr 2435 2440 2445			7344
caa gat gtc caa gca atc ctc tcc tac ggt gac aag gct ggt ttg gcg Gln Asp Val Gln Ala Ile Leu Ser Tyr Gly Asp Lys Ala Gly Leu Ala 2450 2455 2460			7392
aac ggt tgc gag gct ctt gct gtc tct cat ggc atg aac gac tct ggt Asn Gly Cys Glu Ala Leu Ala Val Ser His Gly Met Asn Asp Ser Gly 2465 2470 2475			7440
caa ttc caa ctt gac ttc aac gat ggc aag ttc ctc cca ttc gag ggc Gln Phe Gln Leu Asp Phe Asn Asp Gly Lys Phe Leu Pro Phe Glu Gly 2480 2485 2490 2495			7488
att gcc att gac caa ggc acc ctc acc ctc tcc ttc cca aac gct tcc Ile Ala Ile Asp Gln Gly Thr Leu Thr Leu Ser Phe Pro Asn Ala Ser 2500 2505 2510			7536
atg cca gag aag gga aag caa gcc acc atg ctc aag acc ctc aac gat Met Pro Glu Lys Gly Lys Gln Ala Thr Met Leu Lys Thr Leu Asn Asp 2515 2520 2525			7584
atc atc ctc cac atc agg tac acc atc aag tgagctc Ile Ile Leu His Ile Arg Tyr Thr Ile Lys 2530 2535			7621

## INTERNATIONAL SEARCH REPORT

Internal Application No  
PCT/US 00/22237A. CLASSIFICATION OF SUBJECT MATTER  
IPC 7 C12N9/52 C12N15/82 C07K14/24 C12N15/11

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

STRAND, EPO-Internal, WPI Data, PAJ

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 08932 A (DOW AGROSCIENCES LLC ;WISCONSIN ALUMNI RES FOUND (US)) 5 March 1998 (1998-03-05) cited in the application SEQ ID NO:11 in this document is the unmodified version of SEQ ID NO:3 of the present application. SEQ ID NO:46 corresponds to SEQ ID NO:5. page 16, line 31 -page 19, line 35 -----	1-7
A	WO 97 13402 A (DOWELANCO) 17 April 1997 (1997-04-17) the whole document -----	1-7



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

## \* Special categories of cited documents :

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- \*&\* document member of the same patent family

Date of the actual completion of the international search

1 December 2000

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Name and mailing address of the ISA  
European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl.  
Fax: (+31-70) 340-3016

Authorized officer

Sprinks, M

## INTERNATIONAL SEARCH REPORT

Information on patent family members

Internal application No  
PCT/US 00/22237

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